

APPENDIX S3

MeSH over-representation analysis (Horse)

This is a reproducible report written by an R Markdown.

1. Obtain a list of genes

```
dat <- read.table("quarter.txt", header=FALSE, stringsAsFactors=FALSE)

require(biomaRt)
mart <- useMart(biomart = "ensembl", dataset = "ecaballus_gene_ensembl")
## gene annotation
allgenes <- getBM(c("ensembl_gene_id", "entrezgene", "hgnc_id", "start_position", "end_position",
                   "chromosome_name"), mart = mart)

chr.name<-numeric()
bp.loc <- numeric()
ensembl.id<-numeric()
entrez.id<-numeric()

m<-1
for(k in 1:nrow(dat))
{
  sub.gene <- subset(allgenes, chromosome_name == dat[k,1])
  for(j in 1:nrow(sub.gene))
  {
    if((sub.gene$start_position[j] - 250000) <= (dat[k,2]) &
      (sub.gene$end_position[j] + 250000) >= (dat[k,2]))
    {
      chr.name[m]<-dat[k,1]
      bp.loc[m]<-dat[k,2]
      ensembl.id[m]<-sub.gene$ensembl_gene_id[j]
      entrez.id[m]<-sub.gene$entrezgene[j]
      m<-m+1
    }
  }
}
```

2. Create a vector of background (universe) genes

We first create a vector of all genes.

```
## remove genes with no corresponding Entrez Gene ID
allgenes2 <- allgenes[!is.na(allgenes$entrezgene),] # 17175
## remove duplicated Entrez Gene ID
allgenes3 <- allgenes2[!duplicated(allgenes2$entrezgene),] # 16808
```

3. Create a vector of significant genes

Secondly, we create a vector of significant genes.

```
dat2 <- data.frame(chr.name, bp.loc, ensembl.id, entrez.id) # 179
## remove genes with no corresponding Entrez Gene ID
dat3 <- dat2[!is.na(dat2$entrez.id),] # 113
```

4. MeSH enrichment analysis

Then, we perform a MeSH ORA for the category **Chemicals and Drugs** by setting ‘category=“D”’.

```
library(meshr)
library(MeSH.db)
library("org.MeSH.Eqc.db")
meshParams <- new("MeSHHyperGParams", geneIds = as.integer(dat3$entrez.id),
                 universeGeneIds = allgenes3$entrezgene, annotation = "org.MeSH.Eqc.db",
                 category = "D", database = "gene2pubmed", pvalueCutoff = 0.05, pAdjust = "none")
meshR <- meshHyperGTest(meshParams)
summary(meshR)[!duplicated(summary(meshR)[,7]),c(1,2,7)]
```

##	MESHID	Pvalue	MESHTERM
## 68	D017392	0.006722989	Thiobarbituric Acid Reactive Substances
## 79	D055435	0.006722989	Myostatin
## 1	D000521	0.013401177	alpha-MSH
## 69	D017471	0.013401177	Receptors, Interferon
## 71	D018389	0.013401177	Codon, Nonsense
## 73	D021901	0.013401177	DNA, Intergenic
## 75	D047888	0.013401177	Receptors, Tumor Necrosis Factor, Type I
## 77	D047889	0.013401177	Receptors, Tumor Necrosis Factor, Type II
## 13	D009842	0.033169882	Oligopeptides
## 18	D015232	0.039671804	Dinoprostone
## 24	D015237	0.039671804	Dinoprost
## 30	D015395	0.039671804	Histocompatibility Antigens Class I
## 44	D015850	0.039671804	Interleukin-6
## 3	D007375	0.046130386	Interleukin-1
## 55	D016753	0.046130386	Interleukin-10

Switching to a different category is easily done by the ‘category<-’ function. Here, we use **Diseases** (category = “C”).

```
category(meshParams) <- "C"
meshR <- meshHyperGTest(meshParams)
summary(meshR)[!duplicated(summary(meshR)[,7]),c(1,2,7)]
```

##	MESHID	Pvalue	MESHTERM
## 1	D008173	0.02003486	Lung Diseases, Obstructive

MeSH ORA for **Anatomy** (category = “A”).

```
category(meshParams) <- "A"
meshR <- meshHyperGTest(meshParams)
summary(meshR)[!duplicated(summary(meshR)[,7]),c(1,2,7)]
```

```
##      MESHID      Pvalue      MESHTERM
## 15 D009434 0.006722989 Neural Pathways
## 16 D013116 0.006722989      Spinal Cord
## 12 D008184 0.020034859      Luteal Cells
## 1  D004847 0.046130386 Epithelial Cells
```

MeSH ORA for **Phenomena and Processes** (category = "G").

```
category(meshParams) <- "G"
meshR <- meshHyperGTest(meshParams)
summary(meshR)[!duplicated(summary(meshR)[,7]),c(1,2,7)]
```

```
##      MESHID      Pvalue      MESHTERM
## 79 D006239 0.005704067      Haplotypes
## 118 D010805 0.006389037 Physical Conditioning, Animal
## 57 D005684 0.006722989      Gait
## 117 D009043 0.006722989      Motor Activity
## 150 D011597 0.006722989 Psychomotor Performance
## 153 D013008 0.006722989      Somatotypes
## 163 D020029 0.006722989      Base Pairing
## 151 D012420 0.013401177      Running
## 157 D018389 0.013401177      Codon, Nonsense
## 159 D019898 0.013401177 Autocrine Communication
## 161 D019899 0.013401177 Paracrine Communication
## 164 D021901 0.013401177      DNA, Intergenic
## 154 D015810 0.020034859 Linkage Disequilibrium
## 1  D003341 0.026624330      Luteolysis
## 58 D005784 0.026624330      Gene Amplification
## 146 D010807 0.026624330 Physical Endurance
## 5  D005091 0.030980920      Exons
## 110 D008285 0.033169882 Major Histocompatibility Complex
## 102 D006579 0.039671804      Heterozygote
## 63 D005805 0.046130386      Genes, MHC Class I
```

5. Session Information

```
sessionInfo()
```

```
## R version 3.1.2 (2014-10-31)
## Platform: x86_64-apple-darwin13.4.0 (64-bit)
##
## locale:
## [1] en_US.UTF-8/en_US.UTF-8/en_US.UTF-8/C/en_US.UTF-8/en_US.UTF-8
##
## attached base packages:
## [1] grid      parallel  stats4    stats     graphics  grDevices  utils
## [8] datasets  methods  base
##
## other attached packages:
## [1] org.MeSH.Eqc.db_1.2.0      meshr_1.2.2
## [3] org.MeSH.Syn.db_1.2.0     org.MeSH.Bsu.168.db_1.2.0
```

```

## [5] org.MeSH.Atu.K84.db_1.2.0 org.MeSH.Aca.db_1.2.0
## [7] org.MeSH.Hsa.db_1.2.0      MeSH.PCR.db_1.2.0
## [9] MeSH.AOR.db_1.2.0         MeSH.db_1.2.0
## [11] MeSHDbi_1.2.0             org.Hs.eg.db_3.0.0
## [13] cummeRbund_2.8.2         Gviz_1.10.2
## [15] rtracklayer_1.26.2       GenomicRanges_1.18.3
## [17] fastcluster_1.1.13      reshape2_1.4
## [19] ggplot2_1.0.0           Category_2.32.0
## [21] GO.db_3.0.0             RSQLite_1.0.0
## [23] DBI_0.3.1               AnnotationDbi_1.28.1
## [25] GenomeInfoDb_1.2.3      IRanges_2.0.0
## [27] S4Vectors_0.4.0        Biobase_2.26.0
## [29] BiocGenerics_0.12.1     Matrix_1.1-4
## [31] fdrtool_1.2.13         biomaRt_2.22.0
##
## loaded via a namespace (and not attached):
## [1] acepack_1.3-3.3         annotate_1.44.0
## [3] base64enc_0.1-2        BatchJobs_1.5
## [5] BBmisc_1.8             BiocParallel_1.0.0
## [7] Biostrings_2.34.0     biovizBase_1.14.0
## [9] bitops_1.0-6          brew_1.0-6
## [11] BSgenome_1.34.0       checkmate_1.5.0
## [13] cluster_1.15.3        codetools_0.2-9
## [15] colorspace_1.2-4      dichromat_2.0-0
## [17] digest_0.6.4          evaluate_0.5.5
## [19] fail_1.2              foreach_1.4.2
## [21] foreign_0.8-61        formatR_1.0
## [23] Formula_1.1-2         genefilter_1.48.1
## [25] GenomicAlignments_1.2.1 GenomicFeatures_1.18.2
## [27] graph_1.44.0          GSEABase_1.28.0
## [29] gtable_0.1.2          Hmisc_3.14-5
## [31] htmltools_0.2.6       iterators_1.0.7
## [33] knitr_1.8             lattice_0.20-29
## [35] latticeExtra_0.6-26   MASS_7.3-35
## [37] matrixStats_0.10.3    munsell_0.4.2
## [39] nnet_7.3-8           plyr_1.8.1
## [41] proto_0.3-10         R.methodsS3_1.6.1
## [43] RBGL_1.42.0          RColorBrewer_1.0-5
## [45] Rcpp_0.11.3          RCurl_1.95-4.3
## [47] rmarkdown_0.3.10     rpart_4.1-8
## [49] Rsamtools_1.18.2     scales_0.2.4
## [51] sendmailR_1.2-1      splines_3.1.2
## [53] stringr_0.6.2        survival_2.37-7
## [55] tools_3.1.2          VariantAnnotation_1.12.4
## [57] XML_3.98-1.1         xtable_1.7-4
## [59] XVector_0.6.0        yaml_2.1.13
## [61] zlibbioc_1.12.0

```