

Supplemental Figure 1. Analysis of hybrid seed development. (A) Percentage of seeds at defined developmental stages at 6 days after pollination (DAP). **(B)** Cleared seeds at the indicated developmental stages. **(C)** Germination of hybrid seeds derived after reciprocal crosses.



Supplemental Figure 2. Confirmation of 10 selected MEGs and PEGs by Sanger sequencing of RT-PCR products. The Sanger sequences of RT-PCR products of 10 randomly selected MEGs PEGs confirm the next generation sequencing results. The first two MEGs are imprinted only in one direction of the cross.



Supplemental Figure 3. Principal Component Analysis (PCA) scatter plot for different categories of imprinted and non-imprinted genes in C. rubella. (A) PCA using maternal informative allele to total informative allele ratio (M/T) and FPKM data for each gene. *At* NIH: Homologs of imprinted *A. thaliana* genes that have been classified as non-imprinted in *C. rubella*, Control: Non-imprinted control. (B) log expression level (FPKM) of different categories of genes. *At* NIH: Homologs of imprinted *A. thaliana* genes that have been classified as non-imprinted in *C. rubella*, Expression level of all genes in these three categories is not significantly different (Mann-Whitney pairwise test, p-val > 0.10).



Supplemental Figure 4. Cross comparison of imprinted genes in five different species. Gene sets used in the comparison are limited to those genes where homologs have been identified in *A. thaliana* (blastn, e-value 1e-6). All homologs with a maximum e-value of 1e-6 were included in the comparison. Data of imprinted genes in different species are derived from the following sources: *A. thaliana* (Hsieh et al., 2011; Wolff et al., 2011; Gehring et al., 2011; Pignatta et al., 2014), rice (Luo et al., 2011), maize (Waters et al., 2013), and castor bean (Xu et al., 2014).



Supplemental Figure 5. Frequency distribution of methylated regions within flanking regions of imprinted genes compared to control genes. Flanking regions (1 kb) of MEGs, PEGs and endosperm expressed control genes were binned (100 bp) and the methylation level in each sequence context was determined for each bin. Only bins with at least five cytosines were considered. The bin with the highest methylation value for each gene was used to calculate the frequency methylation distribution. Genes in the control group were selected based on the presence of SNPs and detectable expression in the endosperm.



Supplemental Figure 6. Likelihood of genes being located within 500 bp proximity of MEGs, PEGs or all genes of *C. rubella* (*Cr*) and *A. thaliana* (*At*). Whiskers represent one standard deviation. Asterisks mark significantly decreased likelihood (hypergeometric test, p-val < 0.05) of finding genes in the proximity of imprinted genes compared to all genes of the corresponding species.

	Mapped Reads to the		Reads Mapped to
Genomic Library	Reference	Average Coverage	Annotated Genes
Cr A	60,952,946	43.22	33,330,305
Cr B	45,540,269	32.45	24,498,833
Transcriptomic	Mapped Reads to The	Average Reads Mapped	Reads Mapped to
Library	Reference	per Genes	Annotated Genes
Cr Whole Seed RNA			
Control*	65,672,150	845.94	61,978,839
Cr A x Cr B Rep 1	118,995,845	1797.37	101,109,365
Cr B x Cr A Rep 1	119,297,773	1868.99	108,871,275
Cr A x Cr B Rep 2	137,143,848	1668.66	127,591,292
Cr B x Cr A Rep 2	135,108,495	1651.77	125,937,105

Supplemental Table 1. Properties of sequencing libraries used in the experiment.

* from replicate 1 Cr x Cr (Rebernig et al., 2015).

Supplemental Table 2. Enriched GO categories of conserved imprinted genes in *C. rubella* (*Cr*) and *A.thaliana* (*At*)

*: p-values (p-val) have been corrected using Benjamini-Hochberg method.

GO Category	Description		At.
co category		p-val*	p-val*
Conserved MEC			
GO:0045449	P regulation of transcription	0.0094	0.002
GO:0019219	P regulation of nucleobase, nucleoside, nucleotide and nucleic acid metabolism	0.0094	0.002
GO:0031323	P regulation of cellular metabolism	0.0094	0.002
GO:0003700	F transcription factor activity	0.0094	0.002
GO:0006350	P transcription	0.0094	0.002
GO:0019222	P regulation of metabolism	0.0094	0.002
GO:0051244	P regulation of cellular physiological process	0.0102	0.0026
GO:0050794	P regulation of cellular process	0.0102	0.0026
GO:0030528	F transcription regulator activity	0.0102	0.0026
GO:0050791	P regulation of physiological process	0.0102	0.0026
GO:0050789	P regulation of biological process	0.0149	0.0043
Conserved PEG	S		
GO:0008757	F S-adenosylmethionine-dependent methyltransferase activity	0.0406	0.0087
GO:0006259	P DNA metabolism	0.0406	0.0018
GO:0006325	P establishment and/or maintenance of chromatin architecture	0.0406	0.0018
GO:0006323	P DNA packaging	0.0406	0.0018
GO:0007001	P chromosome organization and biogenesis (sensu Eukaryota)	0.0406	0.0018
GO:0051276	P chromosome organization and biogenesis	0.0406	0.0018
GO:0008168	F methyltransferase activity	0.0406	0.0249
GO:0016741	F transferase activity, transferring one-carbon groups	0.0406	0.0249

Supplemental Table 3. Primer sequences used for validation of imprinted genes.

Gene ID	Homolog	SNP Position	Cr A Geno- type	Cr B Geno- type	Forward Primer	Reverse Primer					
MEGs tested for Sanger sequencing validation											
Carubv10001875m	AT4G00220.1	16515343	TT	CC	ATGAGCGGCAGCGGA	GAGACGCAGCCGTAGATAG					
Carubv10006496m	AT4G17870.1	8892373	GG	СС	ATGCCTTCGGAGCTAA	AATTCGTGAGCCTGTGCTCT					
Carubv10014802m	AT2G04240.2	9694159	TT	СС	CAGAAGACCACCAGC	CAAAATAACTCTCTTTCTTGC					
Carubv10024888m	AT2G34880.1	9637481	TT	СС	ATCGAAATTTCAAAGA	ATGTTTTCTCATCGCCCTCTC					
Carubv10016646m	AT3G47950.1	7747627	СС	TT	TATACCTGCAAGTCAG	GACCATGGAGCGTTCTCTGT					
Carubv10001038m	AT4G09650.1	12711790	TT	GG	CAAAACTACTGCATCT	CATGTCGTTGCCATTAGCAG					
Carubv10014524m	AT3G14770.1	5063987	AA	GG	CTGGTCATGATCGAAG	TAAAACCGCGATGAAAGGT					
Carubv10025240m	AT2G23580.2	1179435	AA	СС	AAAGGTTTGTGCTCGT	CTAGAGGGCGATCAGGTGT					
Carubv10007482m	AT5G40430.1	16106240	TT	GG	AAAACGTCGGCTACTC	TCATCATCATCCACGGGTTC					
Carubv10026752m	AT5G61430.1	10979408	GG	СС	AATCGCTTGTTGGGAT	GAGAAGCAGGGCACGTAGA C					
PEGs tested for Sang	ger sequencing va	alidation			0,010						
Carubv10020400m	AT1G70560.1	10073861	CC	AA	CGCTGATTTCACTCTC	ATGATCTAGAGGTCAATGCT					
Carubv10011407m	AT1G17770.1	6158407	TT	GG	ACCA CTCATTTGTTTCGGGT	TC GTCCTGCAATTAGCCTCAGC					
Caruby10022676m	AT2G22270 1	8528271	۸۸		GCTGG						
Carubv10022070111	A12032370.1	0520571	~~	cc	GATCG	G					
Carubv10017919m	AT3G29575.4	3677370	GG	AA	TGAGTAGGACTTGTTC GATGC	CCAGAGGAACCACTTCCAG A					
Carubv10019919m	AT5G39550.1	7693760	TT	AA	TGTTCAGATTCCTTGC GACG	CAGTCAAAGTCACATCAGCC TC					
Carubv10006293m	AT4G31900.2	3284332	СС	GG	TTGCTGAGGCAGAAC ATGAC	CAGCTACAAAGCTCGCTCCT					
Carubv10011676m	AT1G28370.1	9749192	СС	GG	TCAACAACAACAACCT CGGA	TAGTTCTCAGGTGGAGGAG GG					
Carubv10026429m	AT5G57180.3	9331482	СС	TT	GTCTTCCCCAAACCCA ATTAG	TTCCTCGTTGAGATTGCGAT GGATG					
Carubv10010527m	AT1G32540.3	11453189	AA	СС	GTACAAGTGGACAGA GCCAGTTAG	TCACATCAGGAGTCGTGCTC					
Carubv10007586m	AT4G14580.1	10545469	AA	СС	TATACTGCACCGGAAG TGATTG	ATCAGACCGAACCTCAAGA G					

All primer pairs were designed to amplify a single locus.