SUPPLEMENTARY DATA

Discharging tRNAs: a tug of war between translation and detoxification in *Escherichia coli* Irem Avcilar-Kucukgoze¹, Alexander Bartholomäus^{1,2}, Juan A. Cordero Varela¹, Robert Franz-Xaver Kaml³, Peter Neubauer³, Nediljko Budisa⁴, Zoya Ignatova^{1,2,#}

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Supplementary Figure 1. Aminoacylation level of tRNA isoacceptors in LB medium. (A) Northern blot analysis of the charging levels of tRNA^{Ser}CGA, tRNA^{Ser}GCU, tRNA^{Ser}UGA and tRNA^{Asp}GUC resembles the microarray analysis (Figure 1A). *E. coli* MC4100 were grown in LB at 37 °C and analyzed in the exponential phase at OD_{600} =0.5. Aminoacyl-tRNAs (A) were isolated at pH 4.5 to preserve the aminocyl-tRNAs and total tRNAs (T) were isolated at pH 8.2, which deacetylates all tRNAs. tRNAs were probed with ³²P-labeled full-length tDNAs that specifically pair to only one

tRNA isoacceptor. The numbers denote the percentage of the aminoacyl-tRNA from the total (aminoacylated and deacylated) in the corresponding lane. Black and gray arrows indicate the migration positions of deacylated and aminoacylated tRNAs, respectively. tRNA^{Ser}CGA reads UCG codons, tRNA^{Ser}GCU pairs to AGC/U codons and tRNA^{Ser}UGA reads UCA/U/G codons. tRNA^{Asp}GUC pairing to GAC/U, which is nearly 100% charged, is included for comparison. (B) Microarray of aminoacyl-tRNAs from *E. coli* BL21 (DE3) at OD₆₀₀=0.5 grown in LB at 37°C. For more details see the legend to Figure 1A. Data are means ± S.D. of two biological replicates. (C) Aminoacylation levels of all tRNAs^{Ser} in LB medium supplemented with 0.4% (w/v) glucose (dark gray) or 10 mM serine (light gray) and compared to growth in LB medium (black bars, panel A). Data are means ± S.D. of two biological replicates. For more details see the legend to Figure 1A.



Supplementary Figure 2. Read calibrations and aminoacylation level of tRNA isoacceptors in different growth phases. (A) Cumulative plots using 23-26 nt long sequencing reads for all ORFs detected over the threshold of 60 rpM (1). Start denotes the first nucleotide of the start codon; stop designates the last nucleotide of the stop codon. Upper and lower panels represent LB and MM, respectively. Ribosome-protected fragments are in black and mRNA reads form the RNA-seq are in red. (B) Northern blot analysis of the charging levels of tRNA^{Ser}GCU pairing to the UCG codon at different ODs of *E. coli* MC4100 grown in LB at 37 °C. A denotes aminoacyl-tRNA and T the total deacylated tRNAs. The numbers under the blot denote the percentage of the aminoacyl-tRNA from the total (aminoacylated and deacylated) in the corresponding lane. Specific ³²P-labeled DNA probes were used to visualize 5S rRNA and tRNA^{Ser}GCU. Dark gray, white and black arrows indicate the position of 5S rRNA, aminoacyl-tRNA and deacylated tRNA, respectively. (C) Microarray of aminoacyl-tRNAs from *E. coli* MC4100 grown in LB medium to OD₆₀₀=1.5 (black), OD₆₀₀=2.5 (light gray) and OD₆₀₀=3.5 (dark gray). Data are means \pm S.D. For more details see the legend to Figure 1A.



Supplementary Figure 3. Growth curves and aminoacylation level of tRNA isoacceptors in different media. (A) Growth curves of cells grown in LB (black circle), MM (grey triangle) and MM+AA (open square). Data are means \pm S.D. of two biological replicates. (B) Microarray of aminoacyl-tRNAs from *E. coli* MC4100 grown in LB medium (black), MM+AA (light gray) and MM (dark gray) to OD₆₀₀=0.5. Data are means \pm S.D. of two-three biological replicates. For more details see the legend to Figure 1A. (C) Northern blot analysis of the charging levels of

tRNA^{Ser}CGA and tRNA^{Ser}UGA from *E. coli* MC4100 grown in MM+AA (OD₆₀₀ =0.5) at 37 °C. tRNA^{Ser}CGA reads UCG codons and tRNA^{Ser}UGA reads UCA/U/G codons. A denotes aminoacyl-tRNA and T the total deacylated tRNAs. The numbers denote the percentage of the aminoacyl-tRNA from the total (aminoacylated and deacylated) in the corresponding lane. Black and gray arrows indicate the positions of deacylated and aminoacylated tRNAs, respectively.

SUPPLEMENTARY REREFENCES

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Supplemenatry Table 1: Summary of the *E.coli* tRNAs with the corresponding codon they recognize. Modified nucleosides at the wobbling position in the anticodon are underlined.

tRNA	Recognized codon (5'-3')	tRNA name				
anticodon (5'-3')						
tRNA ^{Ala} <u>U</u> GC, GGC⁴	GCU, GCA, GCG, GCC	Ala1B, Ala2 ¹				
tRNA ^{Arg} ACG⁵	CGU, CGC, CGA	Arg2				
tRNA ^{Arg} CCG	CGG	Arg3				
tRNA ^{Arg} UCU	AGA	Arg4				
tRNA ^{Arg} CCU	AGG	Arg5				
tRNA ^{Asn} GUU	AAC, AAU	Asn				
tRNA ^{Asp} GUC	GAC, GAU	Asp1				
tRNA ^{Cys} GCA	UGC, UGU	Cys				
tRNA ^{GIn} UUG, CUG	CAA, CAG	GIn1, GIn2 ¹				
tRNA ^{Glu} UUC	GAA, GAG	Glu2				
tRNA ^{Gly} CCC	GGG	Gly1				
tRNA ^{Gly} UCC	GGA, GGG	Gly2				
tRNA ^{Gly} GCC	GGC, GGU	Gly3				
tRNA ^{His} GUG	CAC, CAU	His				
tRNA ^{lle} GAU	AUC, AUU	lle1				
tRNA ^{lle} CAU ⁶	AUA	lle2				
tRNA ^{Leu} CAG	CUG	Leu1				
tRNA ^{Leu} GAG	CUC, CUU	Leu2				
tRNA ^{Leu} UAG ⁴	CUA, CUU, CUG	Leu3				
tRNA ^{Leu} CAA	UUG	Leu4				
tRNA ^{Leu} UAA	UUA, UUG	Leu5				
tRNA ^{Lys} UUU	AAA, AAG	Lys				
tRNA ^{Met} CAU	AUG	Metf1, Metf2 ^{1,2}				
tRNA ^{MEt} CAU	AUG	Metm ³				
tRNA ^{Phe} GAA	UUC, UUU	Phe				
tRNA ^{Pro} CGG	CCG	Pro1				
tRNA ^{Pro} GGG	CCC, CCU	Pro2				
tRNA ^{Pro} UGG ⁴	CCA, CCU, CCG	Pro3				
tRNA ^{Sec} UCA	UGA	Sec				
tRNA ^{Ser} UGA ⁴	UCA, UCU, UCG	Ser1				
tRNA ^{Ser} CGA	UCG	Ser2				
tRNA ^{Ser} GCU	AGC, AGU	Ser3				
tRNA ^{Ser} GGA	UCC, UCU	Ser5				
tRNA ^{Thr} GGU	ACC, ACU	Thr1, Thr3 ¹				
tRNA ^{Thr} CGU	ACG	Thr2				
tRNA ^{Thr} <u>U</u> GU⁴	ACA, ACU, ACG	Thr4				
tRNA ^{Trp} CCA	UGG	Trp				
tRNA ^{Tyr} GUA	UAC, UAU	Tyr1, Tyr2 ¹				
tRNA ^{Val} UAC ⁴	GUA, GUG, GUU	Val1				

tRNA ^{Val} GAC	GUC, GUU	Val2A, Val2B ¹

¹Note that the probe on the array recognizes two tRNAs. ²Metf denotes initiator tRNA. ³Metm denotes elongator tRNA. ⁴(m)cmo⁵U at U34 of a tRNA enables base pairing with U, A and G at the third positions of codons (2,3) 5 Inosine enables base pairing with U, C and A at the third positions of codons (3,4) ${}^{6}k^{2}C$ enables pairing with A at the third positions of (3,4)

Amino acid	Concentration, mM						
	MM	MM+AA					
Ala	0.027 ± 0.005	1.21 ± 0.53					
Asn	0.002 ± 0.0004	0.32 ± 0.01					
Gln	n.d.	0.21 ± 0.01					
Gly	0.062 ± 0.005	0.36 ± 0.05					
His	0.008 ± 0.002	$\begin{array}{c} 0.13 \pm 0.03 \\ 0.29 \pm 0.01 \\ 0.25 \pm 0.03 \end{array}$					
lle	0.009 ± 0.006						
Lys	0.024 ± 0.021						
Phe	0.012 ± 0.005	0.27 ± 0.01					
Ser	0.057 ± 0.005	4.00 ± 0.30					
Thr	0.008 ± 0.006	0.34 ± 0.01					
Trp	0.065 ± 0.005	0.77 ± 0.03					
Tyr	0.011 ± 0.004	0.21 ± 0.01					
Val	0.185 ± 0.027	3.00 ± 0.63					

Supplementary Table 2. Intracellular amino acids concentration in exponentially growing *E. coli* MC4100. Concentrations are mean of two biological replicates \pm S.D. n.d., not determined.

Supplemenatry Table 3: Alterations of the onset of exponential growth by some amino acids added to MM. The duration of the lag phase was measured from the growth curves in Figure 3. n,t., not tested; g.i., growth inhibition. Gray shadowed cells represent the concentration of each amino acid in MM+AA (see also Materials and Methods section).

Amino	tRNA	Aminoacylation level (%) in:			Lag phase duration, h										
aciu					Concentration of the corresponding amino acid in the medium, mM										
		LB	MM+AA	MM	0	0.05	0.1	0.2	0.4	0.5	1	5	10	15	30
Ser	SerCGA	8 ± 9.3	31 ± 21.8	35 ± 7.7	3.6	n.t.	n.t.	n.t.	n.t.	13.6	g.i.	g.i.	g.i.	g.i.	g.i.
	SerGCU	8 ± 5.5	26 ± 6.9	54 ± 24.4											
	SerGGA	3 ± 0.2	20 ± 2.4	52 ± 17.2											
	SerUGA	5 ± 3.8	18 ± 2.7	40 ± 12.4											
Thr	ThrGGU	41 ± 14.4	41 ± 5.6	48 ± 1.8	3.1	n.t.	n.t.	3.6	3.6	3.6	4.1	8.8	14.2	17.5	18.4
	ThrCGU	27 ± 5.4	31 ± 0.8	34 ± 1.1											
	ThrUGU	30 ± 6.5	36 ± 1.9	49 ± 4.2											
His	HisGUG	50 ± 3.2	55 ± 22	36 ± 2.6	3.1	n.t.	3.9	3.5	n.t.	3.9	4.8	8.5	12.1	15	19.4
Cys	CysGCA	21 ± 7.3	60 ± 5.2	35 ± 11.5	3.1	10.9	21.8	n.t	n.t.	g.i.	g.i.	g.i.	g.i.	g.i.	g.i.