

В

1	MKKVLLTKAQHIQPVVASSSDVLVIKPVGSLRVISISAPHSASTYIQRKS	5
51	FQPVYEDIFKLLMKIPDKALTQRVIDAINESNNNDKVAISNHGKQCSCGV	10
101	QKVDSSTQTDWDTEETHMPGITNNSCEAKGNRHSHMTSSSSEPIKPPVDP	15
151	TKVP <u>KKRGRKR</u> NTCVPQVVKRSAAEMALKEREEKQLTPV <u>ITKRKKRDVTQ</u>	20
201	AQNFESYQITNKTPVRRNSTLSDISFNSDELTRVENFISGNTEDRIIRIM	25
251	ANEFRKSHIMSEEGLLPIHEEILHGDVYGVKRQIFVCCHAKMDINELLTR	30
301	DGEDCLELALTNDTDTEIVSLILDARMMTDHLYENSNTALHLAVINHINI	35
351	ESIRLLLRRIDLNSLLLTNDDGYTVLHLAVRNNQFLVVEAILDSIDEREL	40
301	DGEDCLELALTNDTDTEIVSLILDARMMTDHLYENSNTALHLAVINHINI	35
351	ESIRLLLRRIDLNSLLLTNDDGYTVLHLAVRNNOFLVVEAILDSIDEREL	40
401	GETVYRRTLEAANPNEWDEKGFAKYYDRACERLELNKTLLKNRAHKRDVI	45
451	NASEARGGNPPLFYAVEGEQEHLCYFLLAHLADPDEENLSGHSPKSYHYE	50
501	YARTLRINLKVARVMEKVISILNT	52

Supplemental Figure S1. The characterization of *Charon* locus in *Drosophila melanogaster*. A. Structure of *Charon* locus in *Drosophila* genome based on Flybase [26]. B. Sequence of *Drosophila* Charon protein. NLS sequences are shown in Red. Ankyrin repeats are shown in blue.

Ceratitis	MKKVILTKSSNREFATTMKVTSPDNAKSKIDYSELFRVVLKIPEKDVQLRILDA
Bactrocera CG5118	MEKVIVRRSNRNTAALKVISPDNAKSKSAYAELFRVVLQIPEKDVQDRILAA MKKVLLTKAOHIOPVVASSSDVLVIKPVGSLRVISIS
Lucilia	MITVRDFKQKNDTKTNKEDEVIKQKAAT-VDDSKNKFSVIHVVSATQQTVAMVPFRKCTI
Musca	MPNIKPTTEKAPGACKDISIVVGSPLPLSITTEMAKKSISRATVMVKNEMIGVPTSNAPV
Ceratitis	VNGPTSTGNIVHAICPNCKHTEDNAVI.SGKEAGTOTDELEGANKSDTTVVVPMDVT
Bactrocera	VNGKASNAQIIHAMCPNCKKTFGNSNDPGKEIATQTDIAEIISVSKNKATKIITPPPPQT
CG5118	APHSASTYIQRKSFQPVYEDIFKLLMKIPDKALTQRVIDAINESNNN
Lucilia	LKHVAPTKDKTPRPLPVLAPAVEGTGNGQVYQQRKRMLPEYKELFAALMKLPD
Musca	VVKNETVKVPTPNATVIVKREMKPPIQNDMEQKPSHIYQQRKYMLNEYKELFSVLMRFSD
Ceratitis	OTVSTSSSNGODENTMSPULOKAKUSLSAKONLSTTPONLTUSKDTESSOTPNMTEPOSA
Bactrocera	SQNRSHSSNGANSVEISSTISRRTVIFTTVPVDHAIAQQSSIINAIESIEPMEDQTAQVE
CG5118	DKVAISNHGKQCSCGVQKVDSSTQTDWDTEETHMPGITNNSCEAKGNRHSHMTSSSSEPI
Lucilia	${\tt KEMQNKLLAVINSHHSPREYRTIETQTDPVEIKDLLSKE-TEEISIRETANGSGNETPNS}$
Musca	KEISQKVVRCLNAKYANKETQTDPVEILETSSSK-ELVPPVNTNTTESTPTSPN-
Ceratitis	DATAVTSNLPMKRKRKRKVCLPOVVKRSHAOMALTHMOPRLKSRKVOKSPELYO
Bactrocera	EPTNVPVKKKRKRKVCLPQVVKRSHAQMAHTQLQPKLKARKIDKSMDLGQ
CG5118	KPPVDPTKVPKKRGRKRNTCVPQVVKRSAAEMALKEREEKQLTPVITKRKKRDVTQAQNF
Lucilia	KDSNASEEMVKKRKRKRKVSLPQANKESRAKQLAKMNKPKTSMMNNDAA
Musca	NSRPPTPKKRKRRRKVAEPQANKESRVAQRSLLSEKSVQRKPSSQT
Ceratitie	
Bactrocera	KASDDONDACKRDDISSIDSDIDSDIIKDVIMISI -DEIKSKEDHIKKIMALECHLADKK-IEG KPLNELIKRNRSDSMISLASDILNFDMYSDPPELHKTKEEHIMRLMAAECSDADIR-IDG
CG5118	ESYQITNKTPVRRNSTLSDISFNSDELTRVENFISGNTEDRIIRIMANEFRKSHIMSEEG
Lucilia	${\tt kpsapngqqlnspkrqridsfsisecssdivnifddhqtnvimefnklnepfeng}$
Musca	PEIAQSNNDCPAKRRRSNSVCSNDSTLFNMITDMDLSRKDELYESFIKDCIIADVPLENG
Comptitie	
Bactrocera	LLATHRTIVQNDLIALRRQIFVWKKIKOVDDLNTLLTDEDENCLQLAIAQDCFPKIIDVL
CG5118	LLPIHEEILHGDVYGVKROIFVCCHAKMDINELLTRDGEDCLELALTNDTDTEIVSLI
Lucilia	LLPIQDGVVKNQTYKLSLHIPIWQTYIG-TDLNDILTEEDEDLLQLAVINMCDPSIISLL
Musca	LLPIYDCIVRDQDRTLKTQILIWKEYKR-VDLNELITDDDEDLLQLAINNNRSPPIIKLL
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Ceratitis	ISEGLNANEIDCHSNTCVHLAILNEIENESLRLLMRKIDPQLLLHLNDDGYTPLHMAVRA
CG5118	LNEGLNVNEIDDQSNICVNLAVLNEIDDDSLILLFRKIDLNLLILINDDGITPLIMAVRA I.DARMMTDHI.VENSNTAI.HI.AVINHINIESIRI.I.RRIDI.NSI.I.TNDDGYTVI.HI.AVRN
Lucilia	LEKGLNPNTFDAESNTIVHLAILNDITVKSLEHLMNKIELKLLLTLNDEGYTPLHLAIRQ
Musca	$eq:ltnglrpnsvdcqgnniihlcvlndlvpdsmdhlmshidlkmllelnndgytclqlavrq^{$ $ $ $ $ $ $ $ $ $ $ $ $ $ $ $ $ $ $ $
	: : :*. :**.::*.: .*: *: :*: : ** *::*** *::*:*
Ceratitis	NSYLRAEIMLNTLDERLLGKPIFNRNIRASLSEKEFGKYYEEICKNLEKQCGNVT
Bactrocera	NGYLRAEIILNEIDRRLLGEPSFKRNSKVLTSEAEFHKYYEEACKKLEKNYSNTT
CG5118 Lucilia	NQFLVVEAILDSIDERELGETVYRRTLEAANPNE-WDEKGFAKYYDRACERLELNKTLLK DRFLLAFCMLNILDFDLMKKVFYKROLDFLFTDFKVLKKOFHNYYFKICLRMALDDFTNT
Musca	DNFLLAECILNGVDKRLSGSIFYTRNYNKIESDEKTMKLDFKDYYKKVCODMIVEDNVAS
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Ceratitis	RNAQPKLKKKVLETGDRKSGNTALFFAVENKLEHFVFFLLAHLTDPRILNYCGQDA
Bactrocera	RSAQPKLKKKFLEMGDRKSGNTALFFAIENKLEHFIFFLLAHLTDPRVMNFSGQDA
CG5118	NRAHKRDVINASEARGGNPPLFYAVEGEQEHLCYFLLAHLADPDEENLSGHSP
Musca	angniikokilovtovrsgstalffatenosehitvellahigodovenis
nubca	· *I I I.* *II*II
Ceratitis	KSYYSEFGKMLQLSLRVDSAMENVVSLLR-
Bactrocera	KSYYTEFGKMLQISLKVDNAMESVVTILG-
CG5118 Incilia	KSYHYEYARTLRINLKVARVMEKVISILNT
Musca	KTFFNEFGKSLKLSLDIDSAMERVIQLLS-
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Supplemental Figure S2. Evolutionary conservation for Charon protein in *Insects*. Multiple sequence alignments of Charon family members: Lucilia - *Lucilia cuprina* (KNC31104.1); CG5118 - *Drosophila melanogaster* (CG5118); Musca - *Musca domestica* (101889410); Ceratitis - *Ceratitis capitata* (101454591); Bactrocera - *Bactrocera cucurbitae* (105210874).



Supplemental Figure S3. Charon is a nuclear chromatin associated protein. A. Charon protein labels ~ 400 loci in *Drosophila* chromatin: *Drosophila* larvae expressing Charon-GFP were cultured for 60hrs with *E.coli* bacteria; salivary glands were dissected from third instar larvae, squashed and stained with monoclonal anti-GFP (green) antibodys; DNA was detected using propidium iodide (red). **B.** Charon is not covalently modified with pADPr. Total nuclear protein extracts from animals expressing Charon-GFP wild type (WT) and *parg*^{27.1} mutants were subjected to Western-blot analysis using ant-GFP, anti-pADPr antibodies. Proteins which covalently modified with pADPr demonstrate super-shift on Western-blots in *parg* mutants. Band corresponding to Charon-GFP does not demonstrate this. Anti-Actin antibody was used as a loading control. **C.** Quality control of MNase treated mononucleosomal chromatin for Co-IP and ChIP. Digested chromatin was checked on agarose gel after different time intervals of incubation with MNase.



Supplemental Figure S4. Charon represents a third PARP-1 dependent pathway. A. Charon protein is not required for PARP-1-dependent hnRNP-A1 pathway: Charon-GFP (green) is not colocalized with a key component of PARP-1 dependent RNA-fate-regulating pathway, hnRNP A1 (red) genomewide in steady state conditions (top). Upon heat shock hnRNP A1 remains bound to chromatin, while Charon relocalizes to soluble nucleoplasm. B. *Charon* is not required for PARP-1-depednent *hsp70* gene transcription. Quantitative RT-PCR experiment is shown. The amounts of PARP-1 dependent hsp70 mRNA was detected using specific primers before (-) and after (+) 30 minutes heat shock in wild type (WT), *parp-1* mutants and *Charon* knockdowns. **C.** Expression of endogenous *Drosomycin* and *Diptericine* genes was detected after 20 hours infection in *wild type (WT), Parp-1*^{C03256} mutant, and *siRNA*^{CG5118}- *expressing* third instar larvae using quantitative RT-PCR.