

Supplemental Information for:

Adenosine-to-inosine RNA editing by ADAR1 is essential for normal murine erythropoiesis

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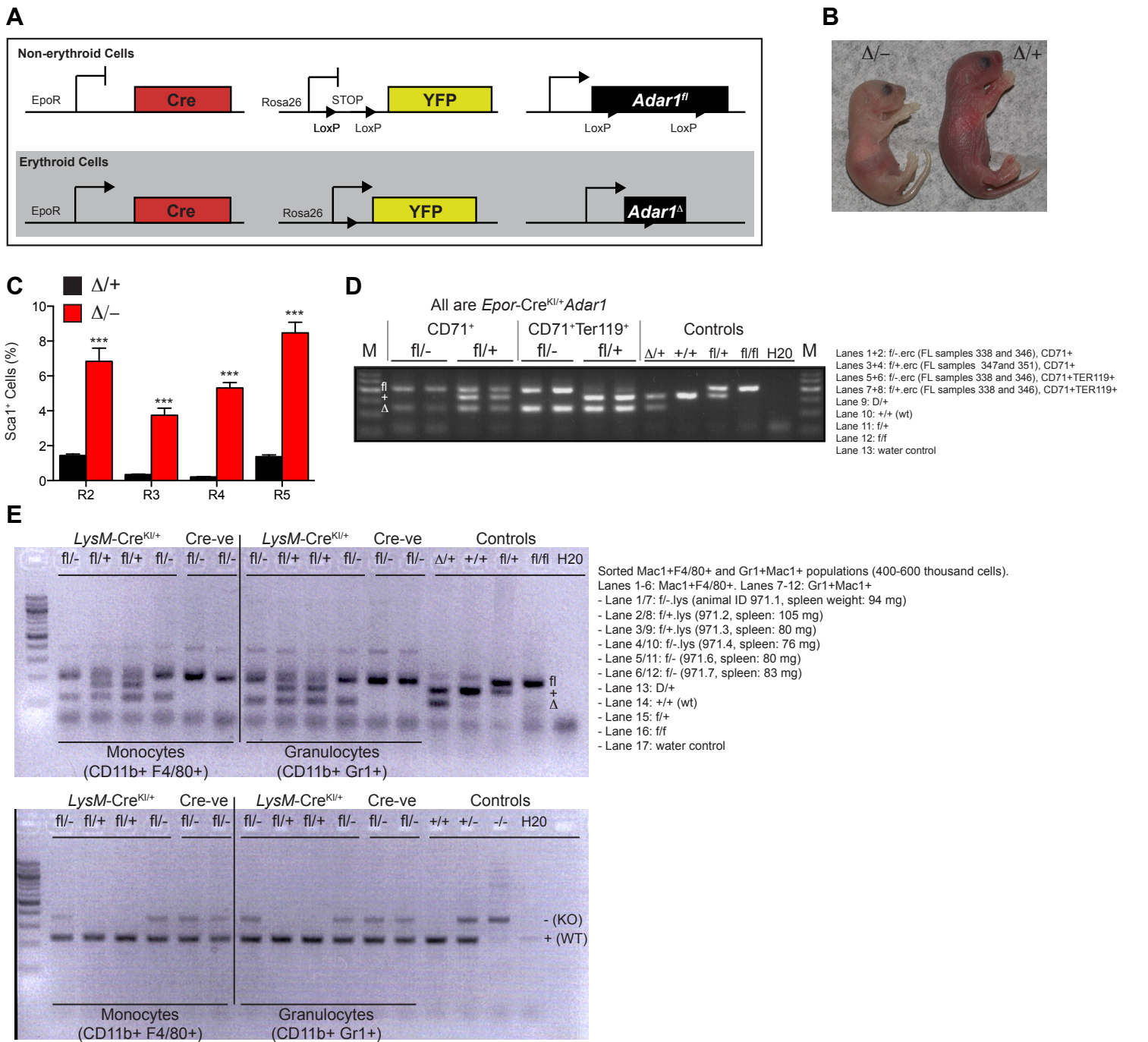
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Running Title: **ADAR1 is essential for erythropoiesis**

Key words: ADAR1, erythropoiesis, RNA editing, dsRNA, retrotransposons, microRNA



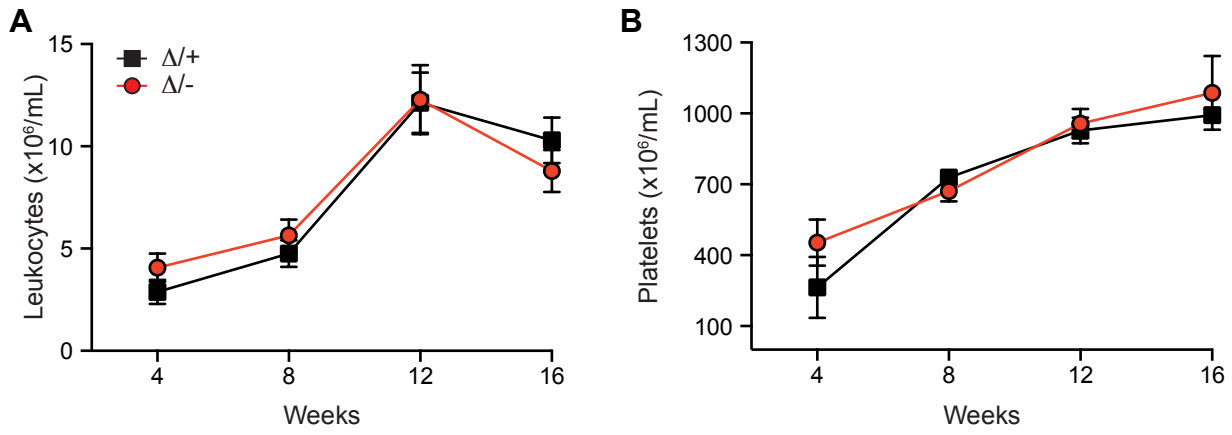


Figure S2. No thrombocytopenia or leukopenia in Epor-Adar1 $\Delta/-$ recipients

Peripheral blood counts of A) leukocytes and B) Platelets in *Epor-Adar1* $\Delta/+$ ($\Delta/+$) and *Epor-Adar1* $\Delta/-$ ($\Delta/-$)FL transplant recipients. Results are mean \pm SEM ($\Delta/+$, n=4 and $\Delta/-$, n=7).

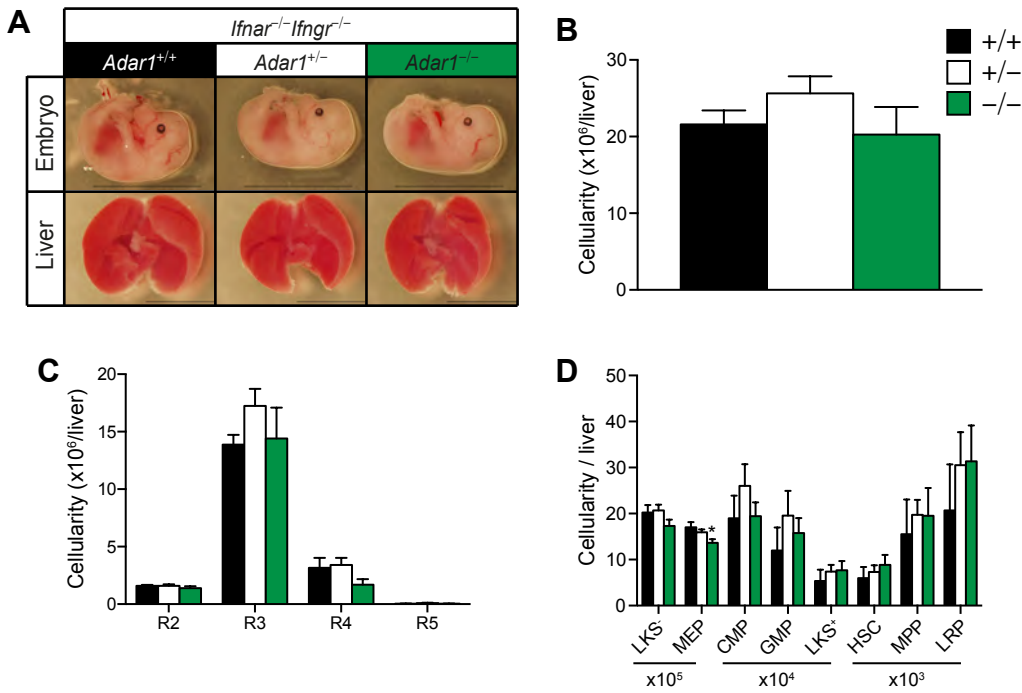


Figure S3. Absence of interferon signaling partially rescues *Adar1*^{-/-} mice

A) Representative images of embryo proper (top) and FL (bottom) at E14.5. Genotypes indicated. Scale bar: 1cm for yolk sac and embryo, 4mm for FL. **B-D)** FL analysis at E14.5. All embryos are *Ifnar*^{-/-}*Ifngr*^{-/-} and *Adar1* genotype is indicated. **B)** Total viable (7AAD⁻) FL cellularity. Enumeration of **C)** erythrocytes and **D)** HSPCs. Results are mean±SEM (+/+ n=4, +/- n=8 and -/- n=4).

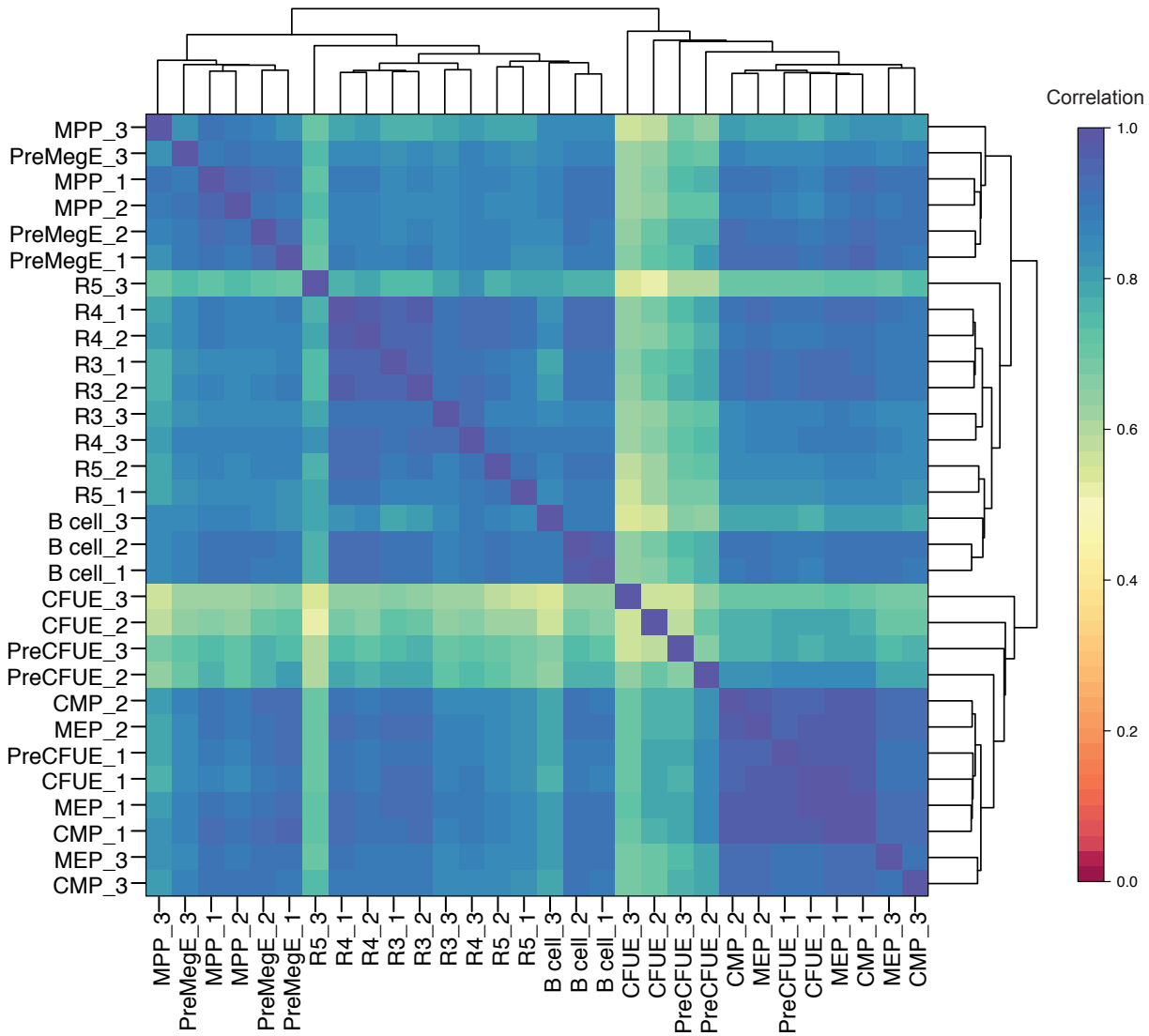


Figure S4. Very little RNA editing differences of known editing sites between diverse cell types

Hematopoietic fractions were isolated from 3 independent wild-type mice. 664 A-to-I editing sites (edited at >1% frequency and coverage >100 reads) were identified by mmP-CR-seq. Colored squares denote total editing frequency correlation between samples, from which hierarchical clustering was based upon. Correlations and statistics were determined by two-tailed Pearson correlation coefficients, $p < 0.05$.

Inc-Tbccd1-IT1
dG = -578.60

Liddicoat_Fig. S5

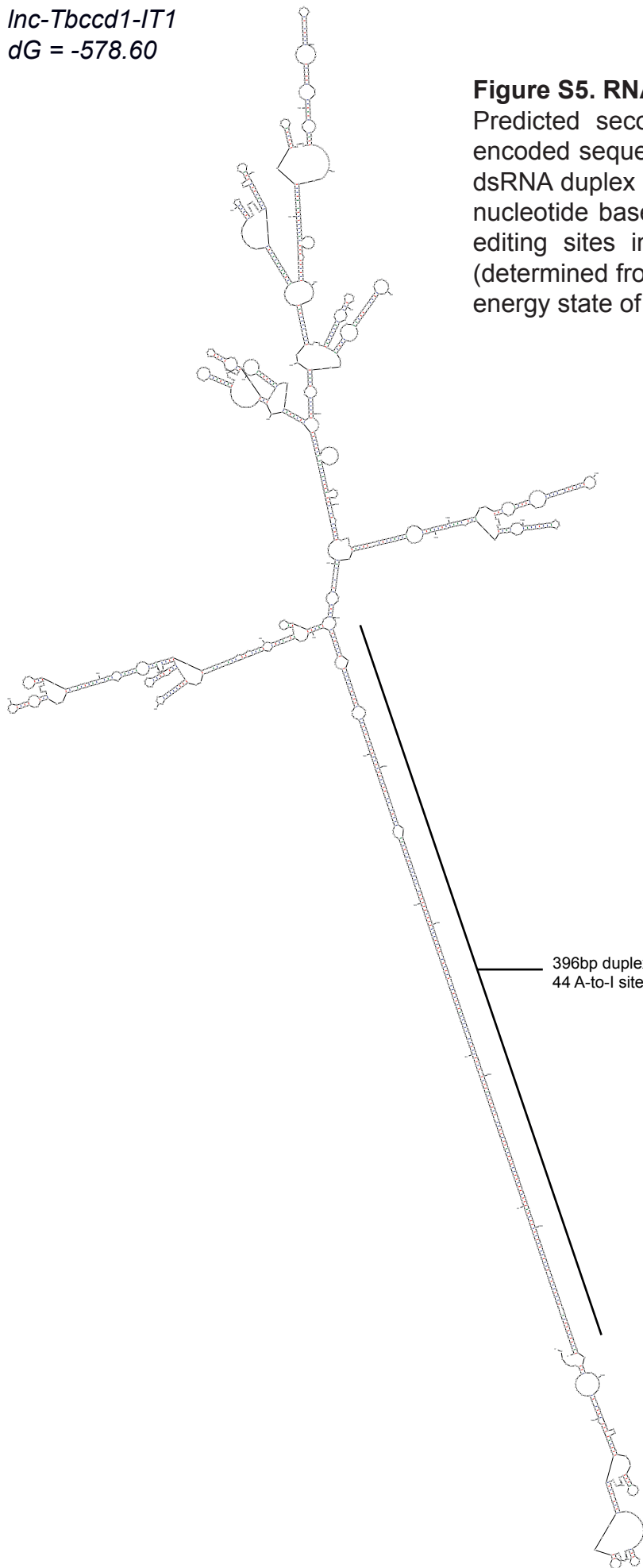


Figure S5. RNA secondary structure of *Inc-Tbccd1-IT1*
Predicted secondary structure of non-edited (genomic encoded sequence) of *Inc-Tbccd1-IT1*. Length of defined dsRNA duplex regions are represented as the number of nucleotide bases in the duplex and the number of A-to-I editing sites in each respective duplexes are defined (determined from editing analysis, but not depicted). Free energy state of predicted structure is depicted as dG.

Primitive

Primitive erythroid cells mature semi-synchronously in the circulation, thus we also used time as a component for their isolation:



Reticulocytes were isolated from peripheral blood, using scatter characteristics to purify them from co-occurring definitive fetal reticulocytes.

All primitive stages were purified using surface expression of Ter119 as well as utilizing a DNA intercalator (Draq5 or Vybrant Violet, VV) and a stain for RNA (Thiazole Orange, TO).

Definitive

Fetal definitive erythroid cells are produced asynchronously and cells were isolated from E14.5 fetal liver, except for reticulocytes isolated from E15.5 peripheral blood.

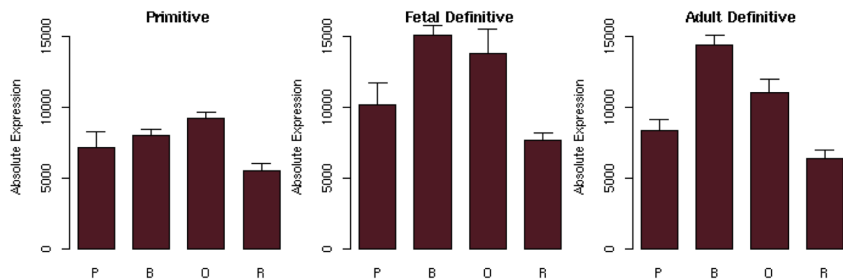
Bone marrow-derived adult definitive erythroid cells also are produced asynchronously and were isolated from mature female bone marrow.

Definitive nucleated erythroblasts were staged by surface phenotype, as well as nuclear condensation and RNA content and size.

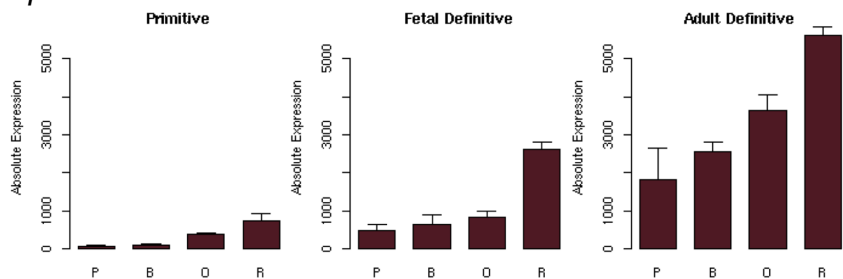


In practice, the more condensed DNA/lower RNA containing definitive erythroblasts were a mix of both polychromatophilic and orthochromatic erythroblasts, so we collected a single fraction and named it PolyO.

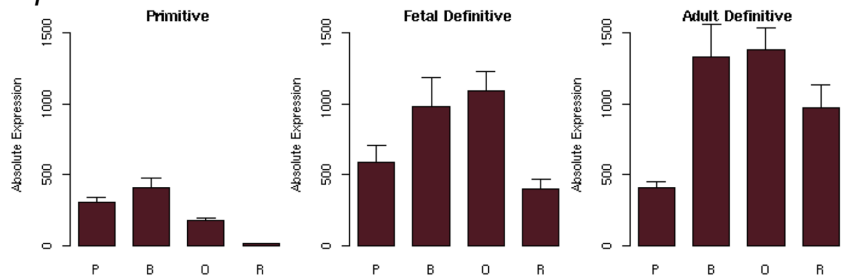
Klf1



Optn



Oip5



Mda5

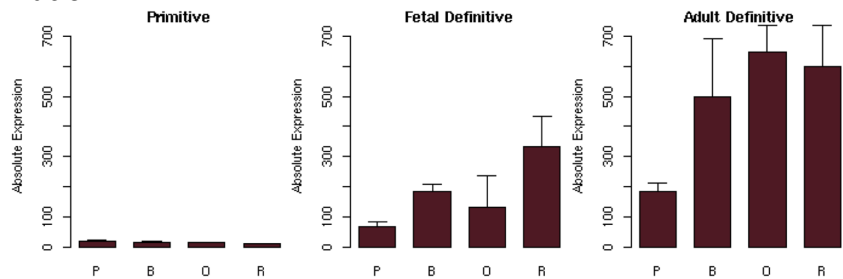


Figure S6. Expression of hyper-edited genes and MDA5 in erythropoiesis

A) Overview of cell-type fractions. B) Expression of hyper-edited genes (*Klf1*, *Optn* and *Oip5*) and MDA5. See <http://www.cbil.upenn.edu/ErythronDB/home.jsp> for details.