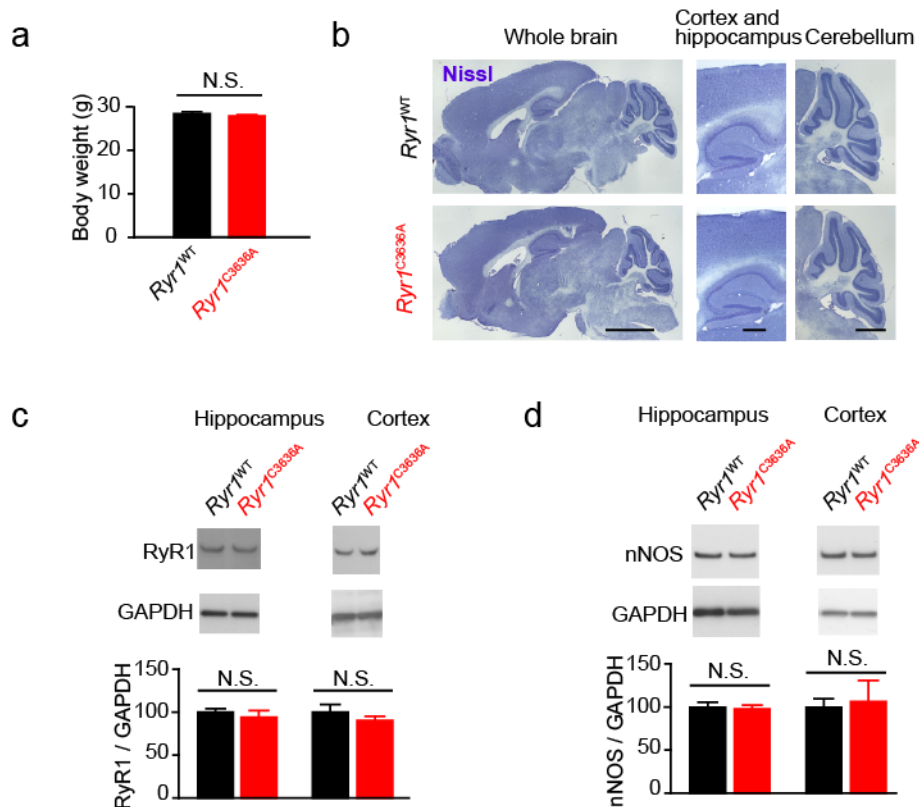


## **SUPPLEMENTARY MATERIALS**

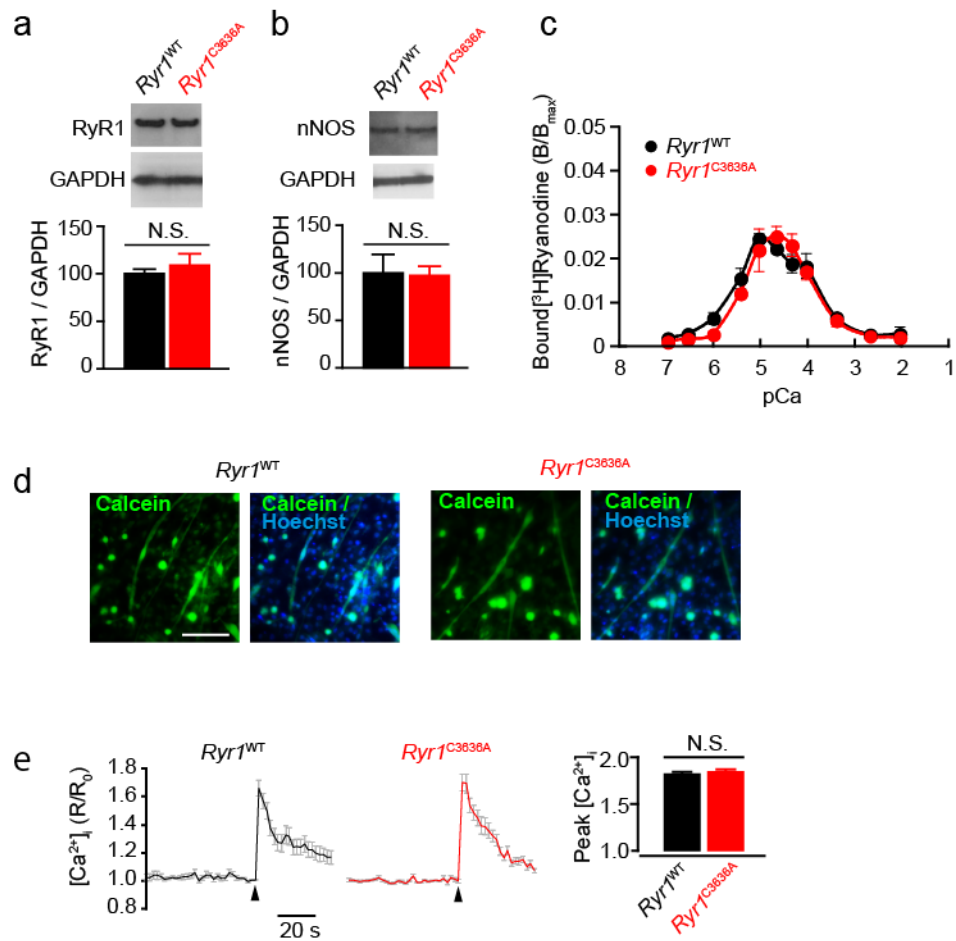
### **Nitric oxide-induced activation of the type 1 ryanodine receptor is critical for epileptic seizure-induced neuronal cell death**

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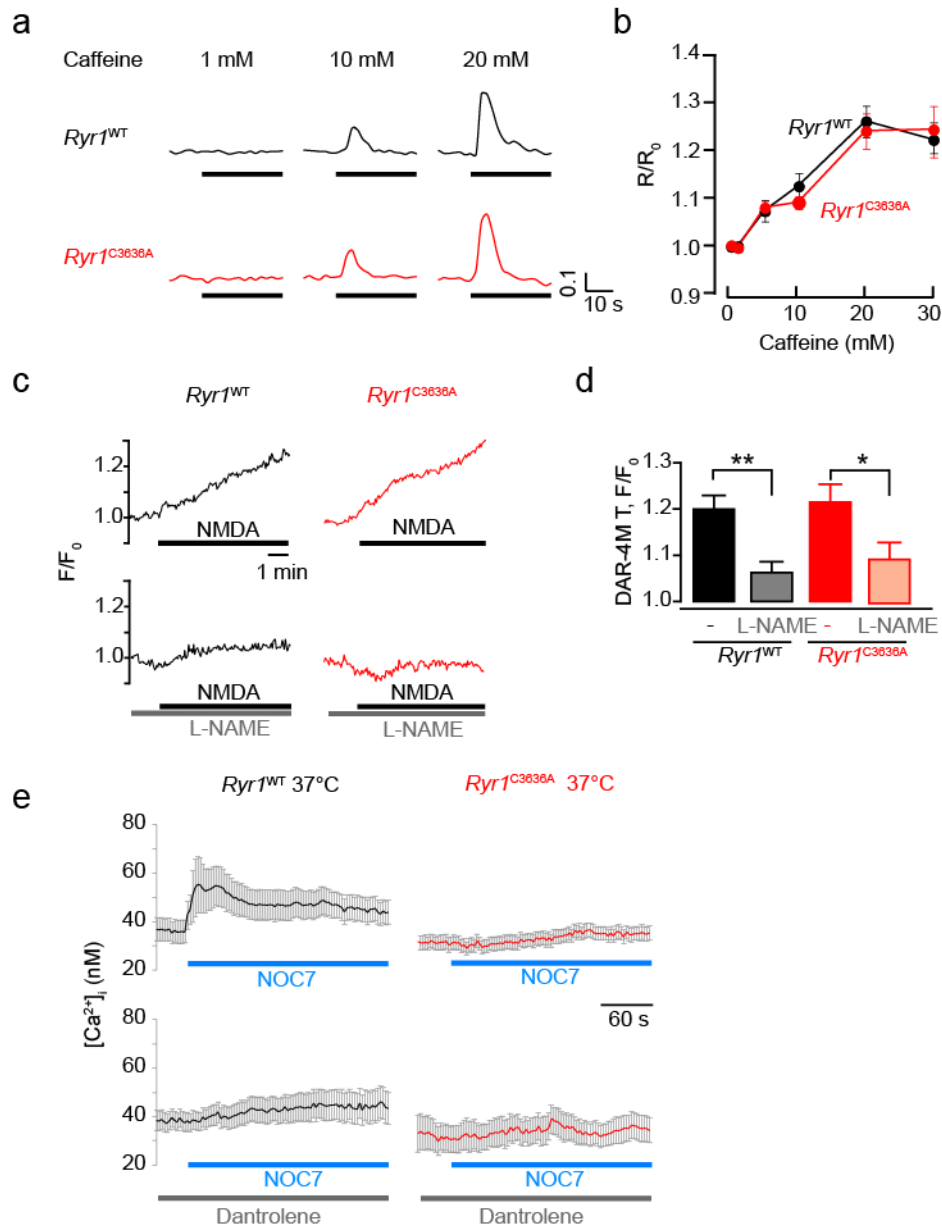
**Figure S1 Characterization of *RyR1*<sup>C3636A</sup> mice and characterization of RyR1<sup>C3636A</sup> channels in brain (Relates to Figure 1)**

**(a).** Body weights of *RyR1*<sup>C3636A</sup> and *RyR1*<sup>WT</sup> littermates at 12 weeks of age.  $n = 7-8$ . Error bars indicate s.e.m. N.S., not significant,  $t$ -test compared with control. **(b)** Representative images of Nissl-stained sagittal sections from brains of *RyR1*<sup>WT</sup> littermates and *RyR1*<sup>C3636A</sup> mice at 12 weeks of age. Scale bar: 5 mm (whole brain), 500  $\mu$ m (cortex and hippocampus), 1 mm (cerebellum). **(c)** Immunoblotting of cytosol and microsome-enriched fractions from hippocampus (left) and cerebral cortex (right) using antibodies against RyR1 and GAPDH (loading control). Samples were collected from *RyR1*<sup>C3636A</sup> and *RyR1*<sup>WT</sup> littermates at 12 weeks of age;  $n = 3-4$ . Error bars indicate s.e.m. N.S., not significant,  $t$ -test compared with control. **(d)** Expression levels were analyzed by immunoblotting using antibodies to nNOS and GAPDH;  $n = 3-4$ . Error bars indicate s.e.m. N.S., not significant,  $t$ -test compared with control.



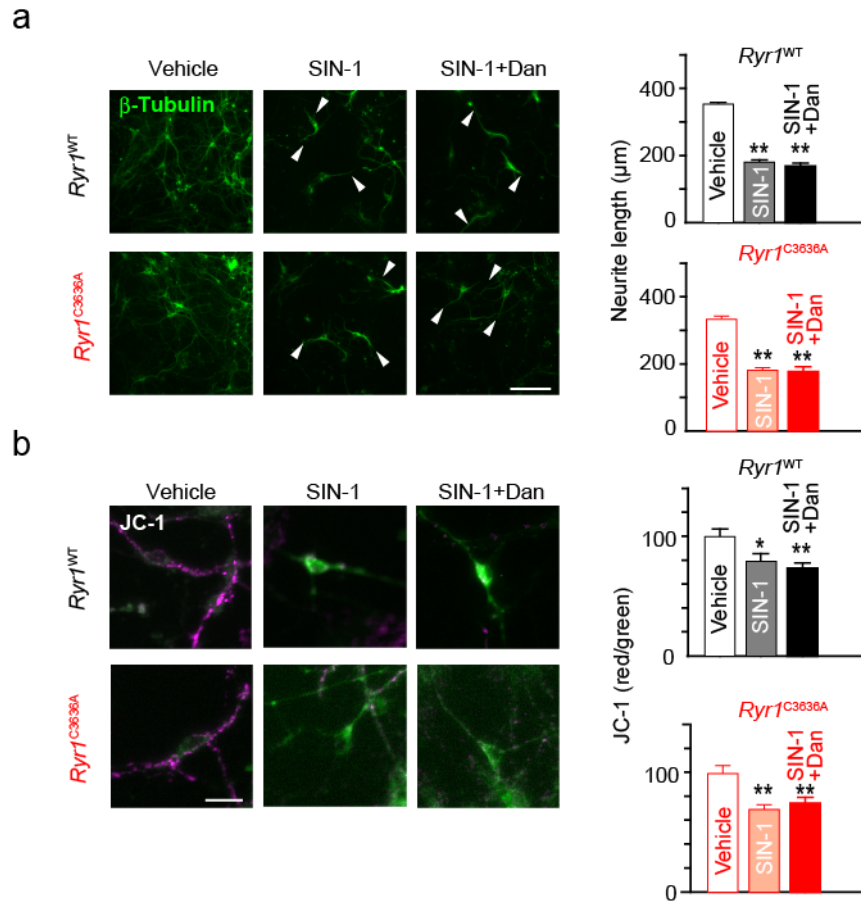
**Figure S2 Characterization of RyR1<sup>C3636A</sup> channels in skeletal muscle (Relates to Figure 1)**

**(a, b)** Immunoblotting of cytosol and microsomal-enriched fractions from skeletal muscle, using antibodies against RyR1 (a) or nNOS (b) and GAPDH (loading control). Samples were collected from *Ryr1*<sup>C3636A</sup> and *Ryr1*<sup>WT</sup> littermates at 12 weeks of age;  $n = 3-4$ . Error bars indicate s.e.m. N.S., not significant,  $t$ -test compared with control. **(c)** Ca<sup>2+</sup>-dependent [<sup>3</sup>H]ryanodine binding to *Ryr1*<sup>WT</sup> (black) and *Ryr1*<sup>C3636A</sup> (red) sarcoplasmic reticulum vesicles. **(d)** Fluorescence images of calcein-AM (green) and Hoechst 33342 (blue) stained primary cultured myocytes. Scale bar: 100  $\mu$ m. **(e)** Intracellular Ca<sup>2+</sup> response to field stimulation in *Ryr1*<sup>C3636A</sup> and *Ryr1*<sup>WT</sup> myocytes measured at 2 frames s<sup>-1</sup>.  $R$  ( $F_{340}/F_{380}$  of Fura-2) was normalized by the initial  $R$  ( $R_0$ ). Peak amplitudes were compared between *Ryr1*<sup>WT</sup> (36 cells) and *Ryr1*<sup>C3636A</sup> (42 cells) myocytes. Error bars indicate s.e.m. N.S., not significant,  $t$ -test compared with control.



**Figure S3 Characterization of RyR1<sup>C3636A</sup> channels in primary cultured neurons (Relates to Figure 1)**

(a) Caffeine-induced Ca<sup>2+</sup> responses (expressed as  $R/R_0$ ) in *Ryr1<sup>C3636A</sup>* and *Ryr1<sup>WT</sup>* neurons. (b) Peak amplitudes of [Ca<sup>2+</sup>]<sub>i</sub> at varied caffeine concentrations in *Ryr1<sup>WT</sup>* and *Ryr1<sup>C3636A</sup>* neurons;  $n = 13\text{--}35$  neurons. (c) NO production measured with DAR-4M with or without a NOS inhibitor L-NAME. Fluorescence intensity of DAR-4M was normalized by the initial value. (d) Magnitude of NO production at 6 min after administration of NMDA in *Ryr1<sup>WT</sup>* ( $n = 15\text{--}19$  cells) and *Ryr1<sup>C3636A</sup>* ( $n = 18\text{--}19$  cells). Error bars indicate s.e.m.  $t$ -test. \*  $p < 0.05$ , \*\*  $p < 0.01$ . (e) NICR was blocked by dantrolene. NOC7 (500  $\mu\text{M}$ )-induced intracellular Ca<sup>2+</sup> increase at 37°C with or without dantrolene (10  $\mu\text{M}$ ) in cultured cerebral neurons of *Ryr1<sup>WT</sup>* or *Ryr1<sup>C3636A</sup>* mice;  $n = 17\text{--}30$  neurons, error bars indicates s.e.m.



**Figure S4** Peroxynitrite donor SIN-1 induces neuronal cell death in both *Ryr1*<sup>WT</sup> and *Ryr1*<sup>C3636A</sup> neurons to the same extent (Relates to Figure 3)

**(a)** Effect of the application of peroxynitrite donor SIN-1 (3-morpholino-sydnonimine; 100 μM) on neuron morphology (stained with anti-β-III tubulin antibody). Arrowheads indicate short and curly neurites. Graphs show the length of the longest neurite of each neuron;  $n = 95-125$ ; error bars indicate s.e.m. Data were analyzed for significance using ANOVA followed by a Tukey-Kramer *post-hoc* test. \*\*  $p < 0.01$ . Dan, dantrolene. **(b)** Cell viability examined by JC-1 assay.  $n = 65-81$ . Error bars indicate s.e.m. Data were analyzed for significance using ANOVA followed by a Tukey-Kramer *post-hoc* test. \*\*  $p < 0.01$ , \*  $p < 0.05$ .