

Supporting information

Activation mechanism of the *Bacteroides fragilis* cysteine peptidase, Fragipain

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Table S1: primers

Primer	Sequence	Comment
R147A-UP	GCAAAGACAG GCT TGGTTTGGT	Primers used to mutate residue R147 present in the cleavage site to alanine.
R147A-LO	ACCAAACCA AGCT GTCTTTGC	
R147K-UP	GCAAAGACAA AAG TGGTTTGGT	Primers used to mutate residue R147 present in the cleavage site to lysine.
R147K-LO	ACCAAACCA CTT GTCTTTGC	
L Δ 5-UP	GAGGGATGGATACCTCGTTGGTTTGGTCAG	Primers used to delete 5 residues (Δ S142-T146) in the cleavable loop.
L Δ 5-LO	CTGACCAAACCAACGAGGTATCCATCCCTC	
L Δ 7-UP	CACGGTGAGGGATGGCGTTGGTTTGGTCAG	Primers used to delete 7 residues (Δ I140-T146) in the cleavable loop.
L Δ 7-LO	CTGACCAAACCAACGCCATCCCTCACCGTG	
L Δ 7+R147A-UP	CACGGTGAGGGATGG GCA TGGTTTGGTCAG	Primers used to delete 7 residues (Δ I140-T146) in the cleavable loop and to mutate residue R147 to alanine.
L Δ 7+R147A-LO	CTGACCAAACCA TGCC CATCCCTCACCGTG	

Table S2: strains

Strain #	Organism	Genetic background	Plasmid	Comment	Reference
FC2470	<i>E. coli</i> Rosetta (DE3) PlysS	-	pET28- <i>fpn</i>	Strains used to express wild-type Fpn.	Choi V. <i>et al.</i> (2016). Nature Medicine.
FC2471	<i>E. coli</i> Rosetta (DE3) PlysS	-	pET28- <i>fpn</i> (H135A+C180A)	Strains used to express Fpn (H135A+C180A) active site mutant.	
FC2472	<i>E. coli</i> Rosetta (DE3) PlysS	-	pET28- <i>fpn</i> (R147A)	Strains used to express Fpn (R147A) cleavage site mutant.	This work.
FC2473	<i>E. coli</i> Rosetta (DE3) PlysS	-	pET28- <i>fpn</i> (R147K)	Strains used to express Fpn (R147K) cleavage site mutant.	
FC2474	<i>E. coli</i> Rosetta (DE3) PlysS	-	pET28- <i>fpn</i> (LΔ5)	Strains used to express Fpn (LΔ5) cleavable loop mutant.	
FC2475	<i>E. coli</i> Rosetta (DE3) PlysS	-	pET28- <i>fpn</i> (LΔ7)	Strains used to express Fpn (LΔ7) cleavable loop mutant.	
FC2476	<i>E. coli</i> Rosetta (DE3) PlysS	-	pET28- <i>fpn</i> (LΔ7+R147A)	Strains used to express Fpn (LΔ7+R147A) cleavable loop mutant.	
FC2477	<i>E. coli</i> Rosetta (DE3) PlysS	-	pET28- <i>bft</i>	Strains used to express BFT.	Choi V. <i>et al.</i> (2016). Nature Medicine.
-	<i>B. fragilis</i>	Δ <i>fpn</i>	-	Strain used for western-blot.	
-	<i>B. fragilis</i>	Δ <i>fpn</i>	pAH2- <i>fpn</i>	Strain used for western-blot.	This work. See [Choi V. <i>et al.</i> (2016). Nature Medicine and Hecht A <i>et al.</i> (2016; in press). EMBO reports] for primer and method information.
-	<i>B. fragilis</i>	Δ <i>fpn</i>	pAH2- <i>fpn</i> (H135A+C180A)	Strain used for western-blot.	
-	<i>B. fragilis</i>	Δ <i>fpn</i>	pAH2- <i>fpn</i> (LΔ7)	Strain used for western-blot.	

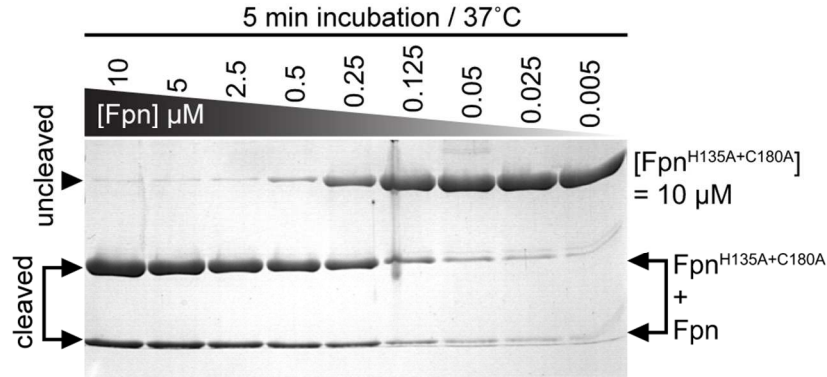


Figure S1: Concentration dependence of Fpn cleavage in *trans*. 10- μ l samples containing 10 μ M of inactive Fpn^{H135A+C180A} and decreasing concentrations of wild-type Fpn (final concentrations tested: 10, 5, 2.5, 0.5, 0.25, 0.125, 0.05, 0.025 and 0.005 μ M) were incubated at 37°C for 5 minutes and resolved by SDS-PAGE.

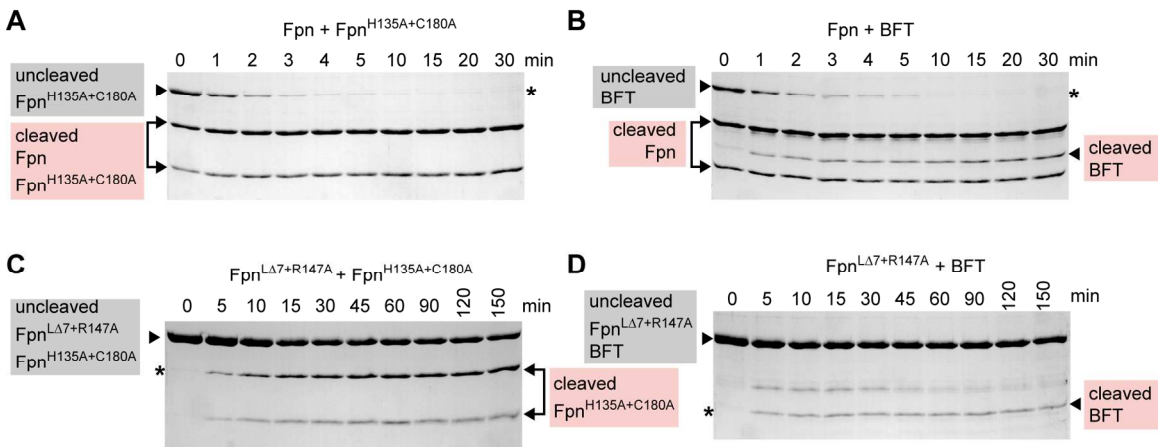


Figure S2: Time course of BFT or Fpn^{H135A+H180} cleavage by wild-type Fpn or the loopless pro form Fpn mutant (Fpn^{Δ7+R147A}). A) Digestion of Fpn active site mutant (Fpn^{H135A+H180}) (5 μ M) by wild-type Fpn (5 μ M). B) Digestion of BFT (5 μ M) by wild-type Fpn (5 μ M). C) Digestion of Fpn^{H135A+H180} (5 μ M) by loopless Fpn^{Δ7+R147A} (5 μ M). D) Digestion of BFT (5 μ M) by Fpn^{Δ7+R147A} (5 μ M). Bands quantified with imageJ are marked with a star.

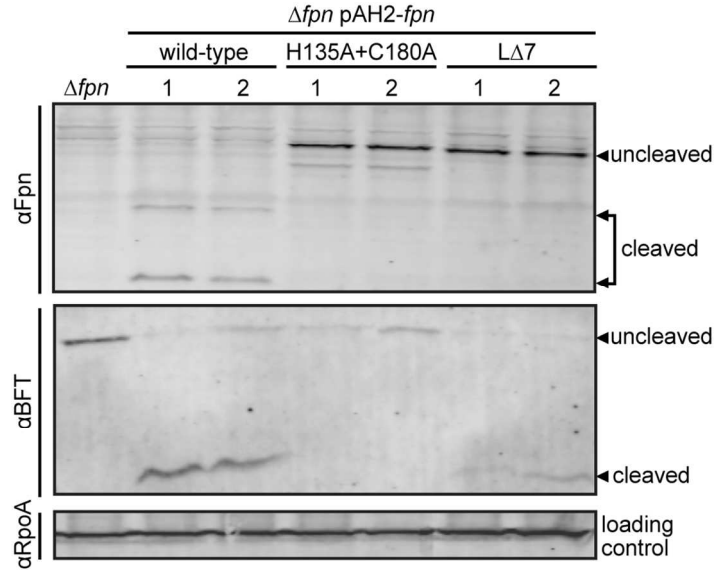


Figure S3: Western blot of *B. fragilis* Δfpn lysate ectopically expressing wild-type Fpn, Fpn^{H135A+C180A} and Fpn^{L Δ 7} proteins. Cell lysate supernatants were used to evaluate the ability of these proteins to auto-cleave (upper blot) and to process BFT (middle blot). RpoA protein (lower blot) was used as a loading control.