

Supplementary Appendix

This appendix has been provided by the authors to give readers additional information about their work.

Supplement to: Cohen MS, Chen YQ, McCauley M, et al. Antiretroviral therapy for the prevention of HIV-1 transmission. *N Engl J Med* 2016;375:830-9. DOI: 10.1056/NEJMoa1600693

(PDF updated September 1, 2016.)

**Antiretroviral Treatment for Prevention of HIV Transmission:
The Final Results of the HIV Prevention Trials Network (HPTN) 052 Trial**

Supplementary Appendix

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Acknowledgements

Acknowledgements for Each Participating Site and Other Collaborating Institutions and Individuals.

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AMERICAS

Rio de Janeiro, Brazil:

Instituto de Pesquisa Clinica Evandro Chaga (IPEC): Valdilea Goncalves Veloso, Ruth Khalili Friedman, Sandra Wagner Cardoso, Guilherme Amaral Calvet, Maria Pia Diniz Ribeiro, Isabel C. Tavares, Maria R. Rocha, Nilo Martinez Fernandes, Ronaldo Ismerio Moreira, Margarete Paiva, Sandra Filgueiras, Daniel Waite, Mariza Goncalves Morgado, Lidiane Tuler, Ingeborg Georg, Ivan Neves Jr., Angela C. Andrade, Lucia Sena, Thiago Torres, Marília Santini de Oliveira, Brenda Hoagland, Juan C. Raxach, Cristina Pimenta, Valeria Ribeiro, Jorge Nunes, Ludmila da Silva Alves, Ricardo H. Freitas, Lucia H. Cardoso de Souza, Sandro Coutinho Nazer da Costa, Soraia Santana de Moura, Tania Krstic, Marcella Feitosa, Flavia Lessa, Luiz Ricardo Siqueira Camacho

Hospital Geral de Nova Iguacu (HGNI): Tania Brum, Flavio Bustorff, Maria Isabel do Nascimento, Lara Somma Portela, Aline Ramalho, Ana Claudia Rodrigues, Cintia Lopes da Silva, Tatiana Muniz de Chã

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Porto Alegre, Brazil: Marineide G. de Melo, Rita A. Lira, Kelin R. Zabtoski, Andréa C. Castro, Rui Flores, Melina Grudzinski, André L. da Silva, Dimas A. Kliemann, Cláudio M. Campello, Ivete C. Canti, Elizabeth S. Magalhães, Consuelo F. Perez, Mara L. Silveira, Julio Barris, Marcelo E. Almeida, Maria C. Seter, Maria L. Turella, Carmen L. Fernandes, Leda Curra, Mariana R. Simon, Salete M. Zabtoski, Inelva Miotto, Fabria Chiarani, Deise T. Ramos, Claudia C. Gottert, Magnus C. de Melo, Roberta F. dos Santos

Boston, Massachusetts, USA (site closed prior to 2007): Marcy Gelman, Ana Maldonado, Robbie Singal, Steven Safren, Rodney Vanderwarker

ASIA

Chennai, India: Aylur Kailasom Srikrishnan, Jabin Sharma, Easter Thamburaj, Jeeva Arumugham, Narayanan Govindarajan, Poongulalli Selvamuthu, Faith Beulah, Syed Iqbal

Pune, India: Ramesh Paranjape, Arun Risbud, Srikanth Tripathy, Raman Gangakhedkar, Seema Sahay, Smita Kulkarni, Manisha Ghate, Madhuri Thakar, Sampada Dhayarkar, Arati Mane, Varsha Kale, Sangita Kulkarni, Bharati Mahajan, Radhika Brahme, Neelam Joglekar, Neeta Pradhan, Sumitra Krishnan, Swapna Kanitkar, Rajeshwari Deshmukh, Archana Beri, Usha Katti, Anuradha Koli, Mallika Alexander, Rewa Kohli, Swapna Deshpande, Asmita Gaikwad, Jyoti Pawar, Prafulla Patil, Ashwini Kokil, Yogesh Nehete, Pratima Sheth, Priyanka Tupekar, Prajakta Patil, Prasad Kulkarni, Yogesh Wagh, Savita Mawal, Prajakta Dhamne, Mahesh Kharat, Aparna Parkhe, Ayesha Momin, Sachin Jadhav, Mufid Baig, Vikram Solas, Ratnaprabha Bihade, Latika Karve, Keshav Gade, Narayan Panchal, Urmila Ghodke, Harshad Nalgirkar, Sunil Gaikwad, Lalit Patil, Swati Jivane, Megha Mamulwar, Deepak Bangar, Rajesh Yadav, Manoj Gorade, Vijay Kad, Ganesh

Palhade, Bipin Bangale, Sonal Shindekar, Vandana Bankar, Kiran Lakhwani, Suchitra Ganeshacharya, Chaitrali Patil, Ashwini Jagdale, Naveen Satyanna, Ashwini Patil, Asha Francis, Vijay Chauware, Sachin Kale, Rohini Bingewar, Ruparani Gujar, Vikas Mallav, Manisha Bhandarkar, Vaishali Chimanpure, Shubhada Mankar, Shubhada Deshpande, Seema Nair, Gauri Vaidya, Deepak More, Anjali Panchanadikar, Kavita Pardeshi, Chetan More, Kishore Kumar, Shubhangi Navlakha, Vijaya Kulkarni, Sudhakar Wankhede, Dhananjay Dadke, Madhura Nene, Sanjay Kulkarni, Sanjay Mehendale, Swarali Kurle, Abhijit Kadam, Anjali Joglekar, Shraddha Bapat, Mansa Angadi, Prachi Athawale, Nilima Lokhande, Gururaj Deshpande, Dhanwanti Inamdar, Mercy Samuel, Yogesh Daware, Ranjana Mundhe, Shweta Chidrawar, Bharati Mahajan, Sushma Jadhav, Chetan More, Vijaya Kulkarni, Ramesh Deoda, Kumar Vaidya, Shilpa Kakru, Pranali Kulal, Pallavi Raut, Ritesh Desai, Nikita Ratnaparakhi, Laxmikant Dudhmal, Sachin Jadhav, Sarita Goli, Mahalaxmi Mayekar, Ashwini Gaikwad, Nitin Tayade, Iranna Mashal, Ujjawala Ghule, Vaibhavi Bodhe, Anjum Shaikh, Sheetal Ghule, Ipsita Choudhari, Devidas Chaturbhuj, Prafulla Lakhare, Nitin Hingankar, Madhuri Chandane, Smita Thorat, Nawaj Shaikh, Winston Umakanth, Mukund Vats, Asia Luwang, Balaji Deshmukh, Jayram Andil, Priyanka Khopkar

Chiang Mai, Thailand: Voravit Suwanvanichkij, Cholticha Ruangyuttikarn, Nuntisa Chotirosniramit, Louise Walshe, Nicole Simmons, Lara Johnson, Marisa Guptarak, Clevetta Chandler, H. Peter Lange, Chanidapa Prasarakkee, Thira Sirisanthana, Kriengkrai Srithanaviboonchai, Voravit Suwanvanichkij, Natthapol Kosashunhanan, Sunida Thetket, Patcharaphan Sugandhavesa, Taweewat Supindham, Kanokporn Chaiklang, Sineenart Nimsakul, Wilawan Chaikan, Thanyalak Thongphan, Saowalak Sarachai, Sontiya Mueanapai, Napha Panyo, Supatra Pookmanee, Boonlure Pruenglampoo, Wipada Cheewawat, Antika Wongthane, Kittipong Rungruengthanakit, Rassamee Keawvichit, Kanlaya Wongworapat, Piyathida Sroysuwan, Rojana Srichan, Boonyarat Puisaeng, Nataporn Kosachunhanan, Veruree Manoyos, Supaporn Sirikunpun, Nittaya Chuenchop, Boonyarat Puisaeng, Niranporn Jaikuar, Praphapin Suriyasorgpi, Kantaphat Dachapratoomwan, Wasun Chanchai, Darika Chittpramodya, Suthathip Wongsrithep, Karnjana Chairungsri, Pimpaka Puangpotha, Tipawal Petthed, Kunnika Jungasathit, Kulthida Chaikul, Waraporn Pasawad, Jiraporn Yamano, Nattanicha Leelasuksaree, Chiraphorn Kaewkosaba, Nattanun Suwannamas, Chamaiporn Naprom, Panida Yodkeeree, Lar Chandee, Sirikwan Dokuta, Chansom Pantip, Panudda Sothanapaisan, Jeitsada Keitkarn, Warunee Jit-Aree, Chayanid Maneechai, Kannika Boursuk, Wirat Niwatananun, Pranee Khad-Umong, Supachai Sakkhachornphop, Parinya Jongpaijitsakul, Nongluck Kabyoy, Taweeluk Vannarit, Charatdao Bunthi, Chaisiri Angkurawaranon, Pranee Sakkhachornphop, Quanhathai Kaewpoowat, Prachern Palanan, Eaksit Chaipin, Metaporn Sompong, Patcharaporn Wongphu-nga, Nunthakarn Saenrak, Pannachart Manop, Ratchanit Chaiban, Saitong Sirinam, Atthawit Khudcum, Aonsutee Jannim, Rattana Kunnapa, Yuttapong Tammachai, Sunisa Butphet, Atita Panyathep, Pachern Putsyainunt, Satitpong Nunjai, Jarun Ontakrai, Paweena Khamdam, Manoo Panyamang, Kadsarin Chantan, Chatsuda Auchiang, Walailuk Hanterdsith, Wonpen Prasertwitayakij, Kesinee Treamrangsee, Darin Ruanpeng, Wathee Sitthi, Patcharaporn Siwilai

AFRICA

Gaborone, Botswana: Priti Dusara, Antonia Bunga, Emily Makunike, Akeem O. Salawu, Sikhulile Moyo, Phibieon Mangwendeza, Chishamiso Mudenyanga, Motswedi Anderson, SheronDzoro, Norah Mawoko, Botshelo Molebedi, Toro Oikantswe, Banno Moorad, Maitseo Malamba, Edwin Mogaetsho, Georinah Modise, Thapelo Mmolawa, Gaone Retshabile, Erik Widenfelt, Karabo Motsisi, Tsholofelo Tsomele, Ernest Moseki, Wellington Mongwa, Masego Kgafela, Chandapiwa Motsamai, Sarah Masole, Martha Moiforay, Kgomotsego Madome, Simon Masopa, Nonvula Sifiwa, Galetlwaelwe Molapisi, Terrence Mohammed, Lucy Mupfumi, Tumalano Sekoto, Obonwe Pule, Thuto Ralegoreng, Boitshepo Sankoloba, Victoria Maselwa

Kisumu, Kenya: Victor Akelo, Bob Chen, Debra Gust, Richard Lando, Kayla Laserson, Charles Lebaron, Beatrice Nyagol, Erick Ondieki, Clement Zeh, Arthur Ogendo, Raymond Goldstine, John Vulule, Victor Mudhune, Elizabeth Ayuo, Anne Gumbe, Vitalis Sewe, Boaz Oyaro, Wairimu Chege, Katrina Kretsinger, Janet Adhiambo, Kevin Achola, Eunice Anyiego, Eucabeth Awuonda, Emily Kerubo, Erica Mimba, Jean Muhanji, Richard Ndivo, Hilary Ngeno, George Nyamao, Mary Nyikuri, Lucy Ochieng, Sylvia Odhiambo, Evans Odipo, Eudia Odum, Phoebe Okola, Benard Okomo, Kenneth Ondenge, George O Ouma, Winnie Ouma, Elizabeth Rambara, Alice Were, Anne Adegga, Frank Angira, Nancy Atieno, Elizabeth Ogutu, Kennedy Imbuki, Abraham Katana, Phylis Mboi, Eleanor McLellan-Lemal, Dancun Okal, John Okanda, Isaiah Okello, Fred Omondi, Tereza Omoro, Pauline Ongwena, Kenneth Onyango, Valarie Opollo, Ruth Opuro, Gloria Orimba, Jecinter Oruko, Joseph Osoga, Fredrick Otieno, Risper Oyaa, Wycliff Akombi, Benson Kesa, Nick Obonyo, Wycliff Ochieng, Caleb Odhiambo, Morris Odongo, William Oremo, Wycliff Ouma, Steve Owoko

Blantyre, Malawi: Faustine Matchere, Nicole Carpenetti, Newton Kumwenda, Mulinda Nyirenda, Fatima Zulu, Linley Seyama, Ben Kalonga

Lilongwe, Malawi: Madalitso Maliwichi, Bertha Maseko, Felix Namalueso, Wiza Kumwenda, Esnath Msowoya Mkandawire, Lucy Kamalizeni, Ida Shumba, Francis Martinson, Robert Krysiak, Dorothy Sichali, George Joaki, Thokozani Nkhalamba, Noel Mumba, Debbie Kamwendo, Jean Tauzie, Franklin Kilembe, Nasi Saukila, Cecilia Kanyama, Gladys Phiri, Lebah Lugalia, Steve Mponda, Agnes Moses, Gerald Tegha, Nkhafwire Mkandawire, Lameck Chinula, Lydda Kandikole, Geoffrey Singini, Tiwonge Kumwenda, Fred Chimzimu, Limbanazo Mindiera, Emma Kachipapa, Wilberforce Mhango, Tchangani Tembo, Lucy Dzama, Innocent Mofolo, Shireen Kharodia, Enock Gamah, Arthur Sungitsa, Chimwemwe Mphande, Charity Potani, Idah Mshali, Henry Eliya, Sarah Chinyama, Egnat Katengeza, Lawrence Nkhwazi, Doreen Kanyika, Chodziwadziwa Kabudula, Tionge Kamvaunamwali, Chiyembekezo Chafuwa, Mwai Chipeta, Allan Jumbe, Alfred Mwanyimbo, Khama Mita, Stanley Kulapani, Kamnkhwani Mtanthira, Dalitso Mzinganjira, Frank Kumbanga, Lameck Gondwe, Florence Lwanda, Omega Banda, Titha Dzowela, Tapiwa Tembo, Dan Namarika, Fredreck Kachiponde, Martha Juma, Mary Chindebv, Roseby Kazembe, Esther Mathiya, Patience Yamba, Christine Chabwera, Felluna Chauwa, Esnath Mtika, Nyanyiwe Mbeye, Louisa Fiacco, Samuel Kamanga, David Chilongozi, Harriet Chanza, Allan Jumbe, Norah Chikhungu, Zane Ramdas, Linga Munthali, Bertha Limburo, Regina Mwausegha, Sophie Mtombosola, Chitsanzi Gadi, Maganizo Majawa, Jacob Phulusa, Bessings Bandawe, Agnes Gumbo, Angella Mwaipape, Hanna Stambuli, Lusungu Msumba, Clement Mapanje, Kenneth Kasanbana, Laston Kayuni, Chimwemwe Kachiwaya, Pokiwe Mwangomba, Towera Banda, Ackim Sankhani, Amanda Varela, Maggie Chigwenembe

Johannesburg, South Africa: Sharlaa Badal-Faesens, Francesca Conradie, Veronica Graham, Jemina Pinkie Thebe, Martha Ntshakala, Mirriam Manamathela, Jabulane Masimula, Fred Phakathi. Keagile Komane, Thembi Kubheka, Marlene Knight, Jenny Baines, Mohammed Rassool, Prudence Ive, Thando Mwelase, Nkuli Mashabane, Desiree van Amsterdam, Mamoloko Mashamaite, Karla Mellet, Belinda Alport, Faizel Laher

Soweto, South Africa: Sarishen Govender, Puleng Dhlamini, Ntombiyenkosi Radebe, Rebecca Lenkokile, Anusha Nana, Carita Marx, Charlene Conradie, Gugulethu Tshabalala, Miriam Montso, Mpho Motlamelle, Noxolo Dyalom, Peter Mafela, Thandekile Essien, Theogene Nshimiyimana, Guy de Bruyn, Banningi Mkhize, Ruth Motlafi, Jesne Kistan

Harare, Zimbabwe: Nehemiah Nhando, Jimijika Batani, Miriam Njaya, Misai Hukuimwe, Patricia F. Mandima, Vernon T. Murenje, Tendayi D. Mutungamiri, Lawrence T. Matanhike, Lucia Chirongoma, Patience N. Ruwende, Mary N. Tichareva, Bevelyn S. Muhwati, Thandiwe H. Chirenda, Felina Mhangami, Elizabeth S Magada, Monica Nyamhuka, Jester Makwara, Beauty Nyamayaro, Loveness Mugari, Mary Manyara, Ernest Chimuka, Nancy Jokonya, Jessie Musundire, Godfrey Matimba, Abigail Mutsinze, Gilton Kadziyanike, Sylvia Manomano, Cleopatra Langa, Christina

Maluwa, Wilfred T. Gurupira, Thembelihle Bafana, Evah Ncube, Alfred Gomo, Marshall Munjoma, Natsai Makanza, Fiona Mtisis, Collen Pamire, Solomon Mashinga, Jacob B. Kagona, Mugove Chahwanda, Fungai Maguramhinga, Memory Chikosha, Violet Mandioma

HPTN Leadership and Operations Center (FHI 360): Jacqueline Talley, Phaedrea Watkins, Jonathan Lucas, Rhonda White, Cheryl Cokley, Nirupama Sista, Melissa Allen, Kathy Hinson, Ward Cates, Timothy Mastro, Nancy Lamson, Gray Davis, Carolyn Yanavich, Andrea Jennings, Michelle Robinson

Statistical Center for HIV/AIDS Research & Prevention (SCHARP): Leslie Cottle, Jami Moksness, Maija Anderson, Sue Tracy-Waisanen, Debbie Lands, Stacie Kentop, Laura Robins-Morris, San-San Ou, Xin Li, Ben Masse, Deborah Donnell, Jing Wang, Kate Ostbye

HPTN Laboratory Center, Johns Hopkins Univ. School of Medicine, Baltimore, MD: Craig Hendrix, Charlotte Gaydos, Thomas Quinn*, Matthew Sievers, and staff of the HPTN Laboratory Center (*Dr. Quinn is also affiliated with the Laboratory of Immunoregulation, NIAID, NIH, Bethesda, MD). The following investigators from other institutions assisted the HPTN Laboratory Center with characterization of partner infections in HPTN 052: Stephen Porcella, Craig Martens, and Daniel Bruno (Genomics Unit, Research Technologies Section, Rocky Mountain Laboratories, DIR, NIAID, NIH, Hamilton, MT); Ronald Swanstrom, Li-Hua Ping, and Elena Dukhovlina (Univ. of North Carolina at Chapel Hill, Chapel Hill, NC); San-San Ou (Statistical Center for HIV Research and Prevention, Seattle, WA); and James Hughes (Dept. of Biostatistics, Univ. of Washington, Seattle, WA).

Additional Acknowledgements: Amita Gupta (Johns Hopkins University), Robert Bollinger (Johns Hopkins University)

Determination of the linkage status of partner infections in HPTN 052.

This file presents an overview of the analysis of linkage status of partner infections in HPTN 052. The methods used for this analysis were published in detail in a previous report, which also includes the results of linkage analysis that were available in May 2011, prior to release of interim study findings (Eshleman, et al. Analysis of genetic linkage of HIV from couples enrolled in the HIV Prevention Trials Network 052 trial. *J Infect Dis.* 2011; 204:1918-1926). The results of the linkage analysis for the entire trial period (through May, 2015) and additional results related to characterization of partner infections were presented in July, 2015 at the 8th IAS Conference on HIV Pathogenesis, Treatment, and Prevention in Vancouver, Canada (Eshleman, et al. Treatment as Prevention: Characterization of partner infections in the HIV Prevention Trials Network 052 trial, Abstract MOAC0106LB).

Methods used to determine the linkage status

Seventy-eight partner infections were observed in the HPTN 052 trial. The three methods were used to assess genetic linkage of HIV from index-partner pairs (see below). The classification “linked” indicates that the index was the likely source of the partner’s infection.

(1) Phylogenetic analysis of HIV *pol* region sequences obtained by population (consensus) sequencing

Linkage analysis was performed using plasma samples from partners who acquired HIV infection during the HPTN 052 trial and the corresponding HIV-infected index participants (index-partner pairs). In most cases, two samples collected on different dates were analyzed for each participant; in some cases, results were only obtained for one sample (e.g., in cases where only one HIV-positive sample was collected from the partner after HIV infection; only one sample from a participant had a viral load sufficient for analysis; or one sample failed analysis). Plasma samples were also analyzed from randomly-selected index participants (control samples); 10 control samples were analyzed from each site, with the following exceptions: one set of 10 control samples was included for the three sites in Brazil; control samples were not included for the site in the United States due to low enrollment.

The ViroSeq HIV-1 Genotyping System (Celera, Alameda, CA) was used to generate HIV *pol* sequences. This system provides population (consensus) sequences that encode HIV protease amino acids 1-99 and HIV reverse transcriptase amino acids 1-335. Alternate primers were used for selected samples if the sample failed to amplify using the primers provided with the ViroSeq kit. *Pol* region sequences from index-partner pairs and local controls were aligned using MegAlign v5.07 with the Clustal W method. Sequence distances were calculated using DNADist. Phylogenetic trees and bootstrap values were generated using PHYLIP (Neighbor-Joining and Consense); for partner infections reported in 2011, results from this analysis were compared to those obtained using other phylogenetic methods. MegAlign was used to generate phylogenetic trees for study sites by geographic region; these trees included sequences from index-partner pairs, local control sequences (matched for HIV subtype), and reference sequences. Additional information about the methods used for this part of the linkage analysis and a representative phylogenetic tree are presented in Eshleman et al, 2011, *Journal of Infectious Diseases* 204:1918-1926. A provisional linkage assignment was based on this analysis; index-partner pairs were provisionally classified as linked if all of the corresponding sequences clustered together on a single monophyletic branch with a high bootstrap value.

(2) Bayesian analysis of the genetic distances between *pol* sequences

The probability of linkage was assessed using an algorithm based on Bayes theorem. These methods were used to compare the genetic similarity of HIV *pol* sequences obtained using the ViroSeq system (see above). The genetic similarity between sequence pairs was determined using MegAlign v5.07. Genetic similarity values were compared for the following sequence pairs (all possible combinations): (a) sequences obtained for one individual from samples collected on different dates, (b) sequences obtained for unrelated study participants (other index participants and local control sequences from the same geographic region or study site), and (c) sequences obtained for index-partner pairs. The analysis assumed that paired sequences from unrelated index participants were not linked, and that the genetic similarity of sequences from index-partner pairs was similar to the genetic similarity of two sequences from the same individual. Bayes theorem was used to determine the probability of linkage between sequences from index-partner pairs. Additional information about the methods used for this part of the linkage analysis and plots showing densities of similarities for linked and unlinked events are presented in Eshleman et al, 2011, Journal of Infectious Diseases 204:1918-1926, and in Supplemental File 1 in that publication. Index-partner pairs were provisionally characterized as linked by the Bayesian analysis if they had a linkage probability of ≥ 0.5 for any pair of sequences. Index-partner pairs were provisionally characterized as unlinked by the Bayesian analysis if they had a linkage probability of ≤ 0.5 for all pairs of sequences.

(3) Phylogenetic analysis of HIV *env* region sequences obtained by next generation sequencing (NGS, selected samples)

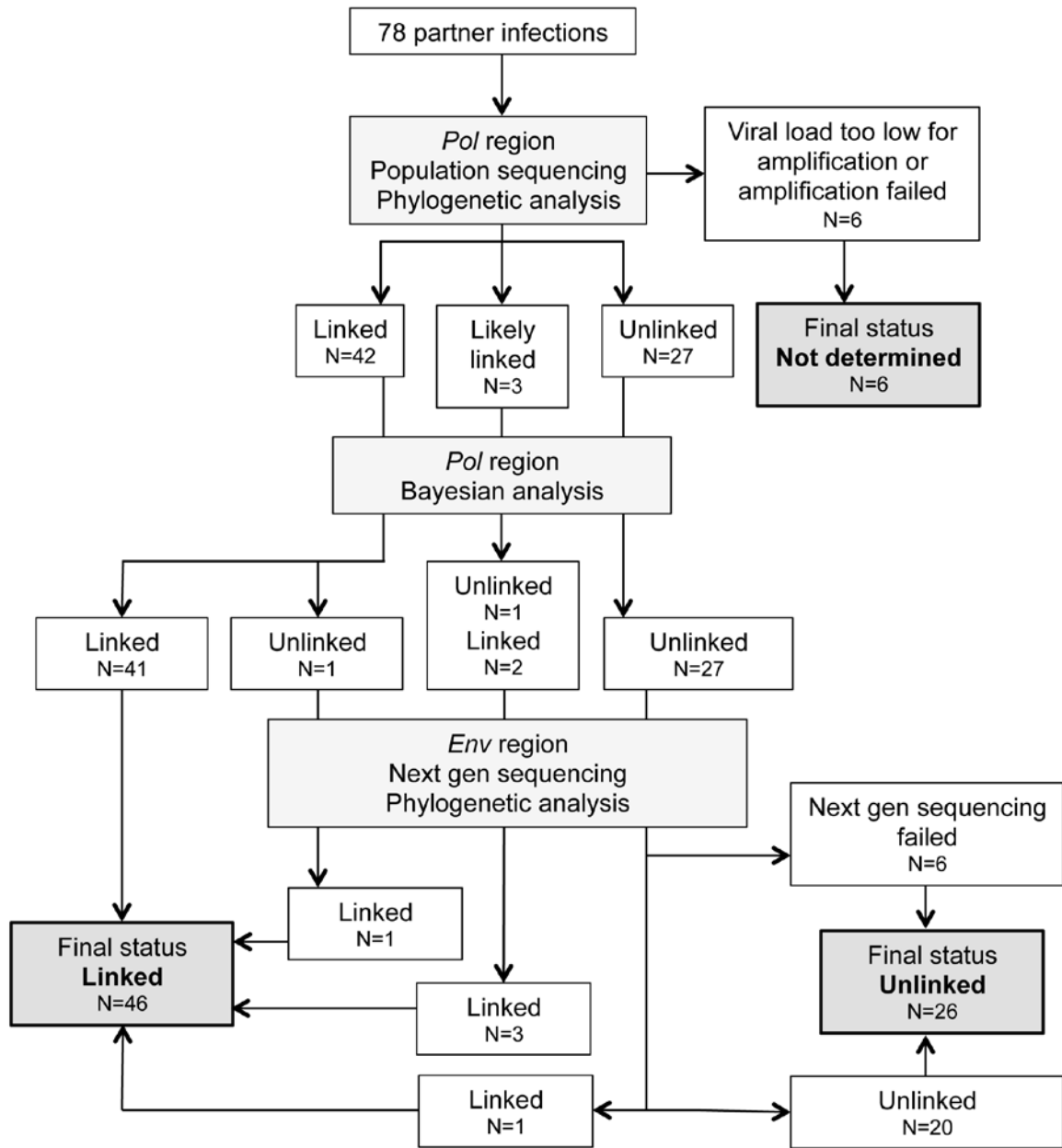
Index-partner pairs that were provisionally characterized as linked by both phylogenetic and Bayesian analysis of *pol* region sequences were classified as linked and were not analyzed further. Additional analysis using NGS was performed for the remaining index-partner pairs. NGS was performed for a region of gp41. The gp41 region was amplified using a combined reverse transcription / nested polymerase chain reaction (PCR). This was followed by a nested PCR that included using primers with DNA barcodes that identified each sample. Amplified DNA was analyzed by gel electrophoresis and was purified using the Amplicon Library Preparation Method (Roche, Branford, CT). Library pools of bar-coded template DNA were prepared, which were used to generate templated beads for NGS. NGS was performed using a Roche 454 instrument (Roche, Branford, CT). The GS Amplicon Variant Analyzer version 2.5 (Roche) was used to generate consensus sequences; quality control procedures were used to determine which sequence reads were included in the analysis. Sequences from samples were aligned with reference sequences using Clustal W. Phylogenetic trees were generated using Neighbor-Joining. Bootstrap values were obtained based on 500 replicate analyses. Additional information about the methods used for this part of the linkage analysis and representative phylogenetic trees are presented in Eshleman et al, 2011, Journal of Infectious Diseases 204:1918-1926. Index-partner pairs were classified as linked if multiple consensus sequences obtained for the index and the partner clustered together on a branch with a high bootstrap value.

Overview of linkage analysis for index-partner pairs in HPTN 052

Seventy-eight partner infections were observed in the HPTN 052 trial (see Figure). *Pol* region sequencing was successful for index and partner samples in 72 cases; linkage status could not be determined for the remaining six cases (two in the early ART arm; four in the delayed ART arm). Forty-two cases were provisionally classified as linked by phylogenetic analysis of *pol* region sequences. The remaining cases were provisionally classified as likely linked (N=3) or unlinked (N=27) by phylogenetic analysis of *pol* region sequences. *Pol* region sequences from all 72 cases were also analyzed using Bayesian methods (see above). In 41 cases, samples were classified as linked by both phylogenetic and Bayesian analysis of *pol* sequences; these cases were classified as linked (no further analysis was performed). Next generation sequencing (NGS, *env* region) was performed for the remaining 31 cases; *env* sequences obtained by NGS were analyzed using

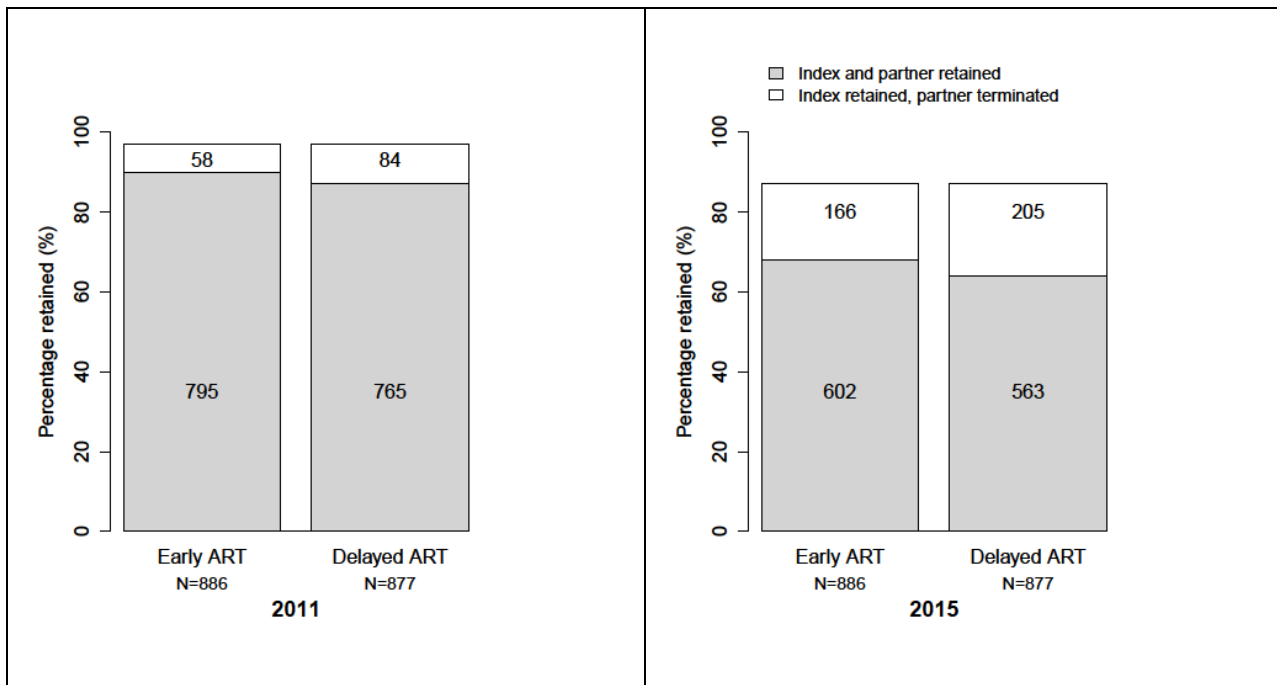
phylogenetic methods (see above). In five cases, partner infections were classified as linked based on NGS. In 20 cases, partner infections were characterized as unlinked based on NGS. NGS failed in six cases (three in the early ART arm; three in the delayed ART arm). All six cases of these cases were clearly unlinked by both phylogenetic and Bayesian analysis of *pol* region sequences; those cases were classified as unlinked based on the *pol* region results. The probability that NGS would have shown linkage in these cases is low. There were 21 cases that were clearly unlinked by *pol* region phylogenetic and Bayesian analysis where NGS results were obtained; only one case was linked by NGS (probability <5%). In that case, the couple was in the delayed ART arm and the index participant had not started ART.

Supplement Figure S1. Linkage analysis of HIV from index-partner pairs.



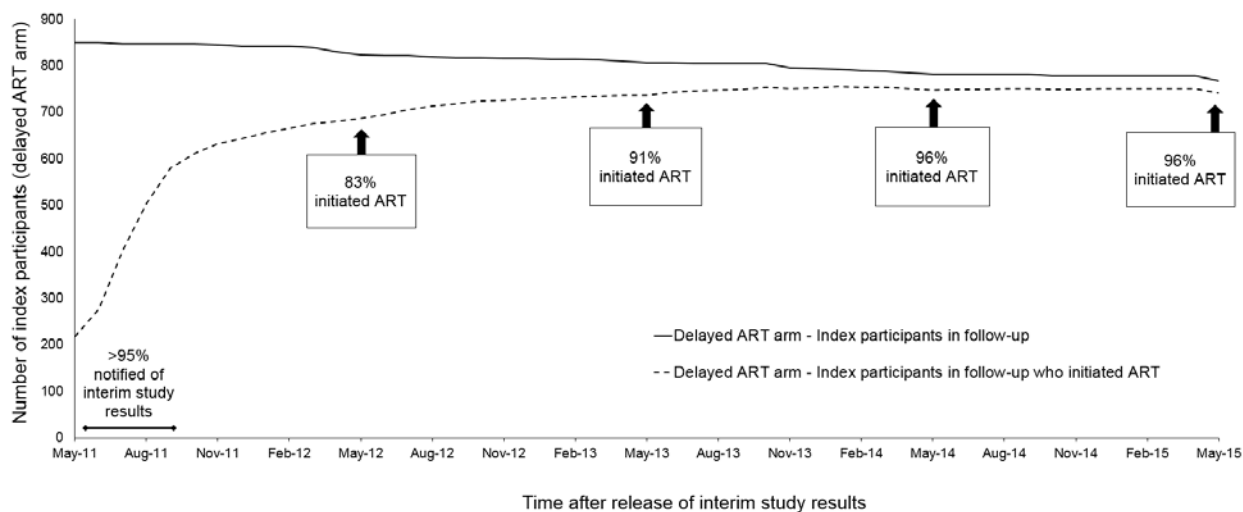
Supplement Figure S2. Retention of study participants.

The graphs show the number and percentage of couples (index and partner participants) retained (shaded) and the number and percentage of index participants retained in cases where the partner was terminated or lost to follow-up (not shaded). Data obtained through May 2011 (prior to release of the interim report) are shown in the left panel; data obtained from May 2011 through May 2015 (after release of the interim study report when all index participants were offered ART) are shown in the right panel.



Supplement Figure S3. Initiation of ART in the delayed ART arm of the study.

The graph shows the number of HIV-infected index participants in the delayed ART arm who were still in follow-up after the release of interim study results (solid line), and the portion of those participants who had initiated ART (dashed line). At the time that the interim results were released (May 2011), 218 (26%) of the index participants in the delayed ART arm had initiated ART, consistent with study guidelines (i.e., due to a fall in CD4 cell count, development of an AIDS-defining illness, or pregnancy). By September 2011, the majority (>95%) of study participants in the delayed ART arm had been notified of the interim study results and offered ART. Most participants chose to start ART as soon as they learned of the interim results, or at their next scheduled study visit; however, others declined for a variety of reasons. Eighty-three percent of index participants in the delayed ART arm who were still in follow-up started ART by May 2012. By the end of the study (May 2015), 96% of index participants who were still in follow-up had started ART.



Supplement Table S1. Cumulative incidence of sexually transmitted infections*.

	Index		Partner	
	Early ART Arm	Delayed ART Arm	Early ART Arm	Delayed ART Arm
Total	886	876	903	889
Person-years	5011.4	5019.9	4324.5	4184.7
HBV ^a	47 (0.94)	47 (0.94)		
Syphilis	78 (1.56)	66 (1.31)	53 (1.23)	50 (1.19)
Gonorrhea	49 (0.98)	53 (1.06)	40 (0.92)	34 (0.81)
C. trachomatis	65 (1.30)	64 (1.27)	49 (1.13)	58 (1.39)
Women	432	441	445	418
Person-years	2435.8	2538.0	2230.7	2115.3
Bact. Vaginosis	190 (7.80)	216 (8.51)	135 (6.05)	121 (5.72)
Trichomonas	74 (3.04)	78 (3.07)	46 (2.06)	41 (1.94)
Yeast	195 (8.01)	204 (8.04)	161 (7.22)	167 (7.89)

* Incidence rates were calculated as number of infections per 100 person-years. Data are shown for men and women combined (Total) and for women only. Person-years of follow-up are shown from the time of study enrollment.

^a Hepatitis B virus (HBV) testing was required only for index participants and was performed only at study enrollment.

Abbreviations: HBV: hepatitis B virus; C. trachomatis: chlamydia trachomatis; Bact: bacterial.

Supplement Table S2. Year-specific incidence rates and rate ratios.

Follow-up period*		Early ART arm			Delayed ART arm			Rate ratio (95% CI) ^a
		Events	PY	Rate (95% CI)	Events	PY	Rate (95% CI)	
		<i>no.</i>		%	<i>no.</i>		%	
All partner infections								
0-1 yr	Before 5/2011	2	834.2	0.2 (0.0-0.9)	19	827.7	2.3 (1.4-3.6)	0.10 (0.01-0.43)
	Total	2	847.6	0.2 (0.0-0.9)	19	839.0	2.3 (1.4-3.5)	0.10 (0.01-0.43)
1-2 yr	Before 5/2011	1	566.5	0.2 (0.0-1.0)	14	563.3	2.5 (1.4-4.2)	0.07 (0.00-0.47)
	Total	2	779.7	0.3 (0.0-0.9)	17	760.1	2.2 (1.3-3.6)	0.11 (0.01-0.48)
2-3 yr	Before 5/2011	1	247.9	0.4 (0.0-2.2)	5	245.3	2.0 (0.7-4.8)	0.20 (0.00-1.77)
	Total	3	721.2	0.4 (0.1-1.2)	8	691.7	1.2 (0.5-2.3)	0.36 (0.06-1.50)
3-4 yr	Before 5/2011	0	60.1	0.0 (0.0-6.1)	4	57.1	7.0 (1.9-17.9)	0.00 (0.00-1.44)
	Total	4	673.6	0.6 (0.2-1.5)	7	638.0	1.1 (0.4-2.3)	0.54 (0.12-2.13)
4-5 yr	Before 5/2011	0	29.8	0.0 (0.0-12.4)	0	24.8	0.0 (0.0-14.8)	-
	Total	4	616.5	0.6 (0.2-1.7)	2	586.7	0.3 (0.0-1.2)	1.90 (0.27- 1.04)
5+ yr	Before 5/2011	0	12.8	0.0 (0.0-28.8)	0	12.8	0.0 (0.0-28.9)	-
	Total	4	686.0	0.6 (0.2-1.5)	6	669.2	0.9 (0.3-2.0)	0.65 (0.13-2.74)
Linked partner infections								
0-1 yr	Before 5/2011	1	834.2	0.1 (0.0-0.7)	17	827.7	2.1 (1.2-3.3)	0.06 (0.00-0.37)
	Total	1	847.6	0.1 (0.0-0.7)	17	839.0	2.0 (1.2-3.2)	0.06 (0.00-0.37)
1-2 yr	Before 5/2011	0	566.5	0.0 (0.0-0.7)	12	563.3	2.1 (1.1-3.7)	0.00 (0.00-0.36)
	Total	0	779.7	0.0 (0.0-0.5)	15	760.1	2.0 (1.1-3.3)	0.00 (0.00-0.27)
2-3 yr	Before 5/2011	0	247.9	0.0 (0.0-1.5)	4	245.3	1.6 (0.4-4.2)	0.00 (0.00-1.50)
	Total	0	721.2	0.0 (0.0-0.5)	6	691.7	0.9 (0.3-1.9)	0.00 (0.00-0.81)
3-4 yr	Before 5/2011	0	60.1	0.0 (0.0-6.1)	3	57.1	5.3 (1.1-15.3)	0.00 (0.00-2.30)
	Total	0	673.6	0.0 (0.0-0.5)	3	638.0	0.5 (0.1-1.4)	0.00 (0.00-2.29)
4-5 yr	Before 5/2011	0	29.8	0.0 (0.0-12.4)	0	24.8	0.0 (0.0-14.8)	-
	Total	1	616.5	0.2 (0.0-0.9)	1	586.7	0.2 (0.0-0.9)	0.96 (0.01-74.7)
5+ yr	Before 5/2011	0	12.8	0.0 (0.0-28.8)	0	12.8	0.0 (0.0-28.9)	-
	Total	1	686.0	0.1 (0.0-0.8)	1	669.2	0.1 (0.0-0.8)	-