1 Supplemental Materials

2	A Novel Tetrahydrofolate (THF)-dependent Methyltransferase
3	Catalyzing the Demethylation of Dicamba in Sphingomonas sp. Ndbn-20
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18	Running title: Tetrahydrofolate (THF)-dependent dicamba methyltransferase
19	Keywords: Dicamba; Tetrahydrofolate (THF)-dependent methyltransferase; Dicamba
20	resistance; Sphingomonas sp. Ndbn-20
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FIG S1 Construction route of the recombinant vector pDBN10879. AtUbi10 represents the promoter of the *Arabidopsis thaliana* polyubiquitin10 gene; AtCTP2 represents the chloroplast transit peptide coding sequence of the *Arabidopsis thaliana* EPSP synthase gene; tCaMV35S represents the terminator of the cauliflower mosaic virus 35S gene; prCaMV35S represents the promoter of the cauliflower mosaic virus 35S gene; PAT represents the glufosinate resistance gene; and Dmt represents the optimized *dmt* gene.



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FIG S2 SDS-PAGE analysis of the purified methyltransferases from scaffold 02 (A) and
Dmt (B). The proteins were separated on a 12% SDS-polyacrylamide gel and stained with
Coomassie brilliant blue. (A) Lane M, molecular mass standards; lane 1, purified
His₆-methyltransferase from scaffold 02. (B) Lane M, molecular mass standards; lane 1,
purified His₆-Dmt.



FIG S3 HPLC analysis of the products during dicamba conversion by Dmt 37 (methyltransferase from scaffold 66). A and C, HPLC spectrum of the dicamba 38 conversion without or with Dmt, respectively. The sample was dissolved in methanol, and 39 the mobile phase was a mixture of ultrapure water (58.4%), acetonitrile (31.7%), 40 41 methanol (7.5%) and acetic acid (2.4%). The dotted line indicates dicamba (275 nm), and 42 the solid line indicates metabolite 1 (319 nm). B and D, HPLC spectrum of the THF conversion without or with Dmt, respectively. The sample was dissolved in 0.1 mol/L 43 KH₂PO₄ buffer, the mobile phase was a mixture of 0.05 mol/L of pH 3.0 KH₂PO₄ buffer 44 45 (90%) and acetonitrile (10%), and the detection wavelength was 298 nm.



- 48 FIG S4 DNA elements in the *dmt* gene cluster promoter. The -35 and -10 boxes are
- 49 shown in boxes, and the TSS is shown by an arrowhead.

-35 box CAA**TTGGCA**AGGAGATTTTTCCCGAA**CGTCATGAT**GGGAGGCGATATAAGCCCTTGCC CATGTGGAAACGTTCCCATATTGATATGGCAGGGTCGTCGGGTGGTTCGCCGTCTTTCC CGCTGGCCAATGCGCATCGCCGCCTTGCCCAGCGGGTCAGGGCGCGGCGGCCGCTAGGGTC GGCTTCGTCTTGCCGGAGTGTCGAAAACGCACCGGCCAGATCCAATTTTATATCACGG AGAGAGGT**ATGGTG** Dmt

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FIG S5 Transcriptional analysis of *dmt*, *metF*, *purU* and *dhc* in scaffold 66 of strain Ndbn-20 grown with glucose or dicamba. A: Agarose gel electrophoresis of the RT-PCR products using the 16S rRNA gene as an internal control. B: Relative expression levels of the four genes, as determined by RT-qPCR. The data were derived from three independent measurements, and the error bars indicate standard deviations.



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FIG S6 Effect of temperature (A) and pH (B) on the activity of Dmt. For the temperature 59 test, Dmt was incubated under standard conditions at pH 8.0 and different temperatures 60 (20-45 °C) for 10 min, and the relative activity was calculated by assuming that the 61 activity observed at 30 °C was 100%. For the pH test, Dmt was incubated in the pH range 62 of 5.0 to 10.0 at 30 ℃ for 10 min, three different buffering systems were used: ■ indicated 63 100 mM phosphate buffer (pH 5.0 to 6.5), • indicated 100 mM Tris HCl buffer (pH 6.2 to 64 8.5) and ▲ indicated 100 mM glycine-NaOH buffer (pH 8.2 to 10.0). The relative 65 activity was calculated by assuming that the activity observed at pH 7.0 was 100%. 66



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69 **FIG S7** Alignment of nucleotide sequences of the *dmt* gene and the codon-optimized *dmt*

70 gene. The nucleotides optimized were marked out in red.

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90 CTC ATC GAC ATC CTT CCG CCT TTC ATT CCT CCC GAG TTC ACG AAT TCG CCC GAG GAG CAG CCC CCT TCG CAC GAA CCG GCG GTG ATG TTC des dami o cohenegnimized_dami CTT ATT GAT ATC TTG COG CCT TTC ATC CCC COC GAT TTT AGG AAC TGG AGG GAG GAA CAG COC CCT TGG CAT GAA ACC GTT ATG TTT ISO Conserves ct at ga atc t cgg cct ttc at cc gcc gag tt acg aa tgg g gag ga cag cgc gct tgg ca gaa gc gc gt atg tt Amino acid L I D I L R P F I P A E F T N W R E E Q R A W H E A A V M F Amino acid dent GAC CAG TCG CAT CAC ATG TCA GTG ACC TAC ATG CA GTG ACC TAC ATG CAG GGT CCG GAT GCC AAG GCG ATG CTG AAT TAT CTG TCG CCG TCC ACC TTC TCC AAT contemptinized_dent GAT CAA ACC CAC ATG TCA GTC ACC TAC ATT AG GCG CCG GAC CCA AAA GCG ATG CTC AAC TAT CTG AGT CCA TCG ACC TTC TCC AAT Consensus ga ca ca atg tca gt ace ta cat aag gg ccg gg gc aa gcg atg ct aa tat ctg cc tc acc ttc tce ca at Amino acid D Q S H H M S V T Y I K G P D A K A M L N Y L S P S T F S N 270 Amino acid

 dmt
 CTG TCC ACC GAT CGT GGC AAA CAA TAT TTCGTC GCC AAT CCC GAC GGC AAG CAT ATC GGC GAC TGT GTC CTT AT TAT GGG GAT GAT coordon-optimized dmt

 coden-optimized dmt
 CTC TCT ACT GAT CGC GGC AAG CAG TAC TTTGTC GCC AAT CCT GAT GGG AAA CAC ATT GGC GAC TGC GTG CTG CAT TAC TAT GGA GAT GAC Gorsenses

 coden-optimized dmt
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 coden-optimized dmt
 CTC TCT ACT GAT CGC GGC AAG CAG TAC TTTGTC GCA AAT CCT GAT GGG AAA CAC ATT GGC GAC TGC GTG CTG CAT TAC TAT GGA GAT GAC Gorsenses

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 G codon-optimized dmt Consensus Amino acid CTT ACC CCG GCC GAT GCC CTG TGG CAG GCC CGG GTC COC CGG ACG CTC 1398 CTT ACT CCA GCA GAC GCT TTG TCC CAG GGA ACG GTC COC CCA ACA TTG 1398 codon-optimized_dmt ctt ac cc gc ga gc tg tc cag gg gg gt cgc gc ac L T P A D A L S Q G R V P Q G Consensus Amino acid