Additional file 14: Positive Control Cell Type Markers

Five positive control genes were chosen for each of the neuron subtypes and intestinal epithelial subtypes, 4 for NG2 Glia, and 10 for every other cell type. These control genes were selected without referring to our results, and whenever possible were chosen from existing lists of cell type markers

Cell Type	Positive Control Genes**	Reference(s)
GABAergic Neurons	GAD2, SLC32A1, SLC6A1, DLX1, DLX2	[8]
Noradrenergic Neurons	DBH , TH , ADRA2A, CYB561, MAOA	[18]
Serotonergic Neurons	TPH2, SLC6A4, DDC, SLC18A2, HTR1B	[22]
NG2 Glia	CSPG4, OLIG1, OLIG2, SOX10	[23]
Neurons	SNAP25, STMN2, ENO2, SYN1, TUBB3	[24]
Astrocytes	SLC1A2, SLC1A3, GFAP, GJB6, FGFR3, AQP4	[24, 25]
Oligodendrocytes (Myelinating)	PLP1, MOBP, SOX10, MAG, MBP	[24, 25]
Microglia	CSF1R, CX3CR1, AIF1, FCGR1A, TREM2	[26]
Adipocytes	ADIPOQ, PLIN, ACSL1, CEBPA, CFD, FASN, GPD1, LIPE, LPL, PPARG	[1, 2]
B Cells	MS4A1 , CD79B , CD79A, CD22, CD40, CR2, FCER2, IGHM, PAX5, TNFRSF13B	[3]
Cardiomyocytes	TNNT2, MYL7, ACTC1, CSRP3, MYBPC3, MYH6, MYH7, MYL2, MYL4, NKX2-5	[4]
Chondrocytes	COMP, MATN1, COL2A1, COL9A1, COL9A2, CHI3L1, SOX9, MATN3, CHAD, SOX6	[5]
Endothelial Cells	FLT1, TIE1, CDH5, EDN1, ENG, KDR, MMRN2, PECAM1, SELE, VWF	[6]
Erythrocytes	GYPA , RHAG , GYPB, GYPC, GYPE, HBA1, HBA2, HBB, RHCE, RHD	adult hemoglobin, glycophorins, Rh blood group antigens, and erythropoietin receptor
Macrophages	EMR1, CD14, CSF1R, ITGAM, CD68, ITGAX, CD33, FCGR1A, FCGR1B, FCGR2A	[7]
NK Cells	KLRD1, KIR2DL1, KLRB1, KLRK1, KIR3DL1, NCAM1, CD244, GZMB, GNLY, PRF1	[3]
Osteoblasts	BGLAP, SP7, ALPL, COL1A1, DLX5, DMP1, MEPE, OMD, RUNX2, SPP1	[5]
Platelets	ITGA2B, GP9, GP1BA, ITGB3, SELP, GP5, MPL, GP6, GP1BB, TBXA2R	[3, 5] plus TBXA2R
Pluripotent Stem Cells	POU5F1 , ZFP42 , SOX2, LIN28A, LIN28B, ZIC3, UTF1, FBXO15, LEFTY1, LEFTY2	[5]
Schwann Cells (Myelinating)	PLP1, PRX, EGR2, MAL, MBP, NGFR, S100B, SOX10, PLP1, PMP2, PMP22	[9, 10]

Simple Epithelial Cells	KRT7, KRT18, AP1M2, CDH1, CGN, DSG2, EPCAM, KRT19, OCLN, TJP3	[11, 12] plus <i>AP1M2</i> and <i>EPCAM</i>
Skeletal Muscle Cells	TNNI1, TNNI2, ACTA1, CHRNA1, ENO3, MYBPH, MYF6, MYL1, MYLPF, TNNC1	[4]
Smooth Muscle Cells	ACTA2, TAGLN, DES, ACTG2, CNN1, CSRP1, LMOD1, LPP, SMTN, VCL	[4]
Stratified Epithelial Cells	KRT14 , KRT15 , DSC1, DSC3, DSG1, DSG3, KRT6A, KRT16, KRT17, PKP1	[11, 12]
T Cells	CD3G, TRA@, CD3E, CD4, CD8A, CD8B, CD28, CD247, TRAC, TRBC1	TCR locus and major TCR co- receptors
Enterocytes	FABP2, SI, APOA1, DPP4, ANPEP	[13, 14]
Enteroendocrine Cells	GCG, PYY, SST, SYP, TPH1	[15]
Goblet Cells	TFF3, MUC5B, CLCA1, FCGBP, AGR2	[16, 17]
Paneth Cells	DEFA6, REG3A, LYZ, REG1A, REG1B	[19, 20]
Podocytes	PTPRO, NPHS1, NPHS2, PODXL, WT1, MME, TJP1, TCF21, CR1	[21]

** Genes in bold were used as negative control markers for non-target cell types in Figure 3. Negative control genes also included the set of reference genes from Eisenberg and Levanon [30]. *KLRD1* and *KIR2DL1* were not included as negative controls for T cells because they are expressed in T cells, *DES* was not included as a negative control for any muscle cell type as it is expressed in all muscle, and *EMR1* and *CD14* were not included as negative controls for microglia because they are expressed in microglia. In addition, bold genes for the neuron subtypes and intestinal epithelial lineages were only included as negative controls for other cell types in the same class (i.e. the GABAergic marker *GAD2* was used as a negative control for the other 3 neural subtypes but not any other cell type).

1. Kim JB, Spiegelman BM: ADD1/SREBP1 promotes adipocyte differentiation and gene expression linked to fatty acid metabolism. *Genes Dev* 1996, **10**:1096–1107.

2. Urs S, Smith C, Campbell B, Saxton AM, Taylor J, Zhang B, Snoddy J, Voy BJ, Moustaid-moussa N: Nutrient-Gene Interactions Gene Expression Profiling in Human Preadipocytes and Adipocytes by Microarray Analysis 1, 2. 2004(August 2003):762–770.

3. Cell Markers [http://www.biolegend.com/cell_markers]

4. Nelander S, Mostad P, Lindahl P: Prediction of cell type-specific gene modules: identification and initial characterization of a core set of smooth muscle-specific genes. *Genome Res* 2003, **13**:1838–54.

5. Research Topics [http://www.rndsystems.com/]

6. Garlanda C, Dejana E: Heterogeneity of Endothelial Cells : Specific Markers. Arterioscler Thromb Vasc Biol 1997, **17**:1193–1202.

7. CD Marker Handbook [https://www.bdbiosciences.com/documents/cd_marker_handbook.pdf]

8. Sugino K, Hempel CM, Miller MN, Hattox AM, Shapiro P, Wu C, Huang ZJ, Nelson SB: **Molecular taxonomy of major neuronal classes in the adult mouse forebrain.** *Nat Neurosci* 2006, **9**:99–107.

9. Jessen KR, Mirsky R: The origin and development of glial cells in peripheral nerves. *Nat Rev Neurosci* 2005, **6**:671–82.

10. Nagarajan R, Le N, Mahoney H, Araki T, Milbrandt J: **Deciphering peripheral nerve myelination by using Schwann cell expression profiling.** *Proc Natl Acad Sci U S A* 2002, **99**:8998–9003.

11. Moll R, Divo M, Langbein L: **The human keratins: biology and pathology.** *Histochem Cell Biol* 2008, **129**:705–33.

12. Franke WW: **Discovering the molecular components of intercellular junctions--a historical view.** *Cold Spring Harb Perspect Biol* 2009, **1**:a003061.

13. Glickman RM: Enterocyte lipid absorption and secretion. :505–526.

14. McConnell RE, Higginbotham JN, Shifrin D a, Tabb DL, Coffey RJ, Tyska MJ: **The enterocyte microvillus is** a vesicle-generating organelle. *J Cell Biol* 2009, **185**:1285–98.

15. Gunawardene AR, Corfe BM, Staton C a: Classification and functions of enteroendocrine cells of the lower gastrointestinal tract. *Int J Exp Pathol* 2011, **92**:219–31.

16. Pelaseyed T, Bergstrom JH, Gustafsson JK, Ermund A, Birchenough GMH, Schutte A, van der Post S, Svensson F, Rodriguez-Pineiro AM, Nystrom EEL, Wising C, Johansson ME V, Hansson GC: **The mucus and mucins of the goblet cells and enterocytes provide the first defense line of the gastrointestinal tract and interact with the immune system**. *Immunol Rev* 2014, **260**:8–20.

17. Sheng YH, Hasnain SZ, Florin THJ, McGuckin M a: **Mucins in inflammatory bowel diseases and colorectal cancer.** *J Gastroenterol Hepatol* 2012, **27**:28–38.

18. Grimm J, Mueller A, Hefti F, Rosenthal A: **Molecular basis for catecholaminergic neuron diversity.** *Proc Natl Acad Sci U S A* 2004, **101**:13891–13896.

19. Bevins CL, Salzman NH: **Paneth cells, antimicrobial peptides and maintenance of intestinal homeostasis.** *Nat Rev Microbiol* 2011, **9**:356–68.

20. Van Beelen Granlund A, Østvik AE, Brenna Ø, Torp SH, Gustafsson BI, Sandvik AK: **REG gene expression** in inflamed and healthy colon mucosa explored by in situ hybridisation. *Cell Tissue Res* 2013, **352**:639–46.

21. Ju W, Greene CS, Eichinger F, Nair V, Hodgin JB, Bitzer M, Lee Y-S, Zhu Q, Kehata M, Li M, Jiang S, Rastaldi MP, Cohen CD, Troyanskaya OG, Kretzler M: **Defining cell-type specificity at the transcriptional level in human disease.** *Genome Res* 2013, **23**:1862–73.

22. Dougherty JD, Maloney SE, Wozniak DF, Rieger M a., Sonnenblick L, Coppola G, Mahieu NG, Zhang J, Cai J, Patti GJ, Abrahams BS, Geschwind DH, Heintz N: **The Disruption of Celf6, a Gene Identified by Translational Profiling of Serotonergic Neurons, Results in Autism-Related Behaviors**. *J Neurosci* 2013, **33**:2732–2753.

23. Dimou L, Gallo V: NG2-glia and their functions in the central nervous system. Glia 2015:n/a-n/a.

24. Zhang Y, Chen K, Sloan S a, Bennett ML, Scholze AR, O'Keeffe S, Phatnani HP, Guarnieri P, Caneda C, Ruderisch N, Deng S, Liddelow S a, Zhang C, Daneman R, Maniatis T, Barres B a, Wu JQ: **An RNA-Sequencing Transcriptome and Splicing Database of Glia, Neurons, and Vascular Cells of the Cerebral Cortex.** *J Neurosci* 2014, **34**:11929–47.

25. Cahoy JD, Emery B, Kaushal A, Foo LC, Zamanian JL, Christopherson KS, Xing Y, Lubischer JL, Krieg P a, Krupenko S a, Thompson WJ, Barres B a: A transcriptome database for astrocytes, neurons, and oligodendrocytes: a new resource for understanding brain development and function. *J Neurosci* 2008, 28:264–78.

26. Elmore MRP, Najafi AR, Koike M a, Dagher NN, Spangenberg EE, Rice R a, Kitazawa M, Matusow B, Nguyen H, West BL, Green KN: Colony-stimulating factor 1 receptor signaling is necessary for microglia viability, unmasking a microglia progenitor cell in the adult brain. *Neuron* 2014, **82**:380–97.