

Title Page

Title: Introducing pharmacogenetic testing with clinical decision support into primary care: a cohort study.

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60**Abstract**

Background: Inappropriate prescribing increases patient morbidity and death due to adverse drug events. The inclusion of genetic information into primary care medication practices is one solution. Our aim was to develop, and to determine the levels of use and usability of, a decision support tool which creates medication options adjusted for patient characteristics, drug-drug interactions, and pharmacogenetics.

Methods: We conducted a cohort study in six primary care settings, enrolling 191 adults with at least one of ten common diseases. Genotyping was undertaken, and genotypic data was linked to an evidence-based prescribing decision support system. The primary end point was ability to obtain and genotype samples. The secondary end points were yield and purity of DNA samples, ability to link results to decision support software, use of the decision support software, and feedback from users.

Results: Genotyping resulted in 189 (99%) patients with pharmacogenetic reports linked to the decision support program. We found 96.8% of samples had at least one actionable genotype for medications included in the decision support system. The medication support system was used by the physicians and pharmacists 236 times over a period of three months. These health professionals stated that the clinical decision support makes it easy to incorporate genetic information into decision-making and helps reduce inappropriate prescribing.

Interpretation: Physicians and pharmacists can collect saliva samples of sufficient quantity and quality for DNA extraction, purification and genotyping. Use of a clinical decision support system with integrated data from pharmacogenetic tests may result in safer prescribing practices.

Trial registration: The University of British Columbia Clinical Research Ethics Board approved the study (H14-02979), and it was registered at ClinicalTrials.gov (NCT02383290).

Introduction.

Drug-related adverse events in primary care are a significant and common cause of morbidity^{1,2} with incidence rates as high as 25%.³ There is high level evidence that pharmacogenetic testing is an effective method of reducing adverse drug events.⁴⁻⁶ Results from a US cohort of 1143 patients experiencing adverse events indicate that a clinically significant 33% were due to drug-gene interactions or drug-drug-gene interactions.⁷ Of the approximately four billion prescriptions filled in the United States in 2013, 18% had actionable pharmacogenetics.⁸

Many of the drugs studied in pharmacogenetic trials are part of the primary care drug formulary and used for common conditions. Pharmacogenetic panels are now available at an affordable price, and patients are requesting the tests and asking physicians to use these results in their care. Prior to implementing a primary care pharmacogenetic panel, it is necessary to consider the ability of healthcare providers to incorporate this information into current medication selection processes.

Computerized order entry systems in electronic medical records, using drug database systems, enable the user to identify potential drug-drug and drug-condition reactions at the time of medication selection. Despite the introduction of these systems, and the increased use of electronic medical records, the prevalence of inappropriate prescriptions (35%) and high risk prescriptions (14%) remains very high.¹⁰⁻¹² Alert systems have proven ineffective at changing prescribing decisions.¹³ Physicians and pharmacists describe alert fatigue, and it has been demonstrated that non-interrupting dynamically annotated visualization are more effective than alerts in reducing inappropriate imaging orders.¹⁴ This may also be true for prescribing; avoiding alerts may lead to a reduction in inappropriate prescriptions. Medication decision support systems (MDSS) need to be able to show not only other classes of drugs for that condition, but assess whether the drug is safe and effective for the individual, taking into account other medications, diseases and the person's physical state.⁹ We have developed a patient-centered MDSS that evaluates a list of potential drugs for a specific condition. The MDSS assesses the potential drug-

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3 drug, drug-condition, drug-gene and drug-drug-gene interactions, and produces a list of drug
4 options least likely to cause harm and most likely to be effective for that person.
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7 We conducted a study to assess the DNA collection process, investigate a panel of
8 pharmacogenetic tests relevant to primary care patients, and assess a condition-based genetic-
9 informed medication decision support system.
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12 **Methods**

13 ***Pharmacogenetic Panel Development***

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15 Evidence for genotype guided dosing recommendations was compiled for drugs commonly
16 prescribed by family physicians. Single nucleotide polymorphisms (SNPs) and copy number
17 variants (CNVs) were ranked according to clinical annotations primarily from PharmGKB,¹⁵ the
18 Clinical Pharmacogenetic Implementation Consortium (CPIC)¹⁶ and the Royal Dutch
19 Pharmaceutical Association review (DPWG)⁵. Based on information from the PharmaADME
20 Consortium (www.PharmaADME.org), as well as guidelines and drug labels, a pharmacogenetic
21 panel was selected. This panel included 33 of the top ranking genetic variants in the following
22 genes: *CY2C9*, *CYP2C19*, *CYP2D6*, *G6PD*, *HLA-B*, *SLC01B1* and *VKORC1*. Modifications were
23 made to this list in light of new evidence of clinically relevant SNP tests and resulted in a
24 customized panel of 24 genetic variants for 22 drugs.
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28 TaqManTM allelic discrimination qPCR assays were chosen based on the manufacturer
29 guarantee of working assays on the Quant StudioTM 12K Flex Platform (Applied Biosystems). We
30 developed a quality control and validation process to test the sensitivity and specificity of the
31 assays. This included using Coriell Biorepository control samples, analyzing the results and
32 comparing the experiment with known genotypes to determine feasibility and accuracy of the
33 genetic test, and using sample replicates to assess concordance. In addition, a subset of
34 experimentally determined genotypes was confirmed using Sanger sequencing. Once validated,
35 these assays were incorporated onto our primary care pharmacogenetic panel.
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3 A pharmacogenetic report was developed giving details on the method of testing, the genes
4 tested, the drugs implicated, the alleles, and the predicted phenotype. Software was developed
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6 to transfer the genotype information into the MDSS.
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9 10 ***Medication Decision Support Development***

11 Ten common diseases were selected as being relevant to primary care and having the potential for
12 pharmacogenetic test use: gout, chronic obstructive pulmonary disease, migraine, depression,
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14 osteoarthritis, hypertension, hyperlipidemia, atrial fibrillation, osteoporosis and epilepsy. As five of
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16 the diseases have two distinct therapeutic approaches, the ten diseases result in 15 “conditions”.
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19 For example, the therapeutic options are very different for a person with an acute flare-up of gout
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21 than a person in the chronic stages of gout. Having identified published epidemiological evidence
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23 for the management of the condition a team of pharmacists, physicians, and experts in
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25 evidence-informed healthcare used, i) the highest levels of evidence for treatment selection,
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27 ii) known drug-drug interactions from standard databases, published studies, and product
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29 monographs, and iii) drug-genetic information from PharmGKB and other resources to form
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31 logic trees. From the logic trees, we developed a set of rules based on factors that affect
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33 selection of optimal drug therapy for each condition, and these were programmed into the
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35 decision support tool.
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39 The result of entering data into the MDSS is the generation of a list of drug options adjusted for
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41 the individual’s medical history, biophysical profile and genetic test results, as demonstrated in
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43 Figure 1.
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46 47 ***Study Design and Participants***

48 We used a prospective cohort study design. Due to the known association between HLA-
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50 B*58:01 and life-threatening SCARs (severe cutaneous adverse reactions), induced by
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52 allopurinol, it is not ethical to perform a randomised controlled study when including the care of
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54 people with gout in a pharmacogenetic study. We enrolled adults at least 18 years of age, who
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56 were not pregnant or breastfeeding and had a diagnosis of gout, chronic obstructive pulmonary
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3 disease, migraine, depression, osteoarthritis, hypertension, hyperlipidemia, atrial fibrillation,
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5 osteoporosis and/or epilepsy. Family physicians and pharmacists recruited patients and obtained
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7 saliva samples.
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9 ***Data Collection, DNA Isolation, Extraction and Genotyping***

10 Patients gave saliva samples using the Oragene DNA collection kit (DNA Genotek) in the
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12 physician's office or pharmacy, and these were transported to the laboratory by research staff,
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14 mail or floatplane.
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17 Genomic DNA was extracted using a magnetic-bead based extraction method (Ambion®
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19 MagMAX™, Applied Biosystems). Each sample was quantified with the Qubit 2.0 fluorometer
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21 (ThermoFisher Scientific) and DNA quality was assessed using the 260nm and 280nm
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23 absorbance ratio (Pure DNA: 1.8-2.0, protein contamination: <1.8, RNA contamination: >2.0).
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25 SNP and CNV genotyping was carried out by qPCR on the QuantStudio™ 12K Flex System
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27 (Applied Biosystems). A genotype report was generated and sent to the MDSS, and a copy
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29 given to the physician/pharmacist.
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33 ***Statistical Analysis***

34 The primary end point was ability to obtain and genotype samples, determined by the number
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36 of samples received in the laboratory, and the number of genetic reports generated. Samples
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38 from all patients who entered the study were evaluated. The secondary end points were yield
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40 and purity of DNA samples, ability to link results to decision support software, and use of the
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42 MDSS. All samples were tracked for linkage to the MDSS, which recorded the number of times a
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44 physician/pharmacist entered the system and for how many patients they used decision support.
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46 DNA purity was determined by the 260/280 ratio with 1.8 to 2.0 taken as pure DNA. The
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48 physicians and pharmacists were asked for their opinions on usability and effect of the MDSS on
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50 their prescribing and advice to patients. For proportions the 95% confidence interval was
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52 calculated using the Wilson Score on Open-Epi v3.03.
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Results

Two family practices on Vancouver Island, and three family practices and one pharmacy in Metro Vancouver, recruited and obtained saliva samples from a total of 191 patients. DNA was isolated from 190 samples and assessed for purity, prepared for analysis, and finally genotyped. The flow through the study is shown in Figure 2.

Of the initial saliva samples, 86.3% (164/190) were successfully genotyped after the first attempt, 7.4% (13/190) were genotyped after re-running the pharmacogenetic panel, and 1.1% (1/190) were genotyped after three or more attempts. The mean DNA concentration for all attempted extractions (n = 190) was 59.6 ng/μL (95% CI: 54.0 to 65.2). The mean 260/280 absorbance ratio of extracted DNA was 1.87 (95% CI, 1.84 to 1.91).

A second saliva sample was collected from 10 of the 12 patients that could not be genotyped. This increased the overall success rate to 99% (189/190 patients), however four of the recollected samples only generated partial reports.

Of the 185 patients with complete reports, 96.8% had at least one actionable genotype for medications included in the MDSS. Single variants were seen in 24.3% of patients, 35.1% had two variants, and 37.3% had three or more of the variants tested. The complete list of variants is shown in Table 1.

The genetic results were linked directly into the MDSS software platform and presented to healthcare professionals in a report that included the method of testing, the drugs implicated and the predicted phenotype. The healthcare professional was able to log in to the system, select their patient and then select the relevant condition. Figure 1 shows how, after completing patient biophysical information, the software displays a list of medication options. The medications are selected and dose-adjusted based on evidence-based drug-drug, drug-condition, or drug-gene interactions within the program.

The physicians, pharmacists and support staff used the software to review the patients' therapy for one or more condition and then provided feedback about the process. The software was

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3 used 236 times over a period of three months, for 11 of the 15 conditions, by the physicians and
4 pharmacists. The frequency with which specific conditions were accessed was, as expected, in
5 proportion to the most commonly seen primary care conditions. Comments from the health
6 professionals fell into three distinct themes: i) The need for instructions on entering variables; ii)
7 The belief that the system will help them to reduce inappropriate prescriptions; iii) The belief that
8 the system successfully integrates the genetic information in a way that makes it easy to
9 assimilate this information into their decision making.

18 **Interpretation**

20 The challenge of identifying the right medication for the right patient at the right time remains as
21 hard today as it has ever been. The current situation is described as a cascade of failure¹⁷ due in
22 part to the problem of multimorbidity and polypharmacy. Pharmacogenetic testing for variants
23 that are associated with alterations in drug response can help prevent adverse reactions,¹⁸ but
24 adds to the problem of increasing complexity in prescribing. The challenge for healthcare
25 professionals is how we actually undertake the process of incorporating this information in the
26 limited time available within a consultation given to deciding on, and writing a prescription.¹⁹ Pre-
27 emptive pharmacogenetic testing, ordered prior to the need for pharmacological prescription,
28 may be one approach, but the need for some sort of system to help professionals use the
29 genetic information is also necessary.²⁰ It has been suggested that to impact the level of
30 utilization of pharmacogenetic test results, pharmacogenetic testing could be ordered by the
31 pharmacist and incorporated into a medication therapy management session or comprehensive
32 medication review²¹. We demonstrated that it is possible for both family physicians and
33 pharmacists to obtain saliva samples from patients, send the samples to the laboratory, complete
34 the biophysical and laboratory data needed for a MDSS, and take decisions based on
35 individualized medication options.

55 The use of electronic medical records has increased over the last 10 years but has not been
56 associated with a drop in adverse drug events.² The use of alerts within the EMR systems when
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3 selecting a drug is not a successful strategy since they are ignored in 49% to 96% of cases,²²
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5 and the alert system is unlikely to be the most effective intervention within the time constraints of
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7 normal family practice.²³ An alternative to alerts is to build a condition-based clinical decision
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9 support system, derived from the clinical guidelines for all conditions, that provides options which
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11 are safe and effective for that patient.⁹ We built a system that starts with all the possible option
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13 pathways for treatment and results in a list of optimal, individualized drug therapy options. These
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15 drug options are already adjusted for renal and hepatic function, comorbidities, concomitant
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17 medications, and genetics. This is the first time a multi-drug, multi-condition approach has been
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19 used in primary care including both family physician offices and a pharmacy. We have
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21 demonstrated the achievability of recruitment of patients, obtaining of saliva samples, DNA
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23 extraction, genotyping, and the use of a MDSS to translate patient genotypes into condition-
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25 based medication options for the health professional and patient.
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29 The list of drugs that have associated pharmacogenetic tests includes drugs used in many
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31 conditions seen and managed in primary care such as cardiovascular conditions, hyperlipidemia
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33 and depression.²⁴ However, as yet the number of studies involving pharmacogenetic testing in
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35 primary care is very limited.²⁵ There has been some exploration of clinical decision support
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37 including genomics and providing genomic interactions as alerts,²⁶ and the largest study to date
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39 shows very significant reductions in hospitalization in those tested (71%) compared with those
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41 untested (36%).²⁷ Preliminary results from two clinical studies, one recruiting from a hospital
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43 system and the other from a long-term care facility, produced actionable genotypes for dose
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45 changes or contraindication for the patients' current medications in 24% and 50% of patients
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47 respectively.^{28,29} Given our finding that 97% of patients had at least one actionable
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49 pharmacogenetic variant, and a 5000 patient US study where 96% had actionable
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51 pharmacogenetic variants,³⁰ it is likely that future Canadian studies will demonstrate similar
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53 numbers. The high proportion of patients with actionable genotypes, coupled with the fact that
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55 11% of Canadians aged 45 to 64, and 30% of seniors aged 65 to 79, take at least five
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3 prescriptions drugs concurrently³¹ indicates that pre-emptive administration of a
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5 pharmacogenetic test has enormous potential. Pharmacogenetic testing should be part of
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7 preventive medicine; if every person is tested prior to a need for medication, when the need for
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9 medication arises there would be no need to delay medication or give medication blindly whilst
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11 waiting for a test result.

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14 The development of condition-based prescribing using an evidence-based approach has been
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16 discussed³² but this is the first time it has been used for multiple-drugs and multiple-conditions in
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18 primary care. To our knowledge, our study is the first that successfully explores the use of
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20 condition-based medication decision support that also incorporates pharmacogenetic
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22 information in community primary care.
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24 25 **Conclusion**

26 Within primary care, it is possible to collect saliva samples of sufficient quantity and quality for
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28 DNA extraction, purification and genotyping. These pharmacogenetic tests can be incorporated
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30 into a condition-based medication decision support tool that provides a list of dose-adjusted
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32 medication options which have been filtered for potential adverse drug reactions. The system
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34 was used extensively, indicating that use of a clinical decision support system with integrated
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36 data from pharmacogenetic tests may result in safer prescribing practices.
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Table 1. Frequency of alleles and diplotypes for fully genotyped patients tested in primary care (n=185).

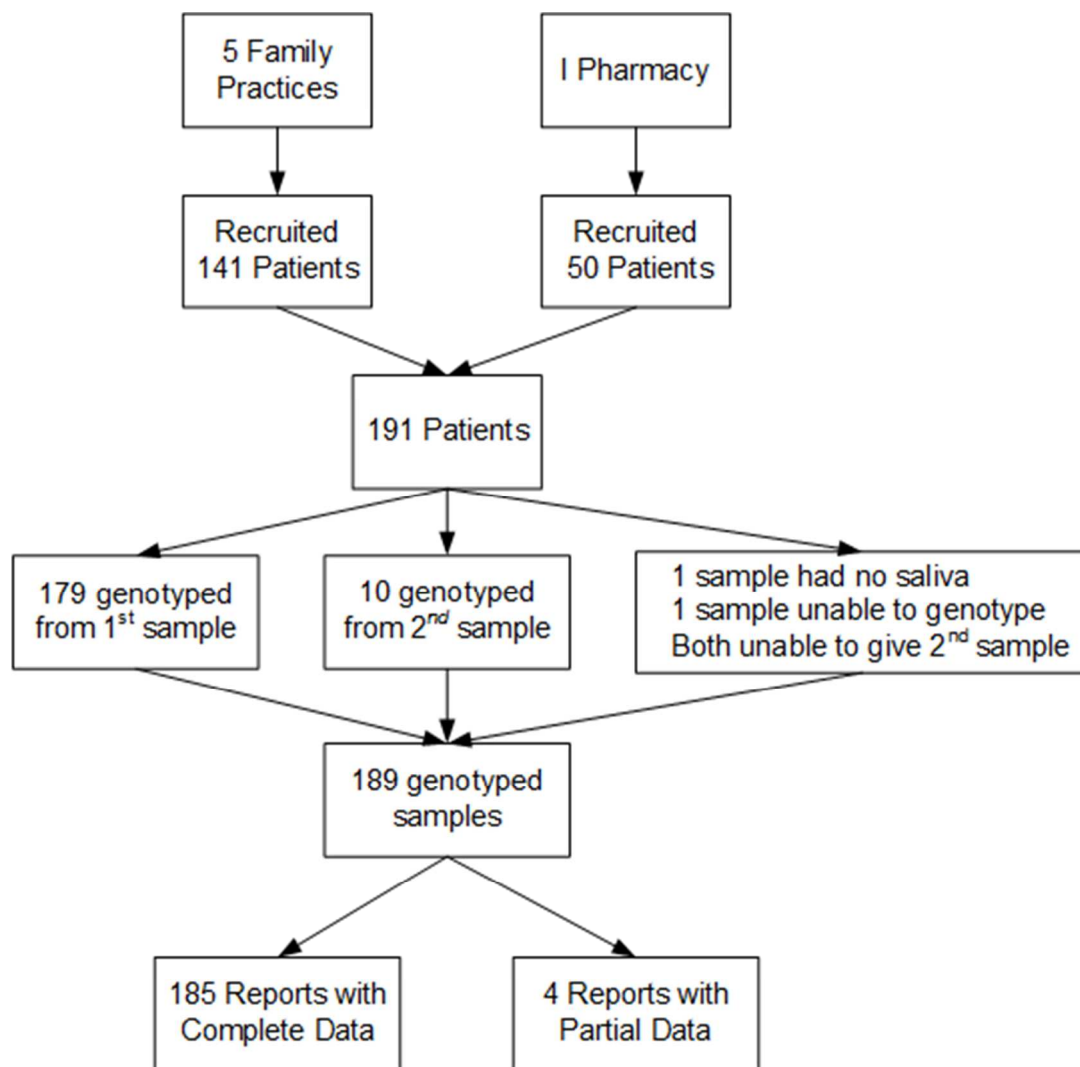
| Gene | Alleles/ Diplotypes | Frequency of Alleles/ Diplotypes | | Phenotype |
|---|---|----------------------------------|--------------|----------------------------|
| | | No. of patients | % (95%CI) | |
| SLCO1B1 rs4149056 | T/T | 132 | 71 (64-77) | Normal function |
| | T/C | 49 | 27 (21-33) | Intermediate function |
| | C/C | 4 | 2 (0-4) | Low activity |
| G6PD rs5030868 | C/C | 185 | 100 (98-100) | Normal variant |
| | C/T | 0 | 0 (0-2) | Variable activity |
| | T/T | 0 | 0 (0-2) | Low activity |
| VKORC1 rs9923231 | G/G | 38 | 21(15-27) | Normal activity |
| | G/A | 89 | 48 (41-55) | Intermediate activity |
| | A/A | 58 | 31 (25-38) | Low activity |
| HLAB *58:01 | *58:01 Negative | 176 | 95 (91-97) | Normal |
| | *58:01 Positive | 7 | 4 (2-8) | Increased hypersensitivity |
| | Undetermined | 2 | 1 (0-4) | Unknown |
| CYP2C19 | *1/*1 | 80 | 43 (36-50) | Extensive metabolizer |
| | *1/*2, *2/*17, *1/*4, *1/*8 | 55 | 30 (24-37) | Intermediate metabolizer |
| | *2/*2, *2/*3, *3/*3 | 8 | 4 (2-8) | Poor metabolizer |
| | *1/*17, *17/*17 | 42 | 23 (17-29) | Ultra-rapid metabolizer |
| CYP2C9 | *1/*1, | 118 | 64 (57-70) | Extensive metabolizer |
| | *1/*2, *1/*3, *2/*2, *2/*3 | 62 | 33 (27-41) | Intermediate metabolizer |
| | *3/*3 | 5 | 3 (1-6) | Poor metabolizer |
| CYP2D6 Codeine CPIC guidelines | *1/*1, *1, *2, *2/*2, *1/*10, (*10/*10)3N, (*2/*4)3N, *1/*17, *1/*3, *1/*4, *1/*5, *1/*41, *2/*10, *2/*4, *2/*5, *2/*9, *2/*41, *10/*10, *41/*41 | 148 | 80 (74-85) | Extensive metabolizer |
| | *3/*41, *3/*9, *4/*10, *4/*9, *4/*41, *5/*10, *5/*9 | 19 | 10 (7-15) | Intermediate metabolizer |
| | *3/*4, *4/*14A, *4/*4, *4/*5, *4/*6 | 13 | 7 (4-12) | Poor metabolizer |
| | (*1/*1)xN, (*1/*10)3N, (*1/*2)3N, (*2/*2)3N | 5 | 3 (1-6) | Ultra-rapid metabolizer |
| CYP2D6 DPWG guidelines [¶] | *1/*1, *1, *2, *1/*10, *1/*17, *1/*41, *2/*2, *2/*10, *2/*41, *2/*9, (*1/*10)3N | 94 | 51 (44-58) | Extensive metabolizer |
| | (*2/*4)3N, *1/*3, *1/*4, *1/*5, *2/*4, *2/*3, *10/*10, (*10/*10)3N, *2/*5, *41/*41, *3/*41, *3/*9, *4/*9, *4/*10, *4/*41, *5/*10, *5/*9 | 74 | 40 (33-47) | Intermediate metabolizer |
| | *3/*4, *4/*14A, *4/*4, *4/*5, *4/*6 | 13 | 7 (4-12) | Poor metabolizer |
| | (*1/*1)xN, (*1/*2)3N, (*2/*2)3N | 4 | 2 (1-5) | Ultra-rapid metabolizer |

[¶]Ambiguous genotypes DPWG guidelines CYP2D6 (*2/*4)3N and CPIC CYP2D6 (*1/*10)3N were assigned the "worst-case" phenotype.

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Figure 1. Example of Medication Decision Support System drug options for a 62-year-old patient with depression who is a CYP2C19 poor metabolizer

| TreatG [®] | |
|---|---|
| Patient | Medication Options |
| Depression not on medication | SSRI or Bupropion or Mirtazapine or Moclobemide |
| Disease Specific None of the above | Citalopram (SSRI) \$ |
| Conditions None of the above | Initial: 10 mg PO daily Maximum: 20mg PO daily Minimum titration interval: 1 week *Reduced dose due to CYP2C19 poor metabolizer for Citalopram |
| Age (years) 62 | Escitalopram (SSRI) \$ |
| Genetics - CYP2C19 Poor metabolizer | Initial: 5 mg PO daily Maximum: 20mg PO daily *Reduced initial dose due to CYP2C19 poor metabolizer for Escitalopram |
| Genetics - CYP2D6 Extensive metabolizer | Fluoxetine (SSRI) \$\$ |
| Lab: eGFR (ml/min) Value: 95 | Initial: 10-20mg PO daily Usual: 20-40 mg PO daily Maximum: 80mg PO daily |
| Lab: Creatinine Clearance (ml/min) Value: 95 | Fluvoxamine (SSRI) \$ |
| Hepatic Impairment Scale (Child-Pugh) No impairment | Initial: 50mg PO at bedtime Usual: 100-200 mg PO at bedtime (for doses > 150mg, divide BID) Maximum: 300mg PO daily |
| Current Medications None | Sertraline (SSRI) \$ |
| | Initial: 25 mg PO daily Maximum: 200mg PO daily Minimum titration interval: 1 week *Reduced initial dose due to CYP2C19 poor metabolizer for Sertraline |
| | Bupropion, SR or XL (NDRI) \$\$ |

Figure 2. Study Flow

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