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Figure S1 Generation of the $\Delta r lm A$ mutant, complemented and RlmA::GFP strain. (A) The gene replacement strategy for the *rlmA* gene. The *pyrG* gene was used as a selection marker. The primer names and binding sites are indicated by arrows (see Table S2 for the primer sequences). The deletion cassette was constructed by in vivo recombination in S. cerevisiae. (B) A Southern blot analysis of BamHI- and Xhol-digested genomic DNA with a probe that binds specifically to the rlmA 5'-region indicating the predicted 4.81 and 4.63 Kb bands in the $\Delta rlmA$ mutant for BamHI and XhoI DNA digestion, respectively. (C) The successful complementation of the *rlmA* gene was confirmed in CFW- and CR-resistant monoconidial transformants by PCR using primer sets rlmA 600 ups / pyrG REV and rlmA ST SC 5F / rlmA ORF REV (upper and lower panel). (D) PCR for the validation of the CWI pathway double mutants. See text for the details and primers. (G) Gene replacement strategy for *rlmA::gfp* strain construction. The *rlmA* genomic sequence with no stop codon was cloned in-frame with the green fluorescent protein (GFP) gene in a C-terminal fusion separated by a Gly-Thr-Arg-Gly linker. The *pyrG* gene was also used as a prototrophy marker. S. cerevisiae in vivo recombination assay was performed as previously described. The PCR-amplified rlmA::gfp cassette was transformed into the A. fumigatus wild-type strain. Transformants were carefully tested by PCR with primer set rImA 5F and GFP REV pyrG to confirm the *rlmA* locus replacement.