

Table S4. Bacterial strains, plasmids, and primers used in this study.

Bacterial strain or plasmid	Description	Reference or source
<u><i>P. aeruginosa</i> strains</u>		
PAO1	Wild type strain	(1)
PAO1 $\Delta lasR$	PAO1 containing an unmarked, in-frame <i>lasR</i> deletion	(2)
PAO1 $\Delta lasR \Delta rhIR \Delta qscR$	<i>lasR</i> , <i>rhIR</i> , <i>qscR</i> triple mutant in PAO1	(3)
<u>Other strains</u>		
<i>E. coli</i> DH5 α	F ⁻ $\Phi 80 lacZ \Delta M15 \Delta(lacZYA-argF)$ U169 <i>recA1 endA1 hsdR17</i> (r _k ⁻ , m _k ⁺) <i>phoA supE44 thi-1 gyrA96 relA1</i> λ^-	Invitrogen
<u>Plasmids</u>		
pSC11	Broad-host-range <i>lasI-lacZ</i> reporter, Ap ^r	(4)
pJN105L	Arabinose inducible <i>lasR</i> in pJN105, Gm ^r	(5)
pECP61.5	Contains an <i>rhlA-lacZ</i> translation fusion and IPTG-inducible <i>rhIR</i> , Ap ^r	(6)
pPROBE-GT	Broad-host-range pVS1/p15a GFP reporter, Gm ^r	(7)
pJF01	pPROBE-GT with -1 through-501 5' region of <i>rhlA</i> inserted with HindIII and BamHI, Gm ^r	This work
pBS351	pPROBE-GT with -1 through-501 5' region of <i>lasI</i> inserted with HindIII and BamHI, Gm ^r	This work
<u>Primers</u>		
<i>lasR</i> .cloning.EcoR1 F	5' – TTTTTGAATTCTAGCGCTATGGCCTTGGTT – 3'	
<i>lasR</i> .cloning.Xba1 R	5' – AAAAATCTAGAGCAAGATCAGAGAGTAATAAGACCC – 3'	
<i>rhlA</i> .GFPcloning.HindIII F	5' – TTTTAAAGCTTGCATGCGAGGCCTGCGAA – 3'	
<i>rhlA</i> .GFPcloning.BamHI R	5' – AAAAAAGGATCCTCTAGAAACCGATACCAACAGACTTTTCGC – 3'	
GFP.QC R	5' – TTCTTTCCTGTACATAACCTTCGGGCA – 3'	

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