TAB3 O-GIcNAcylation promotes metastasis of triple negative breast cancer

Supplementary Materials

SUPPLEMENTARY MATERIALS AND METHODS

Reagents and antibodies

GlcNAcstatin was obtained from GlycoBioChem. Human IL-1 β was from Protech. Click-iTTM O-GlcNAc Enzymatic Labeling System and Click-iTTM Biotin Protein Analysis Detection Kit were obtained from Invitrogen. GlycoProfile β -elimination kit was obtained from Sigma. The antibodies used in this study include: rabbit monoclonal anti-Flag antibody, rabbit polyclonal anti-HA antibody (Sigma), rabbit polyclonal anti-GAPDH antibodies (Santa Cruz), rabbit monoclonal anti-TAB3 antibody, rabbit monoclonal anti-TAK1/phosphorylated-TAK1 antibodies, rabbit monoclonal anti-NF- κ Bp65/phosphorylated-NF- κ Bp65 antibodies, rabbit monoclonal anti-p38/phosphorylated-p38 antibodies mouse monoclonal anti-O-GlcNAcylation CTD110.6 antibody (Cell Signaling Technology). The rabbit polyclonal anti-Thr404 phosphorylated-TAB3 antibodies and rabbit polyclonal anti-Ser408 O-GlcNAcylated-TAB3 antibody were made according to the classic approach.

Cell proliferation assay

Cell proliferation was measured using the MTT assay. Briefly, cells were seeded onto 96-well cell culture cluster plates at a concentration of 10^4 cells/well in volumes of 100 µL, and grown overnight. MTT Reagents were added to the subset of wells, and then incubated for 2 hours at 37°C, the DMSO was added to detergent. Absorbances were quantified using an automated plate reader at 370 nm.



Supplementary Figure S1: Disturbed the TAB3 expression has no effect in TNBC cell proliferation. (A) A representative Western blot image of TAB1, TAB2 and TAB3 expression in 5 human breast cancer cell lines. (B) Equal numbers of indicated stably transfected MAD-MB-468 cells (a) and MAD-MB-231 cells (b) were seed on 96 well plates. MTT assay shown the cell viabilities when TAB3 was knockdown or overexpression. Data were shown as means \pm SEM.



Supplementary Figure S2: TAB3 promoted TNBC cell migration and invasion *in vitro*. (A) Equal numbers of indicated stably transfected MAD-MB-468 cells and MAD-MB-231 cells seeded into six-well tissue culture plates. Wound healing assay was performed to detect cell migration ability. (B) 10⁶ stably transfected MAD-MB-468 cells and MAD-MB-231 cells cultured in matrigel chambers. Transwell assay was performed to detect cell invasion ability.



Supplementary Figure S3: TAB3 involved in OGT promoted TNBC cells migration and invasion. MDA-MB-231 cells were transfected with Flag-OGT and/or TAB3 siRNA vector. Cell migration (A) and invasion (B) were assessed as described.



Supplementary Figure S4: IL-1\beta promotes TAB3 O-GlcNAcylation and TAK1 activation in TNBC cells. (A) The specificity of the antibody which against TAB3 O-GlcNAc S408 was determined using cell lysates prepared from cells transfected with WT TAB3 and S408A TAB3 treated with or without 1 mM GlcNAcstatin. (B) MAD-MB-231 cells were serum starved for 6 h, and then stimulated by IL-1 β for 15 min. Cell lysate were immunoblotted for O-GlcNAcylated-TAB3 (gSer408 TAB3), phosphorylated TAK1and total TAB3 and TAK1. Densitometries for TAB3 OGlcNAcylation and TAK1 phosphorylation were normalized against total TAB3 and TAK1 levels. (C) The TAK1 complexes were pulled down from the cell extracts (1 mg of protein extract) using glutathione-sepharose beads, and TAK1 activity assays were performed as described in the materials and methods section.



Supplementary Figure S5: TAB3 O-GlcNAcylation promoted TNBC cell migration and invasion *in vitro*. (A) Equal numbers of WT and S408A TAB3 stably transfected MAD-MB-231 cells seeded into six-well culture plates in the presence or absence of IL-1β. Wound healing assay was performed to detect cell migration viability. (B) 10⁶ stably transfected and MAD-MB-231 cells cultured in matrigel chambers with or without IL-1β. Transwell assay was performed to detect cell invasion ability.



Supplementary Figure S6: The total O-GlcNAcylation level is correlated with the poorer prognosis in TNBC patients. Kaplan-Meier survival curve of breast cancer patients with low and high total O-GlcNAcylation level.

For cloning the human GST-TAB3	
Forward primer	CCGGGATTCATGGCTTATTCAGAAGAGCATA
Reversed primer	CCGCTCGAGCTACAGTTCAATCTCGAATGTC
For cloning the human Flag-TAB3	
Forward primer	CCGGAATTCGGATGTCCCGCAGCGCGGCGGCC
Reversed primer	CCGCTCGAGACAGTTCAATCTCGAATGTC
For cloning the human HA/Flag-OGT	
Forward primer	CCGGAATTCATGTCCCGCAGCGCGGCGGCC
Reversed primer	CCGCTCGAGACTACAGTTCAATCTCGAATGTC
For cloning the human Flag-S408A TAB3	
Forward primer	TCCCGCAGCGCGGCCGACATG
Reversed primer	TACAGTTCAATCTCGAAGGTCGAG
For cloning the human Flag-T404A TAB3	
Forward primer	GCTTATTCAGAAGAGCATACGAGCG
Reversed primer	CCGAGATCCTCCAGGTCAGTTAAGT

Supplementary Table S1: Primers used in the manuscript