

## Supplemental Information

### Tailored Immunogens Direct Affinity Maturation toward HIV Neutralizing Antibodies

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**Table S1. Related to Figure 1. Purpose of GT3 mutations.**

<b>Mutation in GT3</b>	<b>Purpose</b>	<b>Mutation in GT6?</b>	<b>Mutation in GT8?</b>
N276D	Eliminate 276 glycan	Y	Y
I277F	Stabilize loop D for VRC01-class binding	Y	Y
T278R	Stabilize loop D; possible VRC01-class contact	Y	Y
F353Y	Improve packing behind loop D	N	Y
N356D	Eliminate glycosylation site introduced by I358T	N	N
I358T	Improve packing on $\beta$ -sheet; not VRC01 contact	Y	Y
N363P	Eliminate 363 glycan in BG505 that is uncommon in other HIV strains	N <sup>#</sup>	Y
T372M	Stabilize CD4-binding loop?	N	N
T373N	Add glycosylation site at position 373	N	Y
T455V	Eliminate partially buried and unsatisfied hydroxyl? Possible contact	N	N
S460Y	Stabilize V5? Possible VRC01-class contact	N	Y
T464N	Eliminate the one V5 glycan in BG505	N*	N*

<sup>#</sup>GT6 lacks a glycan at 363 although it does not use P at that position.

\*All V5 glycans have been removed from GT6 and GT8, by other mutations.

**Table S2. Related to Figure 1. Antigenic profile of GT3 SOSIP and native-like trimers**

Table of monovalent binding affinities as measured by SPR using Fabs as analytes and BG505 SOSIP, BG505 GT3 SOSIP, CD4bs-B SOSIP and CD4bs-C SOSIP as ligands.

Dissociation constant Kd (nM)

	Mat-VRC01	GL-VRC01	Mat-12A12	GL-12A12	Mat-CH31	GL-CH31	Mat-3BNC60	Mat-PGV04	PGT145 <sup>a</sup>	PGT151	PGT121	PGT128	35O22
<b>BG505 SOSIP</b>	122	NB	59	NB	137	NB	62	197	6	25*	29	12	276
<b>BG505 N276D-SOSIP</b> <sup>b</sup>	41	-	9	-	69	-	11	67	-	234	26	12	30
<b>GT3 SOSIP</b>	27*	290	19*	1500	38*	2400*	23*	70*	3.7	39*	40	20	411
<b>CD4bs-B N276D-SOSIP</b> <sup>b</sup>	43	-	12	-	811	-	33	165	27	263	19	11	37
<b>CD4bs-C N276D-SOSIP</b> <sup>b</sup>	170	-	212	-	3200	-	10000	17000	18	214	20	10	25

\* complex kinetics are poor fit to simple 1-to-1 binding model

<sup>a</sup> binding determined using IgG

<sup>b</sup> made in HEK 293S (-GnTI) cells

NB: no binding detected at  $\geq 4\mu\text{M}$

**Table S3. Related to Figure 3. L-CDR1 characteristics for selected VRC01-class bnAbs.**

Antibody	VK/VL gene	L-CDR1 sequence*		L-CDR1 length		LCDR1 deletion?
		germline	mature	germline	mature	
12A12	VK1D-33	QDISNY	QGIGSS	6	6	No
12A21	VK1D-33	QDISNY	QGIGSS	6	6	No
3BNC117	VK1D-33	QDISNY	GY	6	2	Yes: 4aa
3BNC60	VK1D-33	QDISNY	GY	6	2	Yes: 4aa
VRC-CH31	VK1D-33	QDISNY	RGIGKD	6	6	No
NIH45-46	VK3-20	QSVSSSY	QSGS	7	4	Yes: 3aa
PGV19	VL2-14	SDVGGYNY	SDFRGFSS	8	8	No
PGV20	VL2-14	SDVGGYNY	STSV	8	4	Yes: 4aa
PGV04	VK3-20	QSVSSSY	SYGH	7	4	Yes: 3aa
VRC01	VK3-20	QSVSSSY	QYGS	7	4	Yes: 3aa
VRC18	VK3-20 <sup>a</sup>	QSVSSSY	QGILSNQ	7	7	No
VRC03	VK3-20	QSVSSSY	QGGNA	7	5	Yes: 2aa
VRC06	VK3-20	QSVSSSY	QGGNS	7	5	Yes: 2aa
VRC23	VK3-15 <sup>b</sup>	QSVSSN	QGVGSD	6	6	No

\*IMGT convention for kappa or lambda chains: L-CDR1 is the stretch between CXXX and XXW.

<sup>a</sup>Georgiev et al., 2013

<sup>b</sup>Zhou et al., 2015

**Table S4. Related to Figure 5. Primers used for antibody gene amplification.**

Round	Chain	Direction	Sequence
1	Heavy	Forward	GAGGTGCAGCTGCAGGAGTCTGG
1	Heavy	Forward	CAGGTGCAGCTGGTGCAG
1	Heavy	Reverse	AGGGGGCTCTCGCAGGAGACGAGG
1	Heavy	Reverse	GGAAGGTGTGCACACCCTGGAC
1	Heavy	Reverse	GGAAGGTGTGCACACCACTGGAC
1	Heavy	Reverse	GGAAGGTGTGCACACTGCTGGAC
1	Heavy	Reverse	AGACTGTGCGCACACCCTGGAC
1	Heavy	Reverse	GAAAGTTCACGGTGGTTATATCC
1	Kappa	Forward	TGCTGCTGCTCTGGGTTCCAG
1	Kappa	Forward	ATTWTCAGCTTCTGCTAATC
1	Kappa	Forward	TTTTCGCTTTTCTGGATTYAG
1	Kappa	Forward	TCGTGTTKCTSTGGTTGCTG
1	Kappa	Forward	ATGGAATCACAGRCYCWGGT
1	Kappa	Forward	TCTGTTGCTCTGGTTYCCAG
1	Kappa	Forward	CAGTTCCTGGGGCTCTTGTGTTC
1	Kappa	Forward	CTCACTAGCTCTTCTCCTC
1	Kappa	Reverse	GATGGTGGGAAGATGGATAAGTT
1	Lambda	Forward	CAGGCTGTTGTGACTCAG
1	Lambda	Forward	CAACTTGTGCTCACTCAG
1	Lambda	Reverse	GTACCATYTGCTTCCAGKCCACT
2	Heavy	Forward	CATCCTTTTTCTAGTAGCAACTGCAACCGGTGTACATTCCCAGGTGCAGCTGGTGCAGTCTGG
2	Heavy	Forward	CATCCTTTTTCTAGTAGCAACTGCAACCGGTGTACATTCCGAGGTGCAGCTGCAGGAGTCTGG
2	Heavy	Reverse	GGAAGACCGATGGGCCCCTTGGTCGACGCTGAGGAGACGGTGACCGTGG
2	Heavy	Reverse	GGAAGACCGATGGGCCCCTTGGTCGACGCTGAGGAGACTGTGAGAGTGG
2	Heavy	Reverse	GGAAGACCGATGGGCCCCTTGGTCGACGCTGCAGAGACAGTGACCAGAG
2	Heavy	Reverse	GGAAGACCGATGGGCCCCTTGGTCGACGCTGAGGAGACGGTGACTGAGG
2	Heavy	Reverse	GGAAGACCGATGGGCCCCTTGGTCGACGCTGAGGAGACGGTGACCAGGG
2	Kappa	Forward	CATCCTTTTTCTAGTAGCAACTGCAACCGGTGTACATTCCGAYATTGTGMTSACMCARWCTMCA
2	Kappa	Reverse	GAAGACAGATGGTGCAGCCACCGTACGTTTGATTTCAGCTTGGTG
2	Kappa	Reverse	GAAGACAGATGGTGCAGCCACCGTACGTTTATTTCCAGCTTGGTG
2	Kappa	Reverse	GAAGACAGATGGTGCAGCCACCGTACGTTTATTTCCAACCTTGTC
2	Kappa	Reverse	GAAGACAGATGGTGCAGCCACCGTACGTTTCAGCTCCAGCTTGGTG
2	Lambda	Forward	CATCCTTTTTCTAGTAGCAACTGCACCGTTTCTGGGCCAGGCTGTTGTGACTCAG
2	Lambda	Forward	CATCCTTTTTCTAGTAGCAACTGCACCGTTTCTGGGCCAACTTGTGCTCACTCAG
2	Lambda	Reverse	GTTGGCTTGAAGCTCCTCACTCGAGAGGACAGTCACTTGGTTCC
2	Lambda	Reverse	GTTGGCTTGAAGCTCCTCACTCGAGAGGACAGTCACTTGGTTCC
2	Lambda	Reverse	GTTGGCTTGAAGCTCCTCACTCGAGAGGACAGTCAATCTGGTTCC
3	Heavy	Forward	ACACTCTTCCCTACACGACGCTCTTCCGATCTNNNNCCAGGGGAAGACCGATGGGCCCTTGGTCCA
3	Kappa	Forward	ACACTCTTCCCTACACGACGCTCTTCCGATCTNNNNCCAGGGAAGATGAAGACAGATGGTGCAGCCACCGT
3	Lambda	Forward	ACACTCTTCCCTACACGACGCTCTTCCGATCTNNNNGTGGCCTTGTGGCTTGAAGCTCCTCACTCCA
3	All	Reverse	AGACGTGTGCTCTTCCGATCTXXXXXXXXXXGGTCATGTATCATCTTTTTCTAGTAGCAACTG
4	All	Forward	CAAGCAGAAGACGGCATAACGAGATCGGCTTCGGCATTCCTGCTGAAGATXXXXXXXXXXGTGACTGGAGTT CAGACGTGTGCTCTTCCGATC
4	All	Reverse	AATGATACGGCGACCACCGAGATCTACACTCTTCCCTACACGACGCTC

Note: Red Xs in the Round 3 reverse primer indicate the position of well-specific barcodes. Blue Xs in the Round 4 reverse primer indicate the position of plate-specific barcodes.