

Supplementary Figure 1. Food hardness and food palatability. (a) Food hardness depending on agarose concentration. n=6. (b) Mean food intake of 10 mM and 100 mM sucrose-containing food depending on its hardness. n=6. All data are presented as means  $\pm$  S.E.M. The ANOVA was used to determine statistical significance.



Supplementary Figure 2. Generation of the *nan*<sup>GAL4</sup> allele and labellar mechanosensory neuron *nan* expression. (a) Structure of the *nan* locus and homologous recombination targeting scheme used to generate the  $nan^{GAL4}$  allele (left). Blue boxes indicate exons. Red triangles indicate the primers used for genomic DNA PCR. Deletion of *nan* confirmed by genomic DNA PCR (right). (b) Expression of *nan* in the labellum as measured by RT-PCR on labella from control and mechanosensory neuron-ablated flies. *rp49* was used as an internal control. (c) Confocal images of labella expressing mCD8::GFP driven by *F-GAL4*. Labella were stained with a rabbit anti-GFP. GFP fluorescence was superimposed on a transmitted light image. Scale bar represents 50  $\mu$ m. (d) Food preference of *F-GAL4* neuron-inactivated

flies. Flies were given a choice between 0.5 mM sucrose in 0.2% agarose and 1 mM sucrose in 2 % agarose. Flies were raised at 21°C and shifted to the indicated temperature for 3 days before assaying. n is indicated in parentheses. The unpaired *t*-test was used to determine statistical significance. All data are presented as means  $\pm$  S.E.M.



Supplementary Figure 3. Functional interaction between mechanosensory neurons and sweet GRNs in the SEZ. (a) Proboscis extension response (PER) upon activation of mechanosensory neurons. The PER assay was performed at the indicated temperatures. ANOVAs with Tukey *post-hoc* tests.  ${}^{*}P < 0.05$ ,  ${}^{**}P < 0.01$ . (**b,c**) Effects of mechanosensory neurons and Nan on pharyngeal pumping. (b) Pumping frequencies upon activation of mechanosensory neurons. Black and white bars indicate the pumping frequencies (Hz) of the indicated genotypes at 21°C and 31°C, respectively. The unpaired *t*-test was used to determine the statistical significance of changes in frequency related to temperature. \*P < 0.05, \*\*P < 0.050.01. The ANOVA revealed no significant changes with respect to genotype. N.S. means no significance. (c) Pumping frequencies of wild-type and nan<sup>36a</sup>. The unpaired t-test was used to determine statistical significance. (d) Structural proximity of mechanosensory neurons and sweet GRNs in the subesophageal zone (SEZ). GRASP images in the SEZ (above) and GRASP schematics (below). GRASP between mechanosensory neurons labeled with R41E11-GAL4 and sweet GRNs labeled with Gr5a-LexA. The genotype of the GRASP flies  $Gr5a-LexA/+;LexAOp-CD4::spGFP_{11}/+;UAS-CD4::spGFP_{1-10}/R41E11-GAL4.$ was nc82 neuropil staining (magenta). Scale bar represents 50 µm. (e) Mechanosensory neurons inhibit calcium responses in the axon termini of sweet GRNs. Time course of calcium responses in the axon termini of sweet GRNs in the SEZ shown as average  $\Delta F/F_0$  traces  $\pm$  S.E.M. The 1<sup>st</sup> and 3<sup>rd</sup> trials were conducted at 21°C and 2<sup>nd</sup> trial was conducted at 31°C to activate mechanosensory neurons via dTrpA1. All data are presented as means  $\pm$  S.E.M.



Supplementary Figure 4. Labellar mechanosensory neurons are GABAergic. (a) PER of flies upon knockdown of cholinergic components in mechanosensory neurons. The PER assay was conducted at the indicated temperatures. n=6. ANOVAs with Tukey post-hoc tests. (b-d) Knockdown efficiency and specificity of Gadl and Vgat RNAi in labellar mechanosensory neurons. Relative expression of Gad1 (b), Vgat (c), and nan (d) transcripts upon knockdown of Gad1 and Vgat. Each C<sub>T</sub> value is normalized to the C<sub>T</sub> value for the rp49 internal control. n=3 biological replicates with technical duplicates. Unpaired Student's *t*-tests (b) or ANOVAs with Tukey post-hoc tests (c,d). (e,f) Representative confocal images of *Vgat-GAL4*>mCD8::GFP labella double-stained with a mouse GFP antibody (green) and a rabbit NOMPA antibody (magenta) to visualize the sensillar base. (f) High magnification view of the white boxed area in (e). White arrowheads indicate sensilla base marked with NOMPA antibody. Scale bar represents 50 µm. (g,h) Food preference between 0.5 mM and 1 mM sucrose either in (g) 2% agarose or (h) 0.2% agarose. The red dashed line at 0.5 represents no preference for either of the two food sources. n=5. ANOVA., and All the data are presented as means  $\pm$  S.E.M. \*P < 0.05, \*\*P < 0.01, and N.S. indicates not statistical significance.

# **Supplementary Table 1**

	Number of	Number of GFP positive neurons		
	mechanosensory			D55D01 ( 14)
	neurons	<i>R41E11</i> (n=13)	V12092 (n=8)	<i>K33B01</i> (n=14)
labellum	63	58.85±1.02	60.86±1.14	26.07±3.10
taste bristle	31	27.62±1.03	29.13±1.29	22.21±2.47
lateral	2	2+0.00	2+0.00	2+0.00
mechanosensilla	2	2_0.00	2_0.00	2_0.00
taste peg	30	29.23±0.97	29.75±1.01	1.86±0.70

Supplementary Table 1. Cells labeled by various mechanosensory *GAL4* drivers Average GFP positive neurons labeled by each mechanosensory neuron driver per half of the labellum. *UAS*-mCD8::*GFP* was used as a reporter. Total mechanosensory cells per taste bristle, labellar mechanosensillum, and taste peg are also indicated. Data are means  $\pm$  S.E.M.