### Recruitment of TBK1 to cytosol-invading Salmonella induces WIPI2dependent antibacterial autophagy

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#### **Appendix Figures**

Figure S1

Figure S2

Figure S3

# Appendix Figure S1. Chemical shift mapping of the interaction between the NDP52 zinc finger and ubiquitin.

**A**) Overlay of the <sup>1</sup>H/<sup>15</sup>N HSQC spectrum of 1mM <sup>15</sup>N labeled NDP52 zinc finger in the absence (black) or presence (red) of 2mM ubiquitin.

**B**) Chemical shifts of NDP52 residue upon addition of ubiquitin.

**C**) Overlay of the <sup>1</sup>H/<sup>15</sup>N HSQC spectrum of 1mM <sup>15</sup>N ubiquitin in the absence (black) of presence (red) of 2mM NDP52 zinc finger

**D**) Chemical shift of ubiquitin residues upon addition of NDP52 zinc finger. Chemical shift changes for amide nitrogen (black) and amide proton (white) resonances are given in Hz. Yellow bars residues of ubiquitin whose peaks are broadened upon addition of NDP52.

# Appendix Figure S2. Chemical shift changes of residues in the NDP52 zinc finger as a function of ubiquitin concentration

**A-B**) Overlay of sections of the<sup>1</sup>H/<sup>15</sup>N HSQC spectra of <sup>15</sup>N labeled NDP52 zinc finger. Peaks correspond to I436 (**A**) and H440 (**B**) at different concentrations of unlabeled ubiquitin.

**C**) Chemical shifts in dependence of ubiquitin concentration. The dissociation constants were obtained by fitting the data to a single site-binding model.

#### Appendix Figure S3. <sup>1</sup>H/<sup>15</sup>N HSQC spectra of the NDP52 zinc finger.

Wild type (black) and D439K (red).





### **Appendix Figure S2**



Appendix Figure S3

