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NSUN3 and ABH1 modify the wobble position of mttRNA^{Met} to expand codon recognition in mitochondrial translation

Sara Haag, Katherine E. Sloan, Namit Ranjan, Ahmed S. Warda, Jens Kretschmer, Charlotte Blessing, Benedikt Hübner, Jan Seikowski, Sven Dennerlein, Peter Rehling, Marina V. Rodnina, Claudia Höbartner and Markus T. Bohnsack

Corresponding authors: Markus T. Bohnsack, University Medical Center, and Claudia Höbartner, Institute for Organic and Biomolecular Chemistry, Georg-August-University Göttingen

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Reporting Checklist For Life Sciences Articles (Rev. July 2015)

This checklist is used to ensure good reporting standards and to improve the reproducibility of published results. These guidelines are consistent with the Principles and Guidelines for Reporting Preclinical Research issued by the NIH in 2014. Please follow the journal's authorship guidelines in preparing your manuscript

A- Figures

- The data shown in figures should satisfy the following conditions:

 the data were obtained and processed according to the field's best practice and are presented to reflect the results of the experiments in an accurate and unbiased manner.
 - figure panels include only data points, measurements or observations that can be compared to each other in a scientifically
 - meaningful way.

 graphs include clearly labeled error bars for independent experiments and sample sizes. Unless justified, error bars should not be shown for technical replicates.
 - if n< 5, the individual data points from each experiment should be plotted and any statistical test employed should be justified
 - → Source Data should be included to report the data underlying graphs. Please follow the guidelines set out in the author ship guidelines on Data Presentation

2. Captions

Each figure caption should contain the following information, for each panel where they are relevant:

- a specification of the experimental system investigated (eg cell line, species name).

- a specimation on the Experimental system investigated teg cut mine, species indire).
 the assay(s) and method(s) used to carry out the reported observations and measurements
 an explicit mention of the biological and chemical entity(ies) that are being measured.
 an explicit mention of the biological and chemical entity(ies) that are altered/varied/perturbed in a controlled manner.

- the exact sample size (n) for each experimental group/condition, given as a number, not a range;
 a description of the sample collection allowing the reader to understand whether the samples represent technical or biological replicates (including how many animals, litters, cultures, etc.).
 a statement of how many times the experiment shown was independently replicated in the laboratory.
 definitions of statistical methods and measures:
 common tests, such as t-test (please specify whether paired vs. unpaired), simple x2 tests, Wilcoxon and Mann-Whitney tests, can be unambiguously identified by name only, but more complex techniques should be described in the methods
 - · are tests one-sided or two-sided?

 - are tests one-sided or two-sided? are there adjustments for multiple comparisons? exact statistical test results, e.g., P values = x but not P values < x; definition of 'center values' as median or average;

 - definition of error bars as s.d. or s.e.m.

Any descriptions too long for the figure legend should be included in the methods section and/or with the source data

Please ensure that the answers to the following questions are reported in the manuscript itself. We encourage you to include a specific subsection in the methods section for statistics, reagents, animal models and human subjects.

he pink boxes below, provide the page number(s) of the manuscript draft or figure legend(s) where the ormation can be located. Every question should be answered. If the question is not relevant to your research,

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B- Statistics and general methods

1.a. How was the sample size chosen to ensure adequate power to detect a pre-specified effect size?	The sample size was chosen based on long-term experience with biochemical analyses in the participating institutes. In general, we chose to perform at least three biological replicates of each experiment. N/A
1.b. For animal studies, include a statement about sample size estimate even if no statistical methods were used.	
2. Describe inclusion/exclusion criteria if samples or animals were excluded from the analysis. Were the criteria preestablished?	The independent experiments were analysed and the data included in the statistical analyses. If experiments failed for technical reasons or could not be analysed (e.g. if the resolution of a gel was not sufficient for quantification of bands) results were excluded.
3. Were any steps taken to minimize the effects of subjective bias when allocating animals/samples to treatment (e.g. randomization procedure)? If yes, please describe.	Datasets were generally analysed independently by different scientists to avoid subjective analysis.
For animal studies, include a statement about randomization even if no randomization was used.	N/A
4.a. Were any steps taken to minimize the effects of subjective bias during group allocation or/and when assessing results (e.g. blinding of the investigator)? If yes please describe.	N/A
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5. For every figure, are statistical tests justified as appropriate?	Statistical analyses were chosen appropriately to the best of our knowledge.
Do the data meet the assumptions of the tests (e.g., normal distribution)? Describe any methods used to assess it.	T-test was applied as indicated in the manuscript.
Is there an estimate of variation within each group of data?	Standard deviation and standard error of the mean were calculated as indicated.
Is the variance similar between the groups that are being statistically compared?	See above.

6. To show that antibodies were profiled for use in the system under study (assay and species), provide a citation, catalog	The origin of commercial antibodies, the corresponding catalogue numbers or references to
number and/or clone number, supplementary information or reference to an antibody validation profile. e.g.,	previous characterisations are given in the manuscript.
Antibodypedia (see link list at top right), 1DegreeBio (see link list at top right).	
7. Identify the source of cell lines and report if they were recently authenticated (e.g., by STR profiling) and tested for	HeLa CCL2 cells originated from ATCC and the Flp-In T-REx 293 cellline was aquired from Life
mycoplasma contamination.	Technologies (Thermo Scientific). Cells are tested regularly for their identity and are confirmed as
	mycoplasma free using a commercial kit.

D- Animal Models

 Report species, strain, gender, age of animals and genetic modification status where applicable. Please detail housing and husbandry conditions and the source of animals. 	N/A
 For experiments involving live vertebrates, include a statement of compliance with ethical regulations and identify the committee(s) approving the experiments. 	N/A
10. We recommend consulting the ARRIVE guidelines (see link list at top right) (PLoS Biol. 8(6), e1000412, 2010) to ensure that other relevant aspects of animal studies are adequately reported. See author guidelines, under 'Reporting Guidelines'. See also: NIH (see link list at top right) and MRC (see link list at top right) recommendations. Please confirm compliance	N/A

E- Human Subjects

11. Identify the committee(s) approving the study protocol.	N/A
12. Include a statement confirming that informed consent was obtained from all subjects and that the experiments conformed to the principles set out in the WMA Declaration of Helsinki and the Department of Health and Human Services Belmont Report.	N/A
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F- Data Accessibility

18. Provide accession codes for deposited data. See author guidelines, under 'Data Deposition'.	The UV and 5-azacytidine crosslinking and analysis of cDNA data are being deposited in the Gene
	Expression Omnibus database. The data base entry has been initiated and the accession codes will
Data deposition in a public repository is mandatory for:	be supplied when available.
a. Protein, DNA and RNA sequences	
b. Macromolecular structures	
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d. Functional genomics data	
e. Proteomics and molecular interactions	
19. Deposition is strongly recommended for any datasets that are central and integral to the study; please consider the	See above.
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datasets in the manuscript as a Supplementary Document (see author guidelines under 'Expanded View' or in	
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20. Access to human clinical and genomic datasets should be provided with as few restrictions as possible while	N/A
respecting ethical obligations to the patients and relevant medical and legal issues. If practically possible and compatible	
with the individual consent agreement used in the study, such data should be deposited in one of the major public access-	
controlled repositories such as dbGAP (see link list at top right) or EGA (see link list at top right).	
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Referenced Data	
Huang J, Brown AF, Lei M (2012). Crystal structure of the TRBD domain of TERT and the CR4/5 of TR. Protein Data Bank	
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format (SBML, CellML) should be used instead of scripts (e.g. MATLAB). Authors are strongly encouraged to follow the	
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at top right) or JWS Online (see link list at top right). If computer source code is provided with the paper, it should be	
deposited in a public repository or included in supplementary information.	

G- Dual use research of concern

23. Could your study fall under dual use research restrictions? Please check biosecurity documents (see link list at top	N/A
right) and list of select agents and toxins (APHIS/CDC) (see link list at top right). According to our biosecurity guidelines,	
provide a statement only if it could.	

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