Supplementary Materials for

Nonsynaptic Communication Through ATP Release from Volume-Activated Anion Channels in Axons

R. Douglas Fields* and Yingchun Ni

*To whom correspondence should be addressed. E-mail: fieldsd@mail.nih.gov

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Fig. S1. Absence of functional gap junction hemichannels in DRG neurons and positive controls.

Table S1. Guide to chloride channels with emphasis on those implicated in release of ATP and small amino acids during cell volume regulation.



Fig. S1. Absence of functional gap junction hemichannels in DRG neurons and positive controls. Astrocytes incubated with 1 mM Lucifer yellow (LY) (Sigma) for ten min at 37° C readily take up the dye (a). Pre-incubation in either carbenoxolone (b) or heptanol (c) (100 μ M, 20 min) inhibits loading of LY into the cells and the intercellular transfer of dye between cells. This positive control shows that the dye enters the cell through gap junction hemichannels and that the drugs are effective in blocking these channels. In contrast, there is no dye loading into DRG neurons, even when 10X higher concentrations of LY and incubation times twice as long as those used to load astrocytes are used. (d) twenty min after incubation in 10 mM LY while neurons are stimulated at 10 Hz. (c) 25 min after washing out LY there is no uptake of the dye into neurons.

Biophysically Identified Chloride Channels	Pharmacological Blockers
Maxi-anion channel	
swelling-induced ATP and glutamate release from	arachidonic acid, DIDS, Gd ³⁺ , NPPB, SITS
non-excitable cells	(not sensitive to glibenclamide, phloretin)
cell volume regulation	
gene identity unknown, VDAC (voltage-dependent anion	
channel) gene disputed as maxi-anion channel	
large-conductance (400 pS), 1.3 nm pore radius	
VRAC (Volume-regulated anion channel)	
also known as VRC, VSOAC, VSOR	DCPIB, DIDS, glibenclamide, NPPB, phloretin, quinine,
cell volume and voltage-dependent ATP conductance	tamoxifen,
cell volume regulation	
efflux of organic osmolytes, including glutamate, from	
non-excitable cells	
gene identity unclear; may not be a single entity	
intermediate conductance (90 pS at positive potentials),	
0.63 nm pore radius	
CFTR (cystic fibrosis transmembrane conductance regulator)	
cAMP regulated Cl ⁻ channel implicated in ATP release	glibenclamide
regulates several conductances, including CaCC, VRAC,	(not sensitive to Gd^{3+})
ORCC, and TRPV4	
0.6-1 nm pore radius	
9 genes with alternative splicing, forming homo and	Cd ²⁺ , DCPIB, DIDS, niflumic acid, NPPB, Zn ²⁺ ,
heterodimeric channels	
Found in intracellular organelles and plasma membrane	
ClC2 activated by cell swelling	
ClC3 relation to VRAC is disputed	
ClC 3/5 is a candidate for VAAC	
ClC3 not sensitive to DIDS and NPPB	
CACC (Calcium-activated chloride channel)	
molecular identity unclear; bestrophins (4 genes) and	DIDS, flufenamic acid (FFA), NPPB, SITS,
anoctamins (10 genes) implicated as candidate CACC	(not sensitive to phloretin)
modulated by CamKII	

Table S1. Guide to chloride channels with emphasis on those implicated in release of ATP and small amino acids during cell volume regulation. Other channels that have not been implicated in volume regulation include ligand-gated chloride channels (GABA, glycine receptors), acid-activated chloride channels (SLC26A7 gene candidate), cGMP-dependent CaCC, intracellular chloride channels (ClCs), ORCC (outwardly rectifying chloride channel), transporter-associated (ClCs), Calcium-activated chloride channel (ClCA family). Abbreviations: DCPIB 4-(2-butyl-6,7-dichlor-2-cyclopentyl-indan-1-on-5-yl); DIDS 4,4'-diisothiocyanostilbene-2,2'-disulphonic acid; FFA flufenamic acid; NPPB 5-nitro-2-(3-phenylpropylamino)benzoic acid; SITS 4'-isothiocyanostilbene-2,2'-disulphonic acid; ORCC outwardly rectifying chloride channel; VACC volume-activated chloride channel; VRAC volume-regulated anion channel (also known as porin); VRC volume-regulated channel; VSOAC volume-activated organic osmolyte-anion channel; VSOR volume-sensitive outwardly rectifying anion channel

Data compiled from information in:

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