SUPPLEMENTAL MATERIAL

Supplemental Table 1 Previous studies reporting fecal calprotectin in neonates with necrotizing enterocolitis.

| Year (Ref) | First Author | Country | Study Design | FCP with no NEC | FCP with medical NEC | FCP with surgical NEC | Conclusion |
|---------------|-----------------|-------------|--|---|--|--|--|
| 2003 (10) | Carroll | England | N=7 with NEC vs 7 controls | N= 7; 98 ±61 μg/g stool | N= 7; 288 ±49 μg/g stool | NR | FCP levels are higher in NEC (P=0.0006) |
| 2007 (13) | Josefsson | Sweden | N=4 with NEC, 3 with perforation of other etiology vs 52 healthy preterm controls | N= 52; 253 µg/g stool | N= 3; Mean 13298 (3646- 22513) μg/g stool | N=1, 685 µg/g stool | FCP >2000 µg/g is a useful but not an early marker of NEC |
| 2008 (14) | Yang | USA | N=14 VLBW serial samples for first month | 123±99 μg/g stool | 380±246 µg/g stool* | NR | FCP may be a marker for early diagnosis of gastrointestinal illness |
| 2010 (15) | Thuijls | Netherlands | N=35 suspected of NEC | N=21; 69.6 (1-556.4) μg/g stool | N=14; 589.2 (146.1-847.6) µg/g stool | NR | FCP is a promising diagnostic marker for NEC |
| 2012 (16) | Aydemir | Turkey | N=25 with NEC and 25 controls | N=25; 365 (58-1006) μg/g stool | N= 16; 1285 (241-2606) μg/g stool | N= 9; 1251 (421- 3337) µg/g stool | FCP >792 μg/g stool has a 76% sensitivity and 92% specificity for identifying NEC |
| 2012 (17) | Dabritz | Germany | N= 145 (127 control and 18 with NEC). Stool samples alternate days for 4 weeks | NR | N=18; 349 (1-850) mg/kg | NR | FCP levels are elevated at onset of NEC but not correlated with disease severity |
| 2012 (18) | Zoppelli | Germany | N= 1988 stool samples from 206 VLBW neonates | N= 38; Mean 52.7 µg/g stool | N= 14 with moderate NEC. Mean NR. | N=5 with fulminant NEC; Mean 14.4 µg/g stool | FCP is a sensitive and specific marker for NEC. However, FCP was unusually low in fulminant NEC. |
| 2012 (19) | Selimoglu | Turkey | N=14 with NEC and 40 healthy preterm controls | N= 40; 172±171 μg/mL | N=14; 167.6±143 µg/mL | N=1; 420 µg/mL | FCP not useful in diagnosing NEC |
| 2012 (20) | Aydemir | Turkey | N=10 with NEC and 9 controls | N= 9; Mean 104 (Range 92- 123) mg/dL | N=10; Mean 185 (Range 165-198) mg/dL | NR | FCP is a useful marker for diagnosis and severity of NEC |
| 2012 | Reisinger | Netherlands | N=62 with clinical suspicion of NEC | N=19; 80 (1-625) | N=16; 402 (108-848) | NR | FCP > 286 µg/g stool has an 81% sensitivity |

| (21) | | | | µg/g stool | µg/g stool | | and 79% specificity for identifying NEC |
|--------------|-----------|-------------|---|---------------------------------|---------------------------------------|--------------------------------------|---|
| 2013 (22) | Jenke | Germany | N= 68 ELBW neonates. Compared gut microbiota and FCP. | NR | NR | NR | No correlation between FCP level and total stool bacterial count of any analyzed bacteria. |
| 2014 (23) | Reisinger | Netherlands | N=29 with NEC (12 medical and 17 surgical or fatal) | NR | N= 12; 375 (146-848) μg/g stool | N=17; 479 (108-684) μg/g stool | FCP did not differentiate between medical vs surgical/fatal cases (p=0.87) |
| 2014 (24) | Yoon | Korea | N= 16 VLBW infants. Stools serially collected. | N=12; 4.34±1.94 mg/kg | N= 4; 5.75±1.98 mg/kg | NR | FCP higher in those with NEC (p=0.02) |
| 2014 (25) | Albanna | Egypt | N=15 with NEC and 20 controls | N=20; 73±57 mg/dL | N=12; 240±58 mg/dL | N=3; 308±4 mg/dL | FCP could be a valuable tool to investigate NEC (p<0.0001) |
| 2015 (8) | Bin-Num | Israel | N= 15 infants, 29 stool samples. Compared bed- side FCP assay to ELISA. | 195 (110- 440) μg/g stool | 3000 (2075- 7875) μg/g stool | NR | FCP useful for rapid diagnosis of NEC (p<0.0001) |

Ref, reference number; FCP, fecal calprotectin; NEC, necrotizing enterocolitis; NR, not reported. FCP values were reported variably as "µg/g stool", "mg/kg", or "mg/dL"; *infants were "sick" but not necessarily with a diagnosis of NEC.

Supplemental Figure 1. *Human NEC gastrointestinal tissue samples demonstrate NET formation.* (A) Using immunohistochemical techniques, we compared NET formation in gastrointestinal tissue samples collected from 5 different infants who underwent surgical treatment for NEC and 6 control infants operated on for indications other than NEC. Representative images from infants with NEC are shown in a 60x magnified image demonstrating PMNs and NETs. Calprotectin is seen in gold fluorescence. Neutrophil elastase, a protein known to be expressed on NETs, is seen in magenta fluorescence. Nuclear and NET-associated DNA is shown in blue fluorescence. White arrowheads denote NETs, and yellow arrowheads denote PMNs which express calprotectin. Bar, $10\mu m$ (B, C) We quantified PMN (B) and NET formation (C) in the immunohistochemical images from infants with NEC represented in A and from control infants. 5 randomly selected high power fields were analyzed for each of the 5 NEC and 6 Control tissue samples. PMNs were defined as cells having neutrophil nuclear morphology and staining positive for both calprotectin and NE. ** denotes p<0.001. We employed the student's T-test as a statistical tool.

Supplemental Figure 2. *LPS treatment fails to induce robust expression of calprotectin mRNA or protein.* (**A**) Using real-time RT-PCR, we quantified calprotectin and IL-8 mRNA expression in control and LPS-stimulated (100 ng/mL) adult PMNs. The y-axis represents LPS-induced mRNA expression shown as a fold change over baseline normalized to β -actin. Baseline mRNA expression is arbitrarily set at 1 (dashed line). Calprotectin mRNA expression (left column) was induced 1.8 fold by LPS while IL-8 mRNA (right column) was induced 8 fold as a control for PMN stimulation. (**B**) We used immunocytochemistry to qualitatively assess calprotectin protein expression in control and LPS-stimulated (100 ng/mL, 2 hours) PMNs. Representative images from LPS-stimulated PMNs are shown here. Calprotectin protein expression is shown in gold fluorescence. Nuclear DNA as a counterstain is shown in blue fluorescence. These images are representative of 3 separate experiments using PMNs isolated from 3 different healthy adult donors. Bar, 20µm.