

**Supplementary Information**

# **Regulation of cell-cell fusion by nanotopography**

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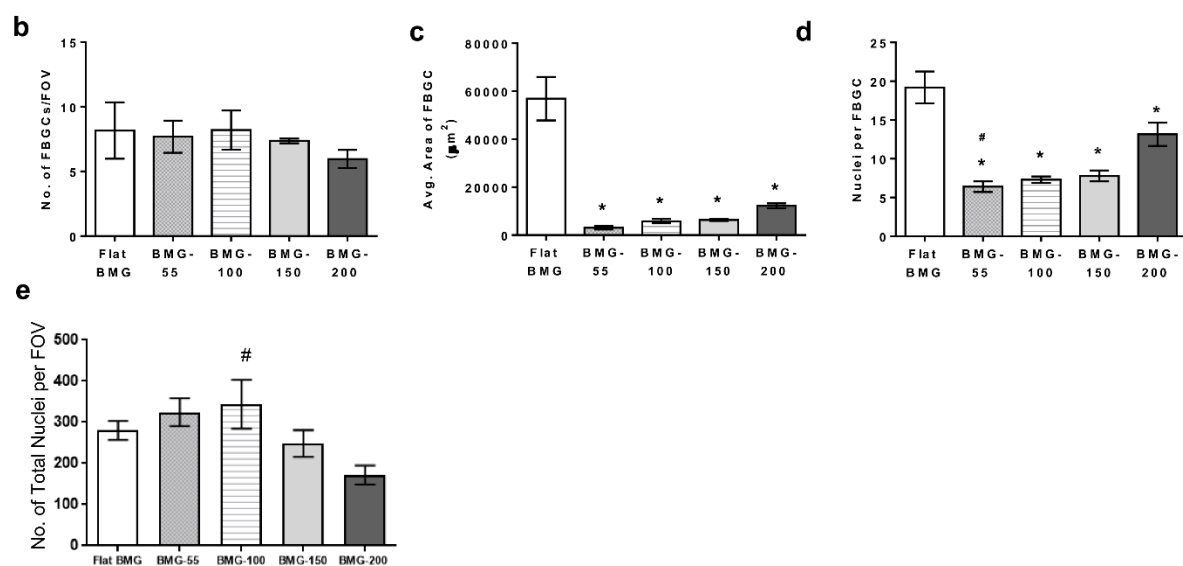
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**a**

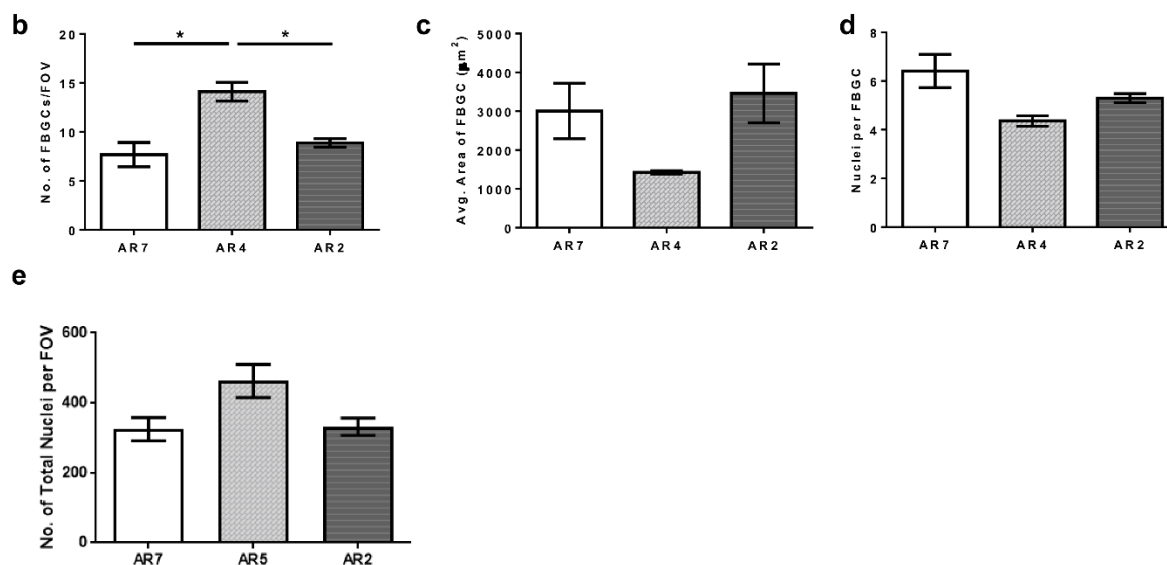
BMG Nanorod Array (nominal rod diameter in nm)	Nanorod Diameter (nm)	Nanorod Height (nm)	Nanorod Aspect Ratio (length:diameter)	Nanorod Stiffness (nN/nm)
Flat BMG	NA	NA	NA	NA
BMG-55	64 ± 10	440 ± 106	6.88	2.75
BMG-100	113 ± 14	1103 ± 146	9.76	1.69
BMG-150	181 ± 18	1134 ± 210	6.27	10.27
BMG-200	247 ± 27	1755 ± 214	7.11	9.61



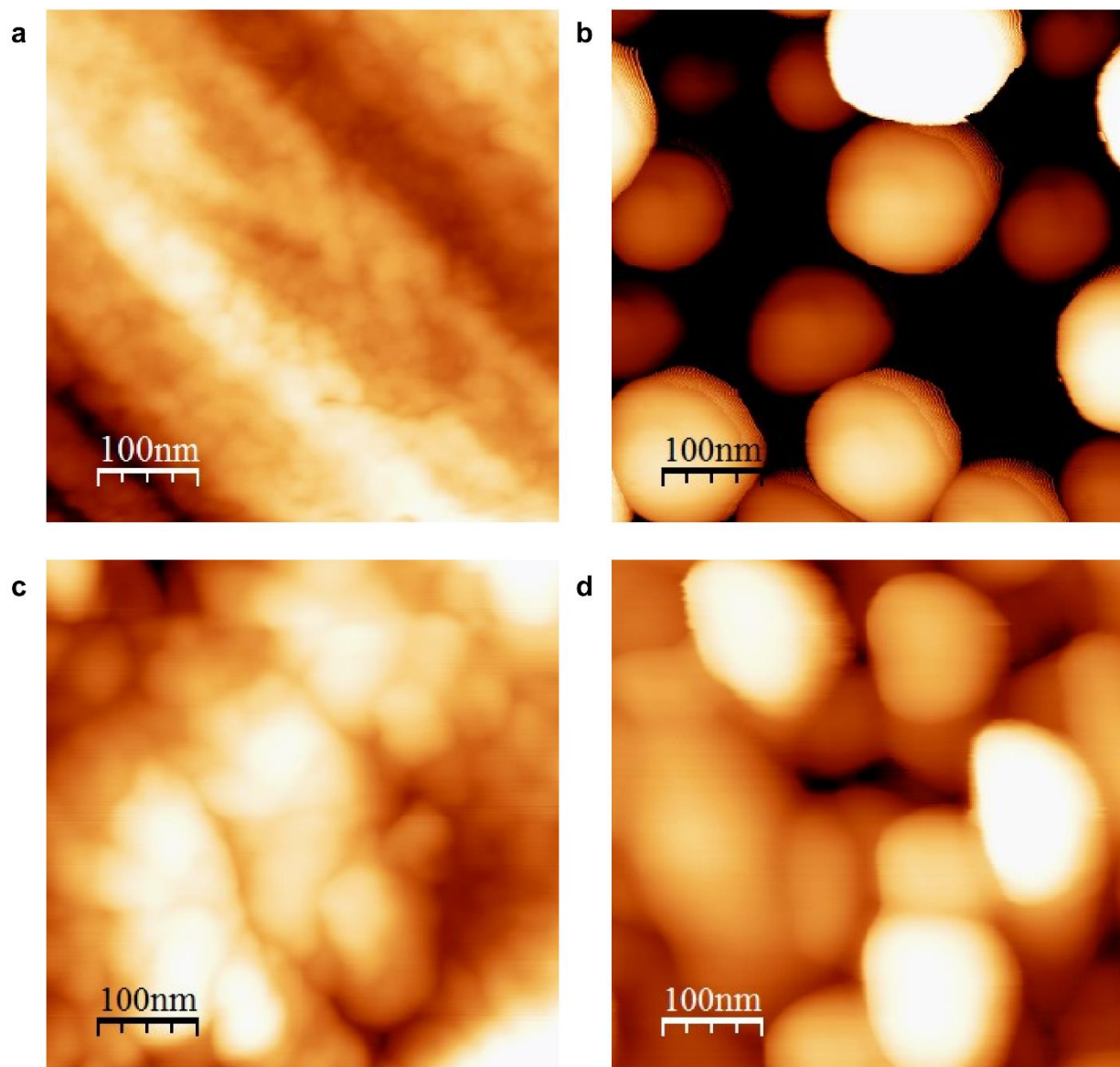
**Fig.S1: Effect of nanotopography on cell fusion** (a) BMG nanorod arrays with varied topography were generated using nanoporous alumina molds as templates. Table showing average nanorod diameter, length and stiffness for each BMG nanorod array. Primary bone-marrow derived murine macrophages were cultured on the nanorod arrays and cell fusion was biochemically induced. Image analysis enabled quantification of (b) Number of FBGCs, (c) FBGC size and (d) nuclei per FBGC. (e) Total nuclei per field of view. Error bars represent standard error mean (SEM) \* represents significant differences as compared to flat BMGs. # represents significant differences as compared to BMG-200. (ANOVA with Tukey's post-hoc analysis,  $n \geq 3$ ,  $p \leq 0.05$  for significance.)

**a**

BMG Nanorod Array	Max. Force applied during TPF (kN)	Nanorod Height (nm)	Nanorod Aspect Ratio (length:diameter)	Nanorod Stiffness (nN/nm)
BMG-55 (AR = 7)	35	440 ± 106	6.88	2.75
BMG-55 (AR = 4)	15	257 ± 50	4.02	13.79
BMG-55 (AR = 2)	5	127 ± 28	1.98	114.34



**Fig. S2: Effect of nanorod stiffness on cell-cell fusion** (a) BMG nanorod arrays with similar topography and varied stiffness were generated using different forming pressures. Nanorod diameters for all BMG-55s were  $64 \pm 10$  nm. Nanorod aspect ratio varied from 7-2. Table showing average nanorod length and stiffness for each BMG nanorod array generated. Primary bone-marrow derived murine macrophages were cultured on the nanorod arrays and cell fusion was biochemically induced. . Image analysis enabled quantification of (b) Number of FBGCs, (c) FBGC size and (d) nuclei per FBGC. (e) Total nuclei per field of view. Error bars represent standard error mean (SEM) \* represents significant differences. (ANOVA with Tukey's post-hoc analysis,  $n \geq 3$ ,  $p \leq 0.05$  for significance.)



**Fig.S3: AFM imaging of BMGs** Flat BMGs (a , c) and BMG-55s (b , d) are shown. Images in c and d show BMGs incubated with serum-containing media prior to AFM whereas a and b were untreated. Scale Bar = 100 nm (a-d). Surface Roughness or RMS values for untreated and treated flat BMGs and the top surface of BMG-55s were found to be within 1-12 nm, much smaller than the nanopattern feature sizes studied here. Percent change in RMS induced by serum protein deposition was 45% for flat BMGs and 177% for BMG-55s.