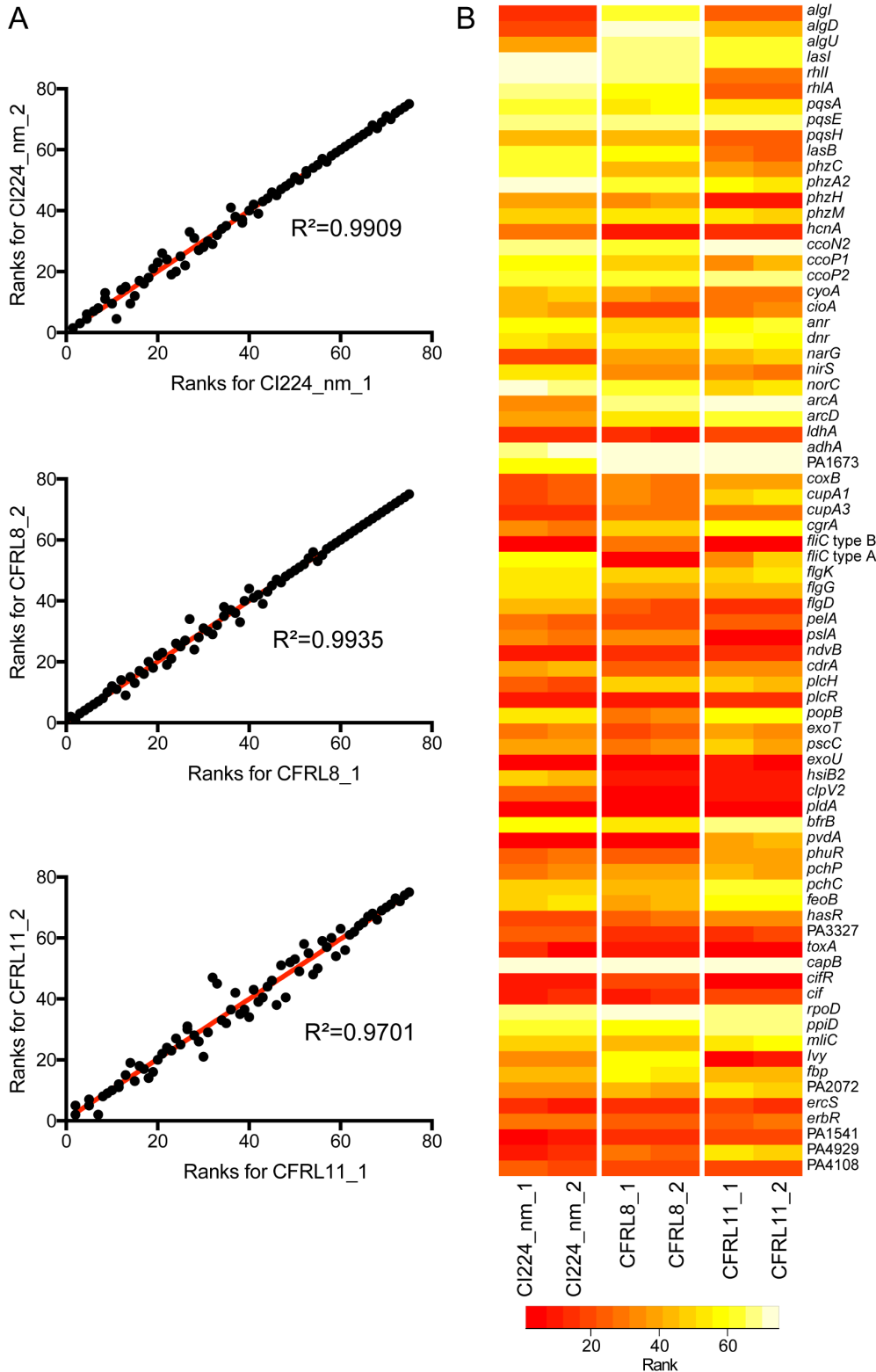
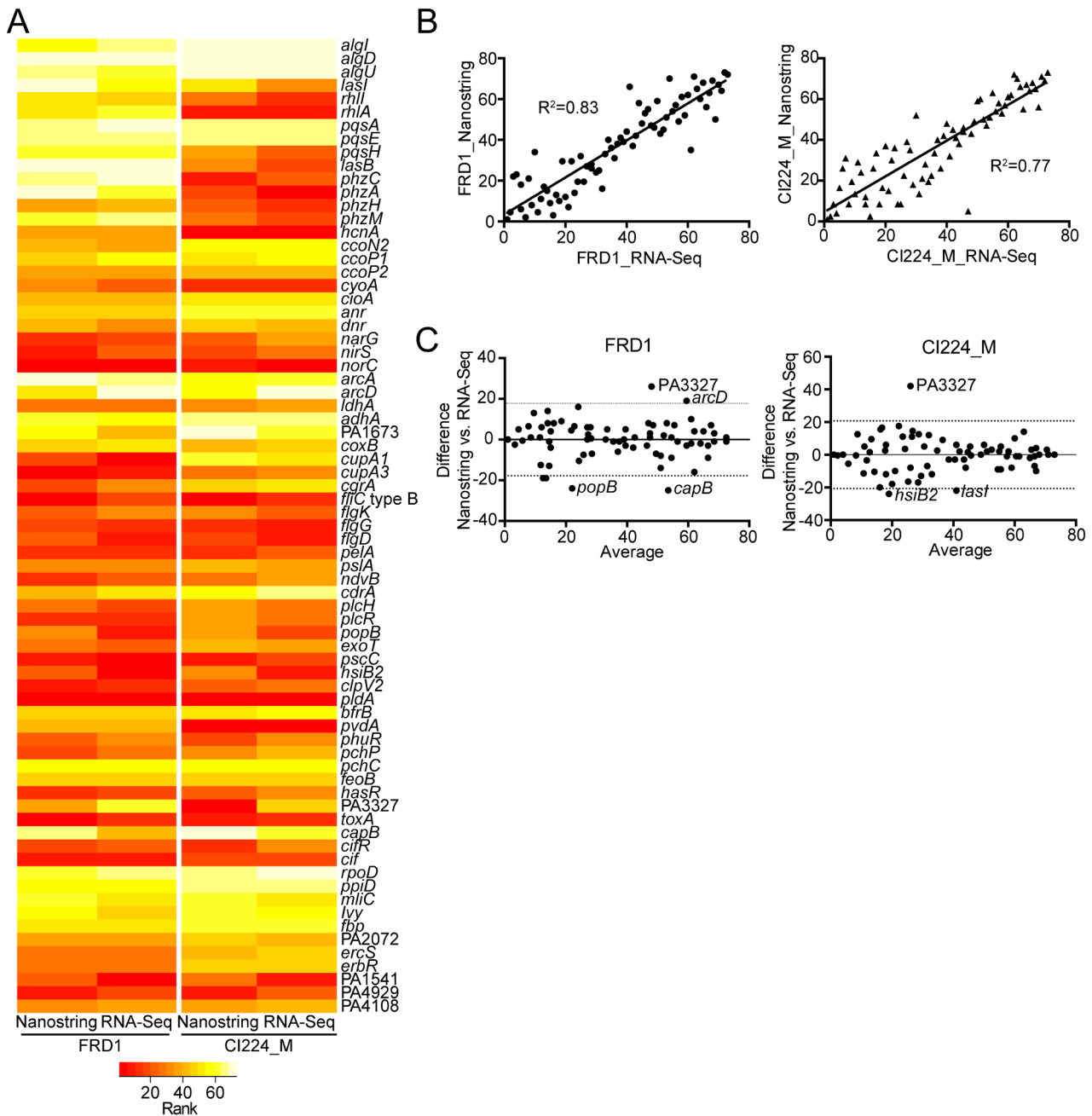


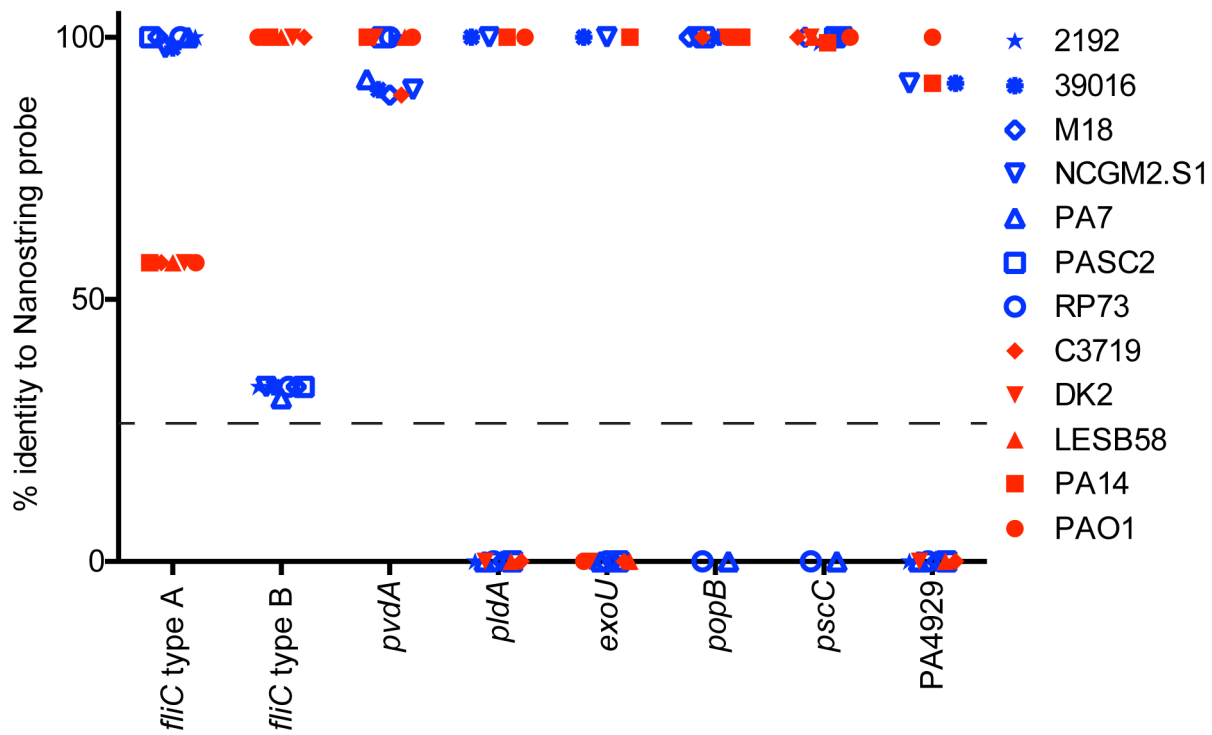
Gifford et al., Supplemental Figures



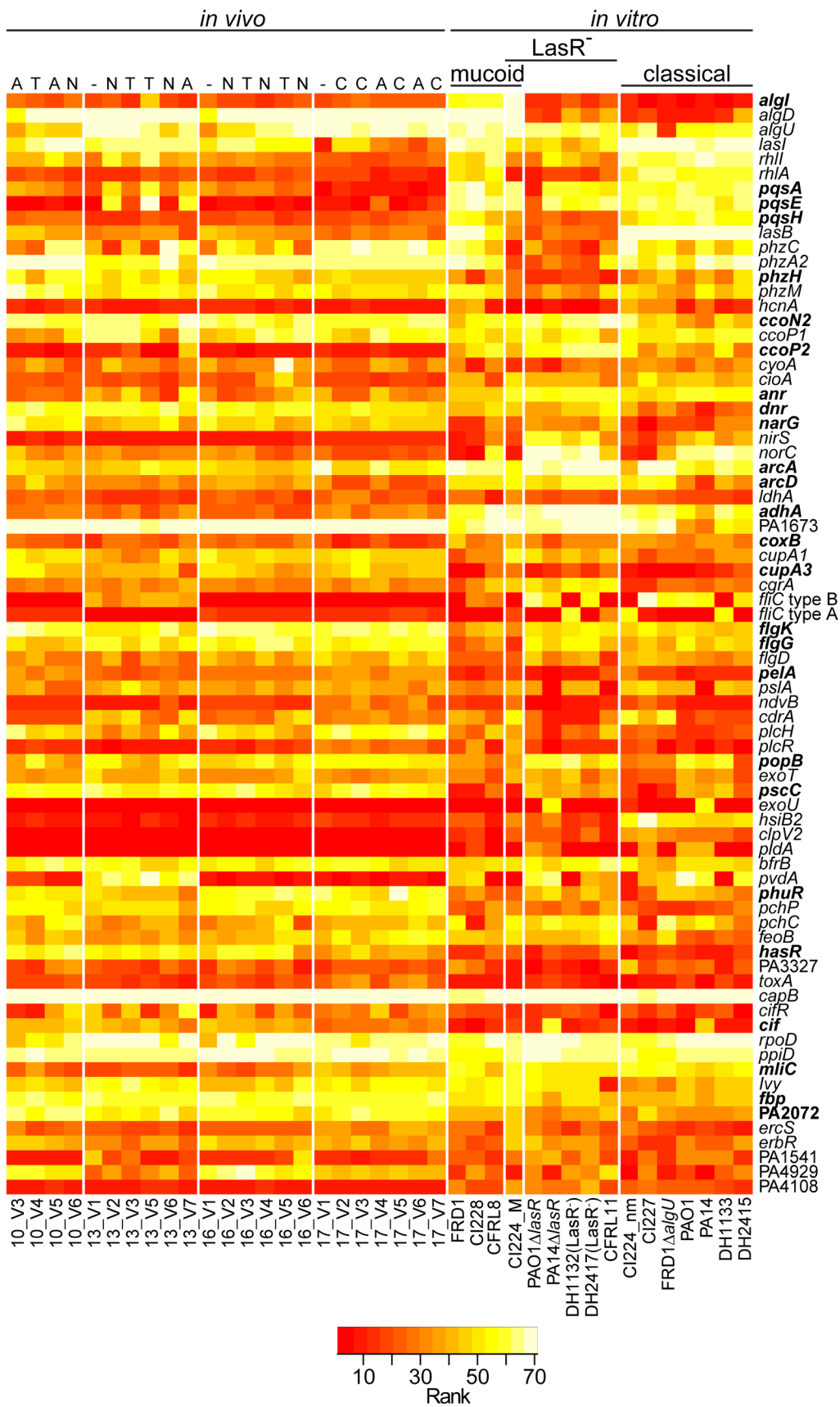
**Fig. S1. Analysis of the reproducibility between duplicate samples.** A. Correlation plot of rank abundances of raw counts obtained from PAV2 Nanostring analysis of colony biofilms formed by three *P. aeruginosa* clinical strains with different colony phenotypes (two colonies per strain). B. Rank abundance heatmap of the three *P. aeruginosa* clinical strains from the PAV2 Nanostring analysis. The raw data are in Table S6.



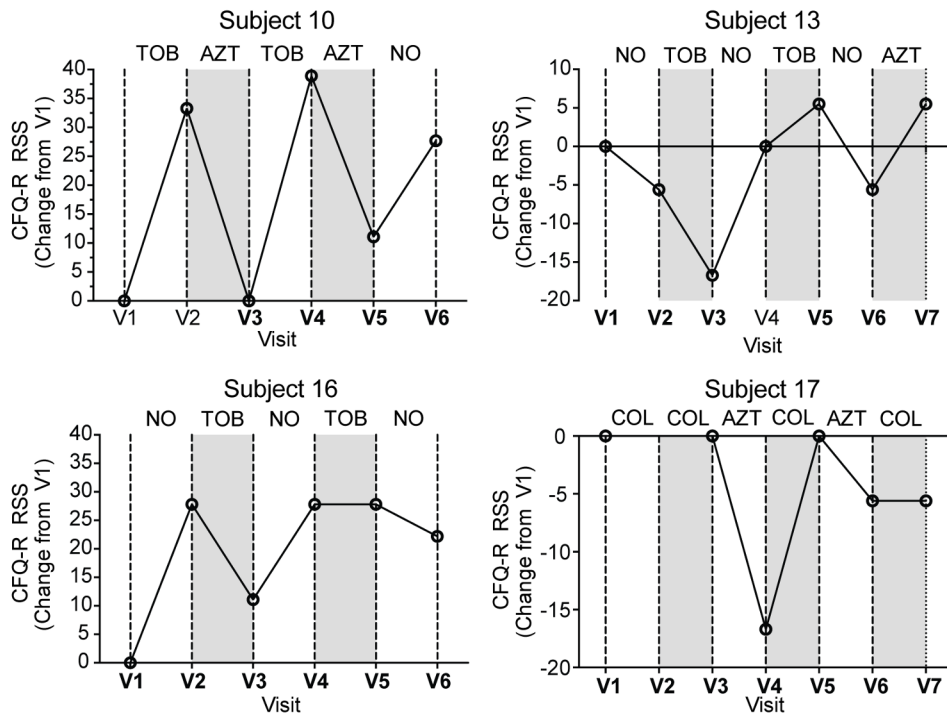
**Fig. S2. Analysis of the agreement between Nanostring analysis and RNA-Seq.** A. Rank abundance heatmap of the *P. aeruginosa* strains FRD1 and CI224\_M from the PAV2 Nanostring analysis (Nanostring) and the RNA-Seq analysis (RNA-Seq). B. Correlation plots of rank abundances of normalized counts obtained from PAV2 Nanostring analysis and the RNA-Seq analysis for the strains FRD1 and CI224\_M. C. Bland-Altman plots of rank abundances of normalized counts obtained from PAV2 Nanostring analysis and the RNA-Seq analysis. The dotted lines represent the 95% confidence interval for agreement between the two transcript level measuring methods. The raw data are in Table S4 (RNA-Seq) and Table S5 (Nanostring).



**Fig. S3. Genomic sequence identities for genes that vary across strains.** Genes with variation in the regions complementary to the Nanostring probe sequence are shown. Each point reflects the percent identity to the PAO1 reference sequence or absence of a detectable homolog. The *P. aeruginosa* strains used in the comparison are shown in the legend. Only genes with at least one instance of a match with less than <90% identity are shown.



**Fig. S4. Analysis of the *P. aeruginosa* transcripts that are significantly different between *in vivo* and *P. aeruginosa* mucoid strains grown *in vitro*.** The heat map shows the rank abundances for all 75 PAV2 transcripts in all samples analyzed. The *in vivo* samples (RNA extracted from sputum) and *in vitro* samples are compared. For the *in vitro* samples, the strain phenotype (classical, mucoid, or LasR-deficient) is shown. For the *in vivo* samples, the subject number and treatment code (A = aztreonam, C = colistimethate, T = tobramycin and N = no treatment) is shown. The sample and strain names are along the bottom. For the *in vivo* samples, the first number indicates the subject (10, 13, 16, and 17) and the number following the “V” indicates the visit number within the six month enrollment period. Letters above each *in vivo* sample column denotes the type of inhaled antibiotic used at the time of sample collection (A = aztreonam, C = colistimethate, T = tobramycin and N = no treatment). Rank abundance analysis indicating levels of transcripts (most abundant in yellow and least abundant in red). The genes highlighted in bold are statistically different between the *in vivo* samples and the mucoid strains grown *in vitro* (FDR corrected and corrected for repeated measures, p-value  $P < 0.05$ ).



**Fig. S5. Changes in the respiratory symptom score (RSS) of the CFQ-R relative to the first visit for each CF subject.** Each plot shows the RSS for a single subject over the course of the six month study. The numbers show the difference relative to the score obtained during the first visit. The type of inhaled antibiotic used the month prior to sample collection is shown (AZT = aztreonam, COL = colistimethate, TOB = tobramycin and NO = no treatment). The visits highlighted in bold were used for *P. aeruginosa* gene expression analysis by Nanostring (PAV2). Samples not used in the analysis were either not collected due to the inability to produce sputum or to the recovery of only minimal or poor quality RNA.