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### **Supplementary Note 1.1**





UPLC-MS extracted ion chromatograms of compounds **1**, **2**, **6**, **7**, **10** and their corresponding *N*-acetylcysteamine (SNAC) thioesters, **12–16**, synthesized via EDC/DMAP coupling reactions.

Structure characterization data of compounds 12a and 12b



The chemical structures of two *N*-acetylcysteamine (SNAC) thioester diastereomers (**12a** and **12b**) of compound **1** (precolibactin-413).



Overlay of the <sup>1</sup>H NMR spectra of **12a** and **12b**. The spectra were recorded in DMSO- $d_6$  at 600 MHz. (Maroon = **12a**; Teal = **12b**).



Marfey's analysis to determine the alanine configurations in **12a** and **12b**. The retention times of DNPA (2,4-dinitrophenyl-5-L-alaninamide)-standard L-Ala and DNPA-standard D-Ala were 9.1 min (a) and 9.9 min (b), respectively. Based on the retention times of DNPA-**12a** Ala [9.1 min, (c)] and DNPA-**12b** Ala [9.9 min (d)], the configurations of alanine in **12a** and **12b** were determined as L and D, respectively.



NMR spectroscopi	e data fo	or <b>13</b> in 1	DMSO- $d_6$ .
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С	δ <sub>C</sub>	$\delta_{\rm H}$ ( <i>J</i> in Hz)	COSY	HMBC ( $^{1}$ H to $^{13}$ C)
1	14.1	0.85 t (6.6)	2	2, 3
2	22.6	1.25 m	1	1, 3
3	31.4	1.23-1.25 m		
4-10	28.8, 29.0, 29.1,	1.23-1.25 m		
	29.2, 29.2, 29.2,			
	29.2			
11	28.8	1.23-1.25 m		
12	25.4	1.46 m	13a, 13b	11, 13, 14
13	35.4	a 2.08 m	12, 13b	11, 12, 14
		b 2.08 m	12, 13a	11, 12, 14
14	172.3	-		
		NH, 7.92 d (8.4)	15	14, 15, 16
15	50.0	4.44 m	14NH, 16a,	16, 18
			16b	
16	37.4	a 2.34 dd (8.4, 15.6)	15, 16b	15, 17
		b 2.43 dd (6.0, 15.0)	15, 16a	15, 17
17	171.5	-		
		a NH, 6.83 s	17bNH	16, 17
		b NH, 7.26 s	17aNH	17
18	170.6	-		
		NH, 7.55 d (8.4)	19	18, 19, 20, 21
19	43.8	3.73 m	18NH, 20, 21	18, 20, 21, 22
20	20.5	1.01 d (6.6)	19	19, 21
21	31.6	a 1.63 m	19, 21b, 22a,	19, 20, 22, 23
			22b	
		b 1.67 m	19, 21a, 22a,	19, 20, 22, 23
			22b	
22	40.5	a 2.54 m	21a, 21b, 22b	19, 21, 23
		b 2.57 m	21a, 21b, 22a	19, 21, 23
23	198.5	-		

24	28.1	a 2.87 m	24b, 25a, 25b	23, 25
		b 2.88 m	24a, 25a, 25b	23, 25
25	38.4	a 3.13 m	24a, 24b,	24, 26
			25b, 25NH	
		b 3.16 m	24a, 24b,	24, 26
			25a, 25NH	
		NH, 8.04 t (5.4)	25a, 25b	25, 26
26	169.4	-		
27	22.2	1.78 s		24, 25, 26



The NMR-based key correlations for the structural assignment of 13.



<sup>1</sup>H NMR spectrum of **13** (recorded in DMSO- $d_6$  at 600 MHz).



<sup>13</sup>C NMR spectrum of **13** (recorded in DMSO- $d_6$  at 150 MHz).



<sup>1</sup>H-<sup>13</sup>C HSQC spectrum of **13** (recorded in DMSO- $d_6$  at 600 MHz).



<sup>1</sup>H-<sup>13</sup>C HMBC spectrum of **13** (recorded in DMSO- $d_6$  at 600 MHz).



<sup>1</sup>H-<sup>1</sup>H COSY spectrum of **13** (recorded in DMSO- $d_6$  at 600 MHz).



NMR spectroscopic data for 14 in DMSO- $d_6$ .

С	δ <sub>C</sub>	$\delta_{\rm H} \left( J \text{ in Hz} \right)$	COSY	HMBC ( <sup>1</sup> H to $^{13}$ C)
1	14.1	0.84 t (6.6)	2	2, 3
2	22.2	1.26 m	1	1, 3
3	31.4	1.18-1.26 m		
4-10	28.7, 29.0, 29.1,	1.18-1.26 m		
	29.1, 29.2, 29.2,			
	29.2			
11	28.8	1.18-1.26 m	12	
12	25.3	1.45 m	11, 13a, 13b	
13	35.3	a 2.10 m	12, 13b	11, 12, 14
		b 2.11 m	12, 13a	11, 12, 14
14	172.4	-		
		NH, 7.90 d (7.2)	15	14, 15, 16
15	50.2	4.46 m	14NH, 16a,	16, 18
			16b	
16	37.5	a 2.38 m	15, 16b	15, 17
		b 2.45 m	15, 16a	15, 17
17	171.6	-		
		a NH, 6.83 s	17bNH	16, 17
		b NH, 7.28 s	17aNH	17
18	170.7	-		
		NH, 7.72 d (7.8)	19	18, 19, 20, 21
19	44.7	3.82 m	18NH, 20,	18, 20, 21, 22
			21a, 21b	
20	20.4	1.05 d (6.6)	19	19, 21
21	35.2	a 1.54 m	19, 21b, 22a,	19, 20, 22

			22b	
		b 1.59 m	19, 21a, 22a,	19, 20, 22
			22b	
22	24.1	a 3.03 m	21a, 21b, 22b	
		b 3.25 m	21a, 21b, 22a	
23	153.2	-		
24	109.5	-		
25	166.9	-		
		NH, 8.48 s		23, 24, 25, 26, 29
26	40.1	-		
27	15.3	a 1.33 m	27b, 28a, 28b	26, 28, 29
		b 1.38 m	27a, 28a, 28b	26, 28, 29
28	15.3	a 1.33 m	27a, 27b, 28b	26, 27, 29
		b 1.38 m	27a, 27b, 28a	26, 27, 29
29	159.9	-		
30	103.1	6.10 s		23, 24, 26, 31
31	161.9	-		
32	52.9	a 4.97 d (16.8)		23, 31, 33
		b 4.99 d (16.8)		23, 31, 33
33	194.6	-		
34	28.2	a 2.94 m	34b, 35a, 35b	33, 35
		b 2.97 m	34a, 35a, 35b	33, 35
35	38.1	a 3.16 m	34a, 34b,	34, 36
			35b, 35NH	
		b 3.21 m	34a, 34b,	34, 36
			35a, 35NH	
		NH, 8.06 t (5.4)	35a, 35b	35, 36
36	169.6	-		
37	22.6	1.79 s		36



The NMR-based key correlations for the structural assignment of 14.



<sup>1</sup>H NMR spectrum of **14** (recorded in DMSO- $d_6$  at 600 MHz).



<sup>13</sup>C NMR spectrum of **14** (recorded in DMSO- $d_6$  at 150 MHz).



<sup>1</sup>H-<sup>13</sup>C HSQC spectrum of **14** (recorded in DMSO- $d_6$  at 600 MHz).



<sup>1</sup>H-<sup>13</sup>C HMBC spectrum of **14** (recorded in DMSO- $d_6$  at 600 MHz).



<sup>1</sup>H-<sup>1</sup>H COSY spectrum of **14** (recorded in DMSO- $d_6$  at 600 MHz).



<sup>1</sup>H-<sup>1</sup>H NOESY spectrum of **14** (recorded in DMSO- $d_6$  at 600 MHz).



NMR spectroscopic data for 10 (10a and 10b two stereoisomers) in DMSO- $d_6$ .

С	δ <sub>C</sub>			$\delta_{\rm H} (J \text{ in Hz})$	COSY	HMBC ( $^{1}$ H to $^{13}$ C)
1	14.1			0.84 t (6.8)	2	2, 3
2	22.3			1.25 m	1	1
3	31.5			1.14-1.27 m		
4-10	28.8,	29.0,	29.0,	1.14-1.27 m		
	29.0,	29.1,	29.1,			
	29.2					
11	28.8			1.14-1.27 m		
12	25.4			1.41 m	13	11, 13, 14
13	35.4			2.05 m	12	11, 12, 14
14	172.5 1	l <b>0a</b> /		-		
	172.4 1	lOb				
				NH, 8.03 <b>10a</b> d (7.7)/	15	14, 15, 16
				NH, 8.01 <b>10b</b> d (7.7)		
15	50.2			4.46 m	14NH, 16a,	16, 18
					16b	
16	37.7 10	)a/		a 2.35 dd (8.5, 15.3)	15, 16b	15, 17
	37.8 10	)b				
				b 2.41 dd (4.3, 15.3)	15, 16a	15, 17
17	171.6			-		
				a NH, 6.84 s	17NHb	16, 17
				b NH, 7.31 s	17NHa	17
18	170.6 1	l <b>0a</b> /		-		
	170.5 1	l0b				
				NH, 7.58 <b>10a</b> d (7.7)/	19	18, 19, 20, 21
				NH, 7.66 <b>10b</b> d (7.7)		

19	44.7	3.70 m	18NH, 20, 21a, 21b	18, 20, 21, 22
20	20.4	0.99 d (6.0)	19, 21b	19, 21
21	30.3 <b>10a</b> /	a 1.44 m	19, 21b, 22a,	19, 22
	30.4 <b>10b</b>		22b	
		b 1.57 m	19, 20, 21a, 22a, 22b	19, 22
22	34.9 <b>10a</b> /	a 1.69 <b>10a</b> m/	21a, 21b, 22b	19, 21, 23
	35.0 <b>10b</b>	a 1.79 <b>10b</b> m	, ,	<i>, ,</i>
		b 1.79 <b>10a</b> m/	21a, 21b, 22a	19, 21, 23
		b 1.89 <b>10b</b> m		
23	107.8	-		
24	45.8 <b>10a</b> /	a 2.60 m	24b	22, 23, 25
	46.1 <b>10b</b>			
		b 3.31 d (12.8)	24a	22, 23, 25
25	170.1 <b>10a</b> /	-		
	170.2 <b>10b</b>			
		NH, 7.86 br s		25, 26, 27, 28
26	39.4	-		
27	22.1	a 0.87 m	27b, 28a, 28b	26
		b 1.10 m	27a, 28a, 28b	29
28	22.1	a 0.87 m	27a, 27b, 28b	26
		b 1.10 m	27a, 27b, 28a	29
29	205.4	-		
30	48.4	a 3.10 d (14.5)	30b	26, 29, 31
		b 3.71 m	30a	26, 29, 31
31	166.9	-		
		NH, 9.06 s	32a, 32b	31, 32
32	40.6	a 4.27 d (14.5)	31NH, 32b	33
		b 4.78 dd (6.8, 17.0)	31NH, 32a	33
33	168.1	-		
34	120.1	8.03 s		32, 33, 35, 36, 37
35	154.1	-		
36	107.6	-		
37	160.4	-		
38	156.3	-		
39	127.6	8.17 s		38, 40, 41
40	156.1	-		
41	163.9	-		



NMR-based key correlations for the structural assignment of 10.



HRESIMS of 10.



UV spectrum of **10** (c) in comparison with those of **5** (a) and **7** (b).



MS<sup>n</sup> fragmentation pattern of **10**.







Frag. 1

Frag. 2









Frag. 4



Frag. 6



ΗÓ H₂Ņ ΝH



Frag. 8

Frag. 7

Frag. 7





Frag. 9

Frag. 10



Frag. 11

Frag. 12

$H$ $S$ $H_2N$ $N$ $N$	$\nabla$ $H$ $S$ $H_2N$ $O$	о 9	Fragmentation	MS <sup>n</sup>	Obs. mass	Calc. mass	Error [Da]
H <sub>2</sub> N T N T S OH	Y Y N Y Y OH	H <sub>2</sub> N	1	MS <sup>2</sup>	870.371	870.353	0.018
0 0 0 0	0 0 0 3-	NH	2	MS <sup>2</sup>	852.369	852.342	0.027
		0 111	3	MS <sup>2</sup>	660.183	660,155	0.028
Erag 13	Eraa 14	Eraa 17	4	MS <sup>2</sup>	642.193	642.144	0.049
Flay. 15	Flay. 14	Flag. 17	5	MS <sup>2</sup>	591.416	591.376	0.040
			6	MS <sup>2</sup>	574.381	574.349	0.032
			7	MS <sup>2</sup>	545.156	545.128	0.028
			8	MS <sup>2</sup> /MS <sup>3</sup>	528.141	528.101	0.040
			9	MS <sup>3</sup>	527.136	527.117	0.019
sH_N	S NHa	0	10	MS <sup>2</sup> /MS <sup>3</sup>	510.152	510.091	0.061
$\nabla$ $H$ $3$ $N$		HN	11	MS <sup>3</sup>	483.156	483.127	0.029
$H_2N$ $\Upsilon$ $\Upsilon$ $N$ $\Upsilon$ $\Upsilon$	H <sub>2</sub> N N T T		12	MS <sup>2</sup>	449.383	449.302	0.081
0 0 0 0	O S V OH	-NH	13	MS <sup>2</sup> /MS <sup>3</sup>	422.124	422.059	0.065
		0	14	MS <sup>2</sup> /MS <sup>3</sup> /MS <sup>4</sup>	405.074	405.033	0.041
Frag 15	Frag 16	Frag 18	15	MS⁴	378.075	378.070	0.006
11ay. 15	11ag. 10	1 ag. 10	16	MS <sup>2</sup> /MS <sup>3</sup> /MS <sup>4</sup>	297.081	297.012	0.069
			17	MS <sup>3</sup>	249.214	249.124	0.090
			18	MS <sup>3</sup>	231.116	231.113	0.003

The major fragmentation species from  $MS^n$  measurement of **10**. Fragmentation was acquired with collision energy of 28 V. Obs. = observed; Calc. = calculated.



<sup>1</sup>H NMR spectrum of **10** (recorded in DMSO- $d_6$  at 850 MHz).



<sup>13</sup>C NMR spectrum of **10** (recorded in DMSO- $d_6$  at 212.5 MHz).



<sup>1</sup>H-<sup>13</sup>C HSQC spectrum of **10** (recorded in DMSO- $d_6$  at 850 MHz).



<sup>1</sup>H-<sup>13</sup>C HMBC spectrum of **10** (recorded in DMSO- $d_6$  at 850 MHz).



<sup>1</sup>H-<sup>1</sup>H COSY spectrum of **10** (recorded in DMSO- $d_6$  at 850 MHz).



<sup>1</sup>H-<sup>1</sup>H TOCSY spectrum of **10** (recorded in DMSO- $d_6$  at 850 MHz).



<sup>1</sup>H-<sup>1</sup>H NOESY spectrum of **10** (recorded in DMSO- $d_6$  at 850 MHz).



<sup>1</sup>H-<sup>15</sup>N HSQC spectrum of **10** (recorded in DMSO- $d_6$  at 850 MHz).



Expanded <sup>1</sup>H-<sup>13</sup>C HMBC spectrum of **10** showing the key correlations of H-34 to C-32, C-33, C-35, C-36 and C-37 (recorded in DMSO- $d_6$  at 600 MHz with a 1.7 mm cryoprobe).



Expanded <sup>1</sup>H-<sup>13</sup>C HMBC spectrum of **10** showing the key correlations of H-39 to C-38, C-40 and C-41 (recorded in DMSO- $d_6$  at 850 MHz).



NMR spectroscopic data for 6 in DMSO- $d_6$ .

С	δ <sub>C</sub>			$\delta_{\rm H} \left( J \text{ in Hz} \right)$	COSY	HMBC ( <sup>1</sup> H to <sup>13</sup> C)
1	14.1			0.84 t (7.2)	2	2, 3
2	22.3			1.26 m	1	1, 3
3	31.5			1.19-1.24 m		
4-10	28.9,	29.1,	29.2,	1.19-1.24 m		
	29.2,	29.2,	29.2,			
	29.2					
11	28.8			1.19-1.24 m	12	
12	25.3			1.46 m	11, 13a, 13b	11, 13, 14
13	35.5			a 2.11 m	12, 13b	11, 12, 14
				b 2.17 m	12, 13a	11, 12, 14
14	172.6			-		
				NH, 8.13 br s	15	14
15	50.2			4.50 m	14NH, 16a,	16, 18
					16b	
16	37.7			a 2.38 m	15, 16b	15, 17
				b 2.47 m	15, 16a	15, 17
17	171.8			-		
				a NH, 6.80 s	17bNH	16, 17
				b NH, 7.37 s	17aNH	17
18	170.9			-		
				NH, 7.62 d (7.8)	19	15, 18, 19, 20, 21
19	44.7			3.82 m	18NH, 20,	18, 20, 21, 22
					21a, 21b	
20	20.6			1.03 d (6.6)	19	19, 21, 22
21	35.0			a 1.56 m	19, 21b, 22a,	19, 20, 22
					22b	-
				b 1.64 m	19, 21a, 22a,	19, 20, 22
					22b	

22	24.3	a 2.92 m	21a, 21b, 22b	
		b 3.26 m	21a, 21b, 22a	
23	153.3	-		
24	108.9	-		
25	167.3	-		
		NH, 8.37 s		23, 24, 25, 26, 29,
				30
26	40.1	-		
27	15.2	a 1.30 m	27b, 28a, 28b	26, 28, 29
		b 1.36 m	27a, 28a, 28b	26, 28, 29
28	15.2	a 1.30 m	27a, 27b, 28b	26, 27, 29
		b 1.36 m	27a, 27b, 28a	26, 27, 29
29	159.3	-		
30	102.8	6.03 s		23, 24, 26, 29, 31
31	162.2	-		
32	46.0	a 4.56 d (18.0)		23, 31, 33
		b 4.67 d (16.8)		23, 31, 33
33	169.7	-		



UV spectrum of **6**.



NMR-based key correlations for the structural assignment of 6 as compared to the structure of known 7 (precolibactin-712).

Note for structure elucidation of 6: Compound 6 was obtained as white and amorphous powder. The molecular formula was determined as  $C_{33}H_{51}N_5O_7$  based on the HRESIMS analysis (m/z 630.3842  $[M + H]^+$ , calcd 630.3867). A comparison of the 1D and 2D NMR spectra of 6 and 7 indicated that 6 contains the *N*-myristoyl-D-asparagine residue same and 1H-pyrrolo[3,4-c]pyridine-3,6(2H,5H)-dione unit as in 7. However, the thiazole proton resonance at  $\delta_{\rm H}$  8.09 (H-34), and the carbon signals at  $\delta_{\rm C}$  164.2 (C-33),  $\delta_{\rm C}$  126.9 (C-34) and  $\delta_{\rm C}$  151.7 (C-35) in 7 were not observed in 6, suggesting an absence of the thiazole ring in 6. Instead, C-32 ( $\delta_{\rm C}$  46.0) in 6 was observed to connect the 1H-pyrrolo[3,4-c]pyridine-3,6(2H,5H)-dione unit and the terminal carboxyl group, as evidenced by the HMBC correlations from the methylene protons at  $\delta_{\rm H}$  4.56 (H-32) to the carbon signals at C-23 ( $\delta_{\rm C}$  153.3), C-31 ( $\delta_{\rm C}$  162.2) and C-33 ( $\delta_{\rm C}$  169.7). As an intermediate from the *clb* pathway, **6** can be envisioned to be released from the first NRPS module of ClbJ, which would fill the gap between previously isolated precolibactin-546 (5) and precolibactin-712 (7).



<sup>1</sup>H NMR spectrum of **6** (recorded in DMSO- $d_6$  at 600 MHz).



<sup>13</sup>C NMR spectrum of **6** (recorded in DMSO- $d_6$  at 150 MHz).



<sup>1</sup>H-<sup>13</sup>C HSQC spectrum of **6** (recorded in DMSO- $d_6$  at 600 MHz).



<sup>1</sup>H-<sup>13</sup>C HMBC spectrum of **6** (recorded in DMSO- $d_6$  at 600 MHz).



<sup>1</sup>H-<sup>1</sup>H COSY spectrum of **6** (recorded in DMSO- $d_6$  at 600 MHz).



<sup>1</sup>H-<sup>1</sup>H NOESY spectrum of **6** (recorded in DMSO- $d_6$  at 600 MHz).