

Multiphoton imaging reveals that nanosecond pulsed electric fields collapse tumor and normal vascular perfusion in human glioblastoma xenografts.

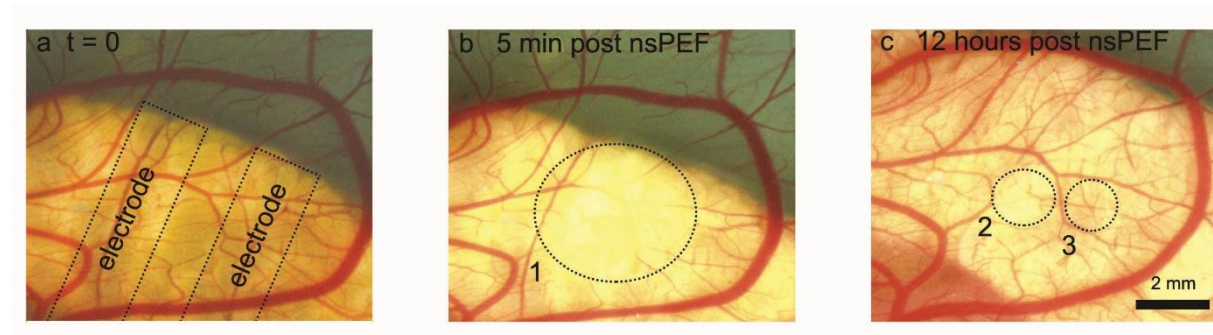
Sylvia M Bardet¹, Lynn Carr¹, Malak Soueid¹, Delia Arnaud-Cormos¹, Philippe Leveque¹
& Rodney P O'Connor¹

¹ XLIM Research Institute, UMR CNRS No 7252, University of Limoges, Faculty of Science and Techniques, 123 Avenue Albert Thomas, 87060 Limoges, France.

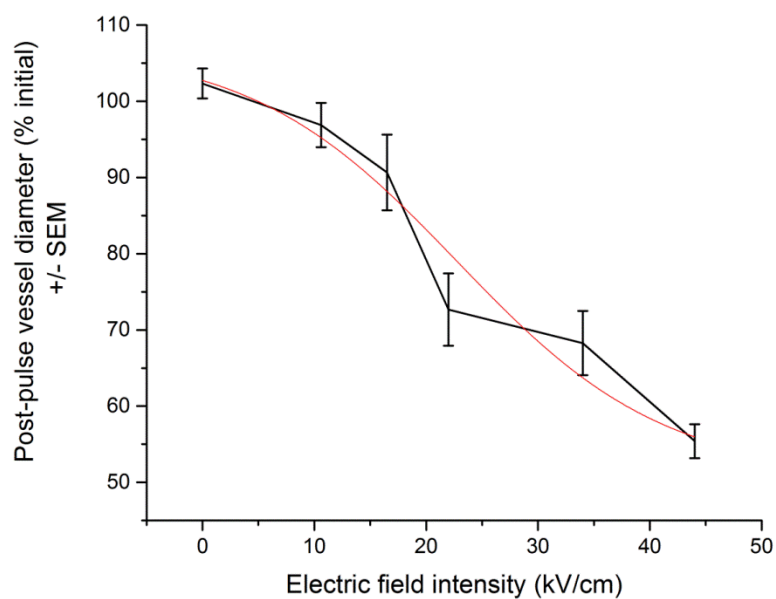
Corresponding author: Rodney P. O'Connor rodney.oconnor@xlim.fr

SUPPLEMENTARY FIGURES

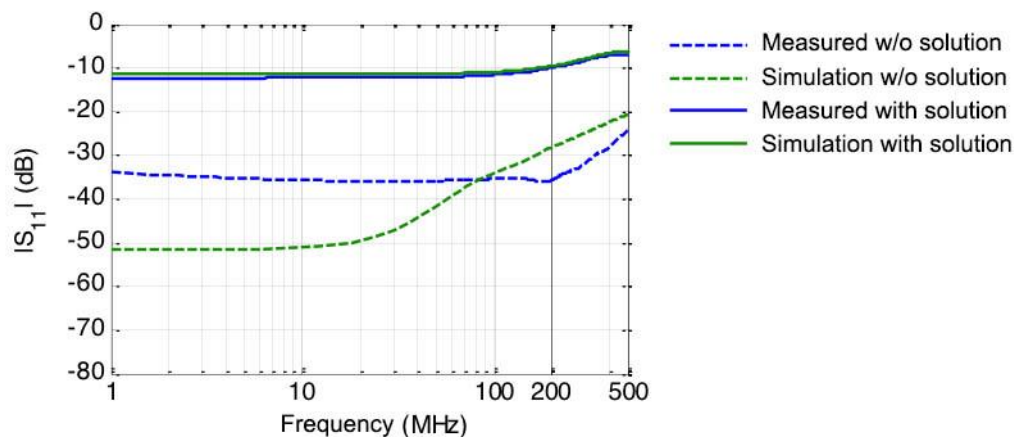
Supplementary figure 1: The nanosecond pulsed electric field effects on capillary perfusion in the CAM persist at 12h. CAM vascularization was visualized with bright field microscopy before nsPEF application (a), 5 min after (b), and at 12 hours (c). Electrodes site were drawn on (a) with dotted rectangles. The circular zone (1) on (b) highlights the clear loss of perfusion of the treated zone (large vessels and capillaries) 5 min after nsPEF treatment. Circle 2 in (c) shows an example zone where perfusion did not return in capillaries, compared with another zone (c, circle 3) where vascularization recovered after 12 hours. Scale bar in (c) = 2 mm applies to all images.



Supplementary figure 2: The dose-response relationship for the vascular effects of nsPEF was investigated with respect to electric field intensity. A total of 18 independent CAM samples were injected with Rhodamine B-dextran 70k and treated with a single nsPEF at a range of electric field intensities including 0 kV/cm (control condition with the same placement of electrodes, n=4), 10.6 kV/cm (n=2), 16.5 kV/cm (n=2), 22.5 kV/cm (n=3), 34 kV/cm (n=3) and 44kV/cm (n=4). Vessel diameter was measured and the peak decrease in vessel diameter followed a sigmoidal trend with increasing electric field intensity, as shown by the dose-response curve fit (R-Square=0.99).



Supplementary figure 3: The measured and simulated reflection coefficient curves of the nsPEF delivery electrodes. To determine the efficiency of the energy transfer between the generator and the delivery system, reflection coefficient (S_{11}) evaluation was carried out through measurements and simulations. Reflection coefficients of less than -13 dB over the 0-500 MHz frequency bandwidth (without biological solution, dotted lines) and less than -10 dB over the 0-200 MHz frequency bandwidth (with biological solution, solid lines) were obtained corresponding to a good impedance matching and energy transfer. It can be noticed that both measured and simulated results are in good agreement with each other.



Supplementary video 1 :

Multiphoton imaging of quail CAM vasculature visualized in a 3D rotating movie showing intravascular Rhodamine dextran labeled capillaries and vessels in a field of $500 \times 500 \times 200 \mu\text{m}$ (case 1).

Supplementary video 2 :

Multiphoton imaging of quail CAM vasculature visualized in a 3D rotating movie showing intravascular Rhodamine dextran labeled capillaries and vessels in a field of $500 \times 500 \times 400 \mu\text{m}$ (case 2).

Supplementary video 3 :

Multiphoton imaging of quail CAM vasculature visualized in a 4D movie (3D over time) showing intravascular Rhodamine dextran labeled capillaries and vessels in a field view of $500 \times 500 \times 400 \mu\text{m}$ (case 2) that was pulsed at $t=6$ min with a single 10 ns PEF.

Supplementary video 4 :

Multiphoton imaging of quail CAM vasculature visualized in a 3D rotating movie showing intravascular Rhodamine dextran labeled capillaries and vessels in a field of 500*500*350 μm (case 3).

Supplementary video 5 :

Multiphoton imaging of quail CAM vasculature visualized in a 4D movie (3D over time) showing intravascular Rhodamine dextran labeled capillaries and vessels in a field view of 500*500*350 μm (case 3) that was pulsed at t=6 min with a single 10 ns PEF and displayed extravascular fluorescence.

Supplementary video 6 :

Multiphoton imaging of quail CAM vasculature visualized in a 3D rotating movie showing intravascular Rhodamine dextran labeled capillaries and vessels in a field of 500*500*500 μm (case 4).

Supplementary video 7 :

Multiphoton imaging of quail CAM vasculature visualized in a 4D movie (3D over time) showing intravascular Rhodamine dextran labeled capillaries and vessels in a field view of 500*500*500 μm (case 4) that was pulsed at t=6 min with a single 10 ns PEF and displayed extravascular fluorescence.

Supplementary video 8 :

Intravascular Rhodamine B-dextran and GFP-U87 grafted on CAM were observed with multiphoton imaging in a series of image sections of the tumoral spheroid over 500*500*250 μm (case 5).

Supplementary video 9 :

Intravascular Rhodamine B-dextran and GFP-U87 grafted on CAM were observed with multiphoton imaging and shown in a 3D rotating movie over 500*500*200 μm (case 6).

Supplementary video 10 :

Intravascular Rhodamine B-dextran and GFP-U87 grafted on CAM were observed with multiphoton imaging and shown in a 3D rotating movie over 500*500*200 μm (case 7).

Supplementary video 11 :

Intravascular Rhodamine B-dextran and GFP-U87 migrating cells on CAM were observed with multiphoton imaging and shown in a 3D rotating movie over 500*500*200 μm (case 8).