

Supplemental Fig. 1. Agreement between the experimental pI values and the reference pH gradient.

Five hRSV-infected lysates (hRSV) and five mock-infected lysates (Mock) were individually separated into 24 fractions using protein OGE. For each OGE separation, the calculated pI values for selected proteoform sequences observed to focus to each OGE fraction were used to represent the experimental pI values.

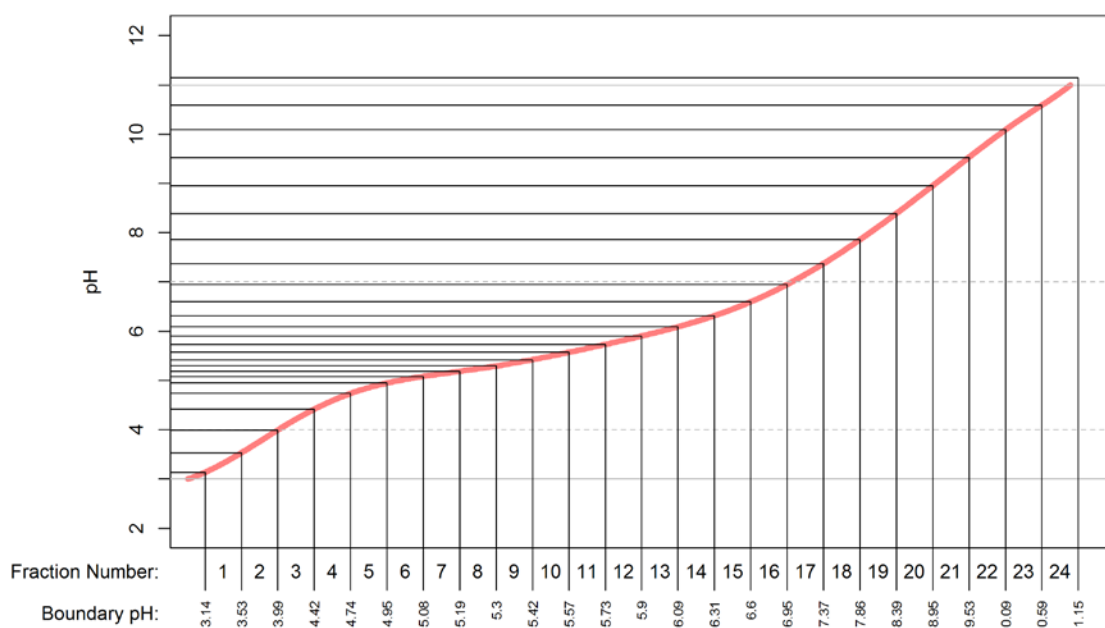
In the following gradient plots, the red line represents the 3-11 nonlinear reference pH gradient (Immobiline DryStrip pH 3-11 non-linear gradient, GE Healthcare). To represent the experimental fractionation patterns, the distribution of pI values for each OGE fraction is superimposed. A boxplot is used to represent the distribution of pI values for each fraction, with the exception of fractions that had less than ten pI values, in which case all pI values were plotted. The number of pI values representing each OGE fraction is presented in Supplemental Table 1.

The boxplots were generated using the “boxplot” function from the graphics package in R (version 2.15.1 (1)). Accordingly, the median pI is represented as a thick black line that is surrounded by a box that represents the interquartile range (i.e., the median \pm 25% of the data). The whiskers represent the range of pI values observed in the fraction and open circles represent outliers. Here an outlier is defined as a pI value that is greater than 1.5 times the interquartile range above the upper quartile or below the lower quartile.

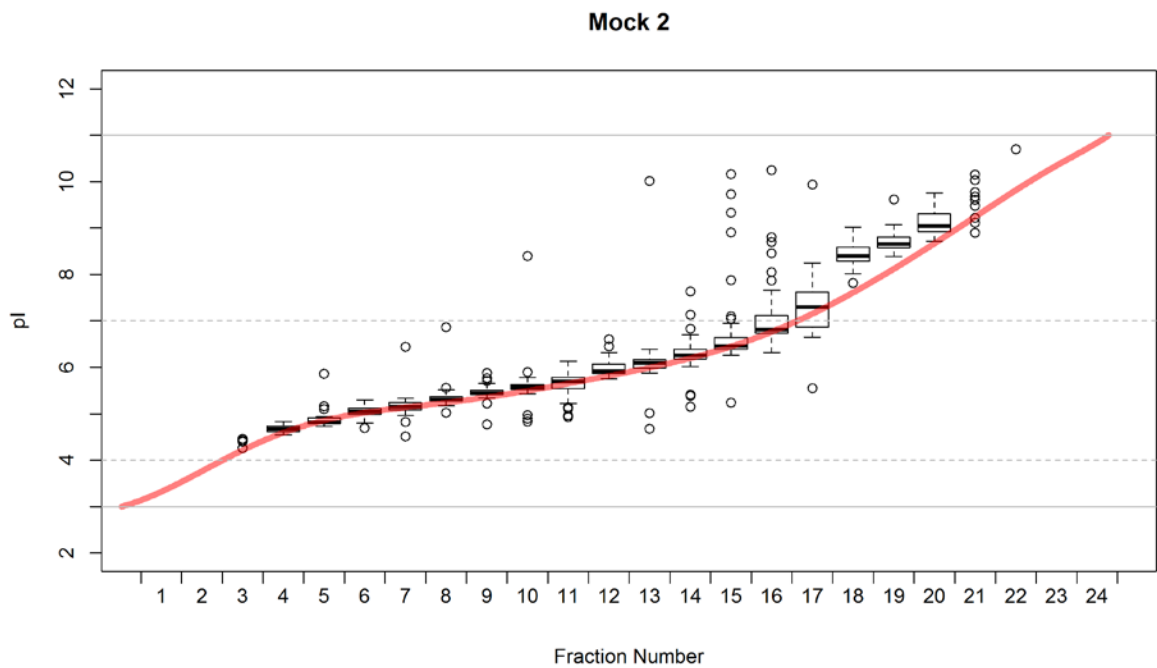
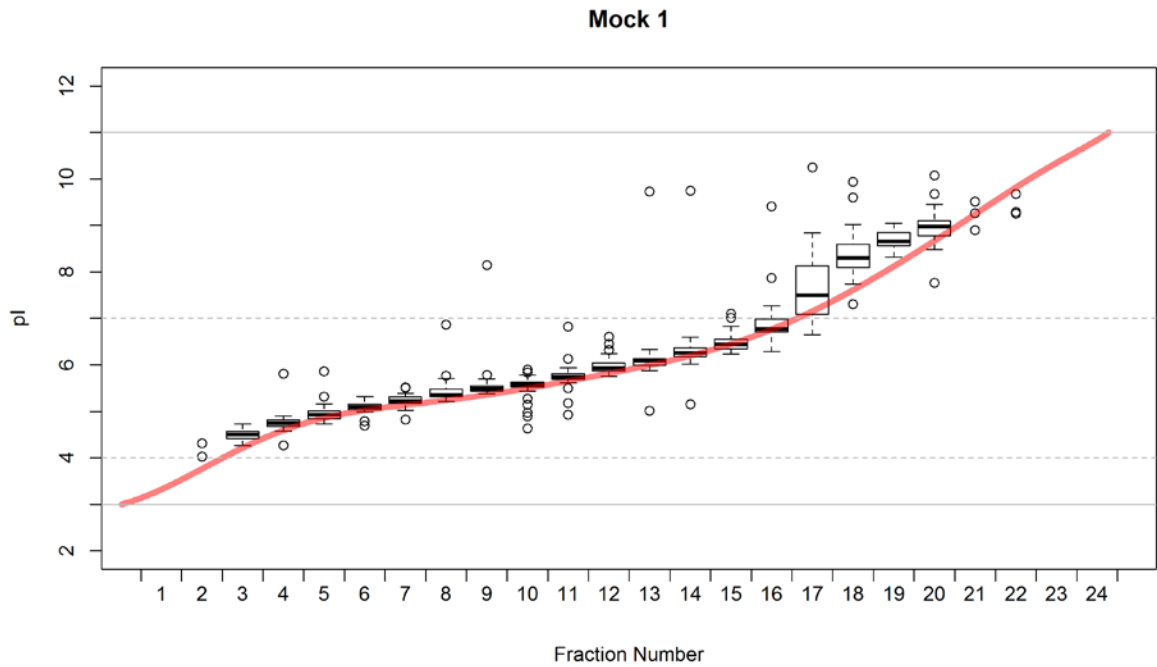
Reference:

1. R Core Team (2016). R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria.

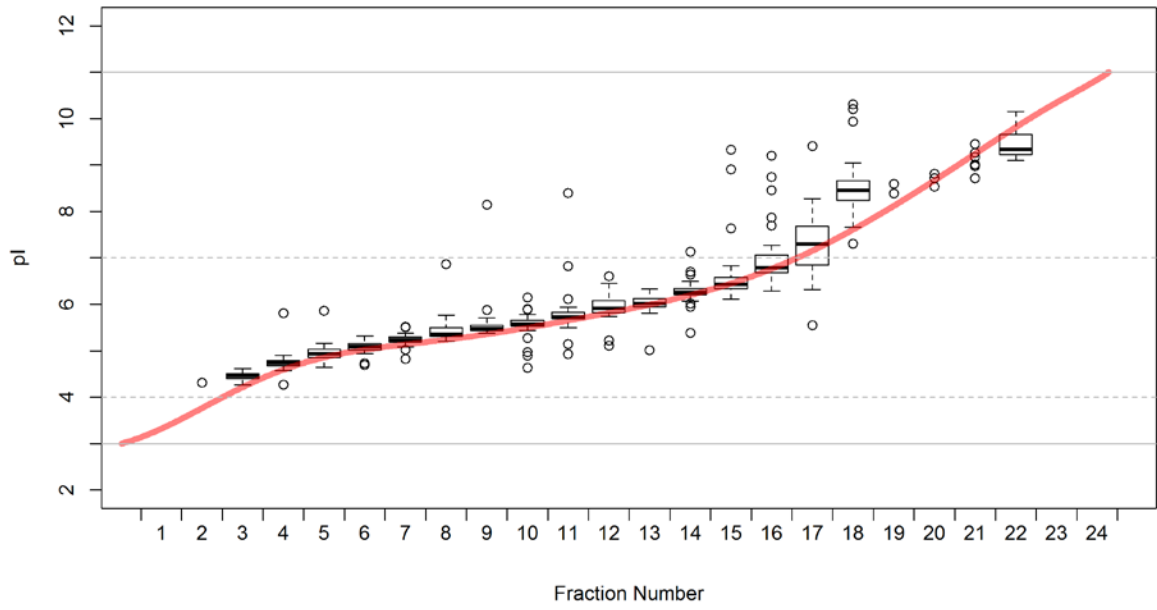
A) Reference pH gradient



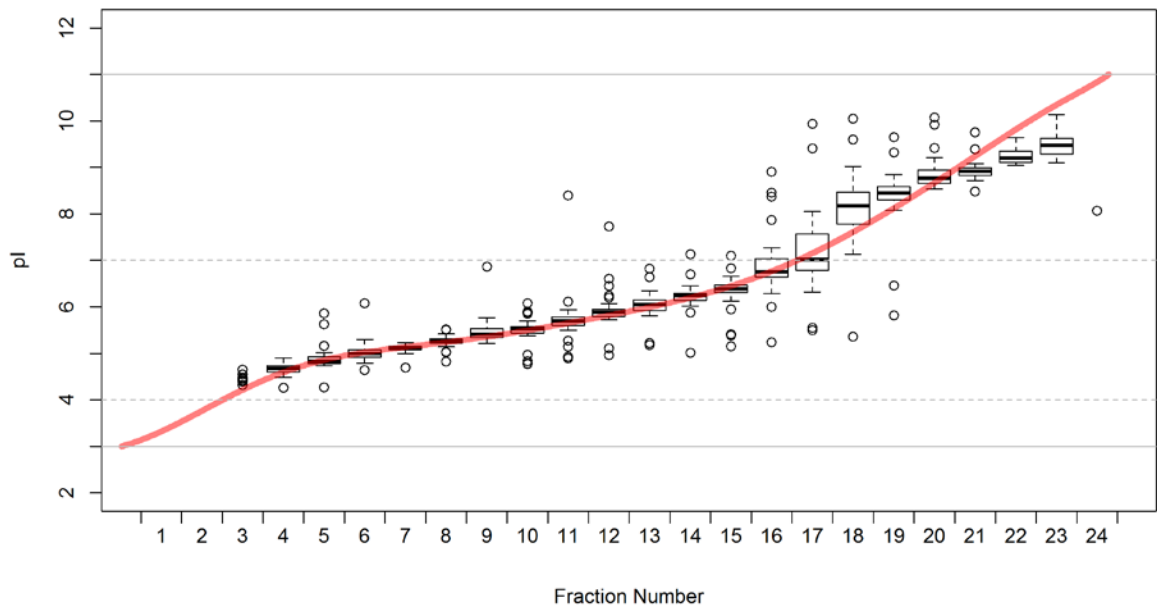
B) Agreement between the distribution of experimental pI values and the reference pH gradient for the ten OGE separations. Five replicate sets of uninfected (Mock) and hRSV-infected A549 cell lysates were fractionated.



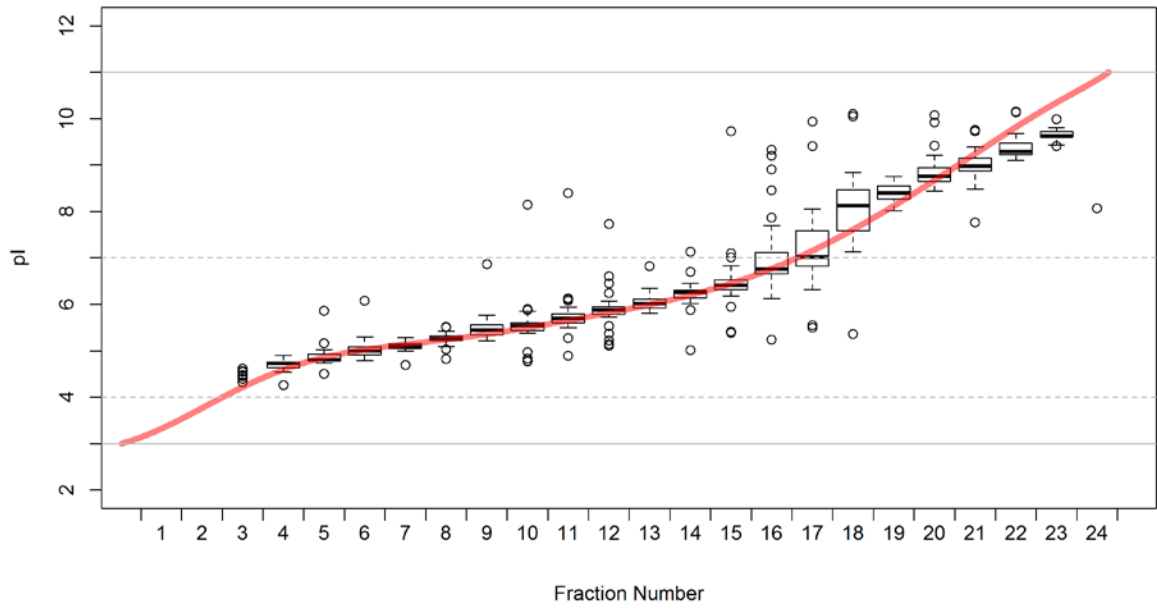
Mock 3



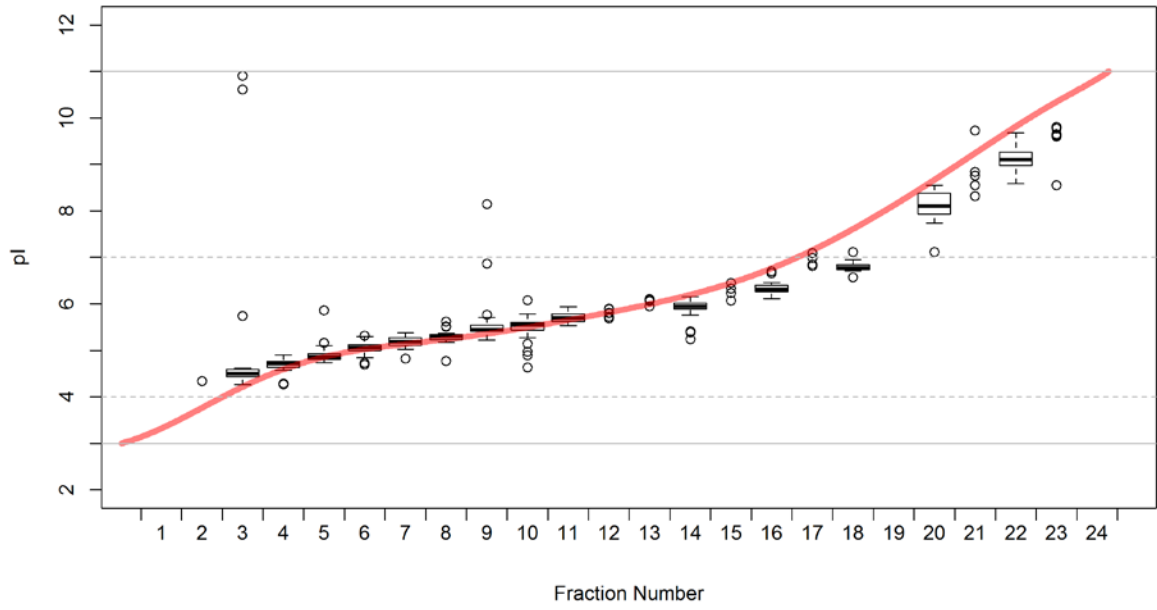
Mock 4



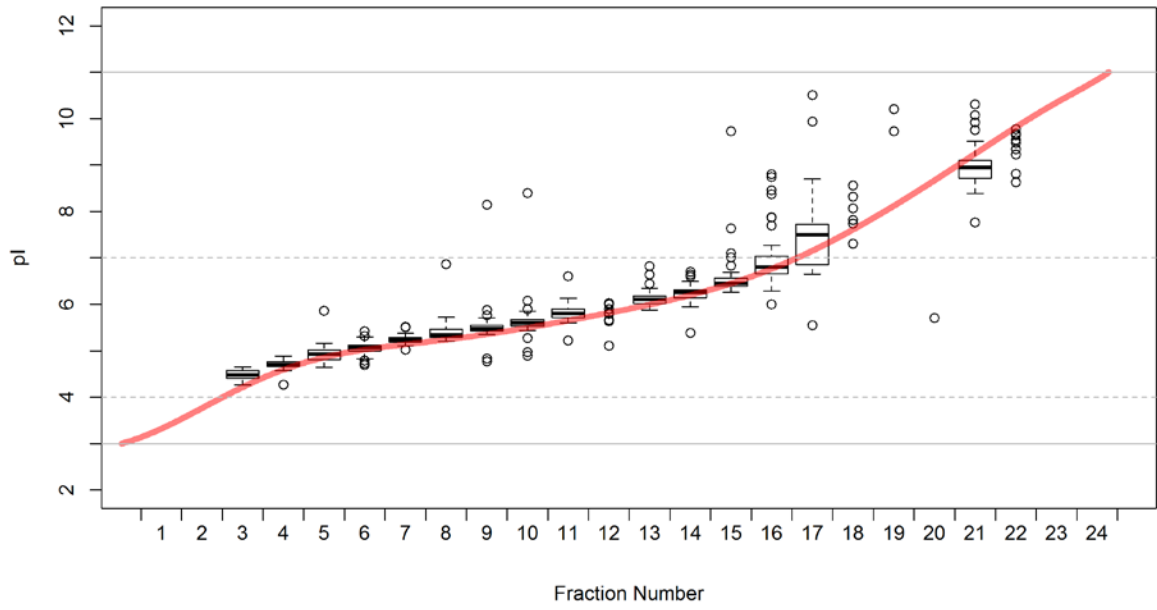
Mock 5



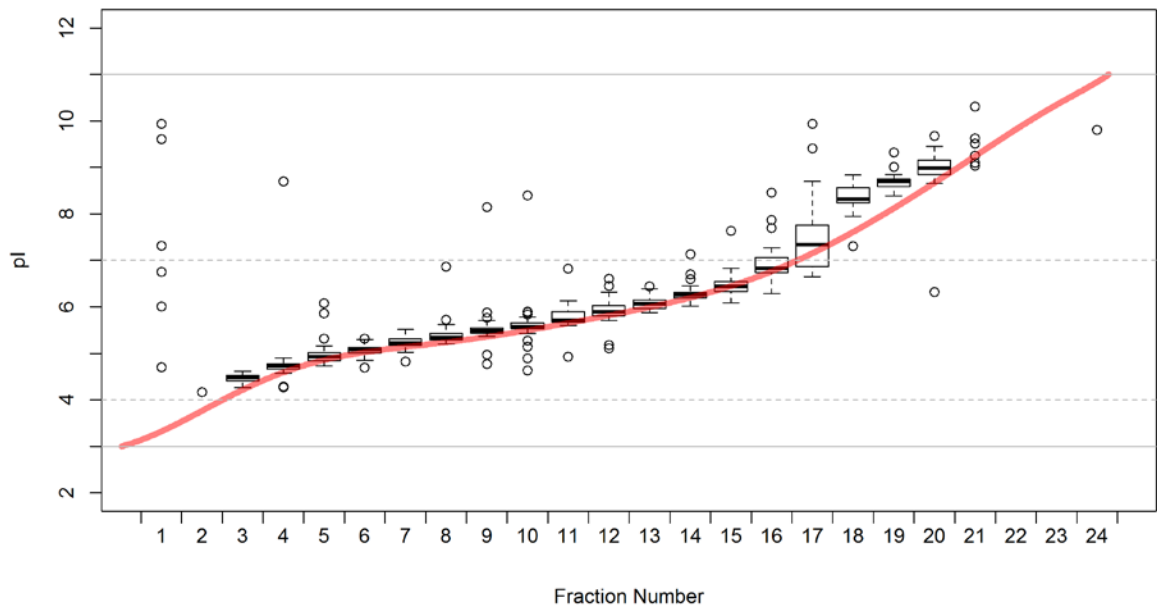
hRSV 1



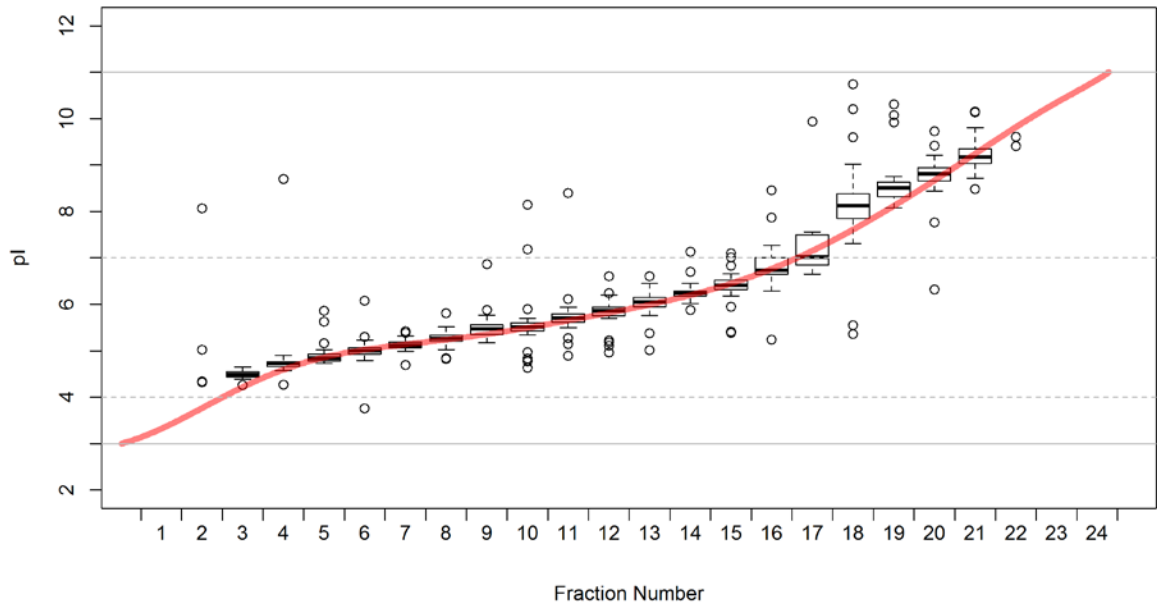
hRSV 2



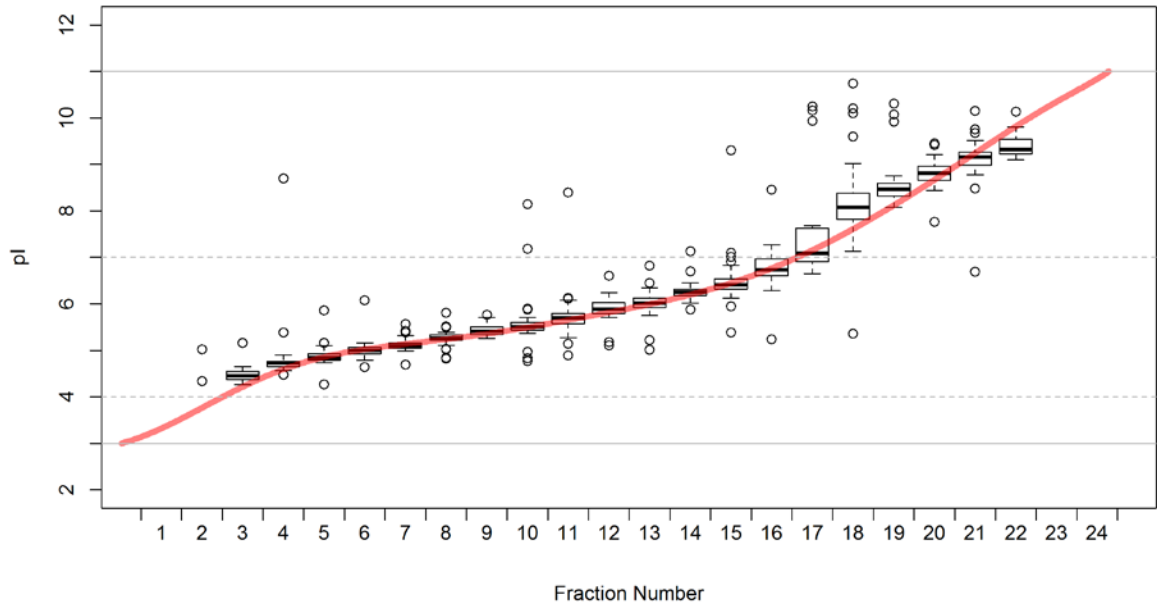
hRSV 3



hRSV 4

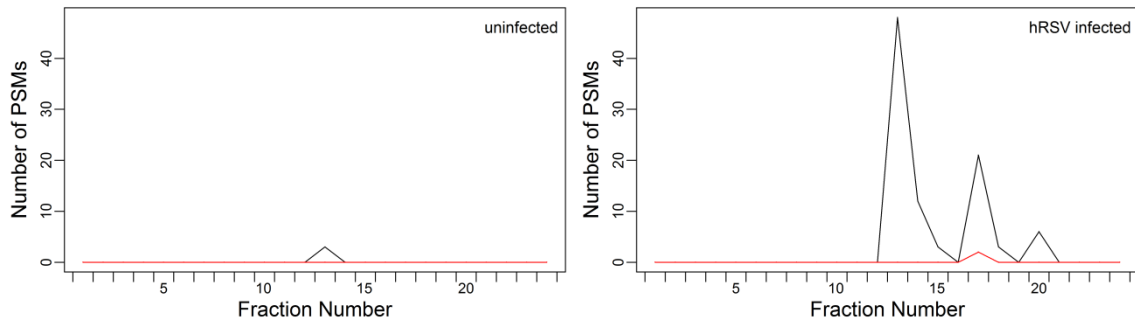


hRSV 5



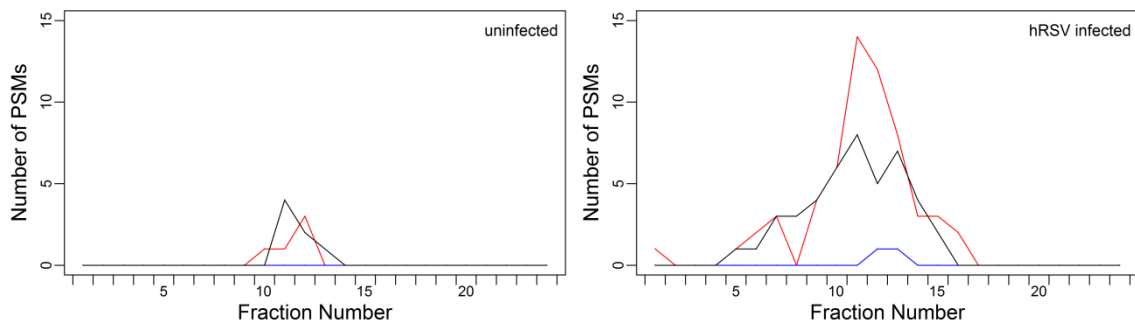
Supplemental Fig. 2. **Traces for proteoform-specific peptides for the A) PSMB10, B) STAT1 and C) WARS protein groups.**

A) PSMB10



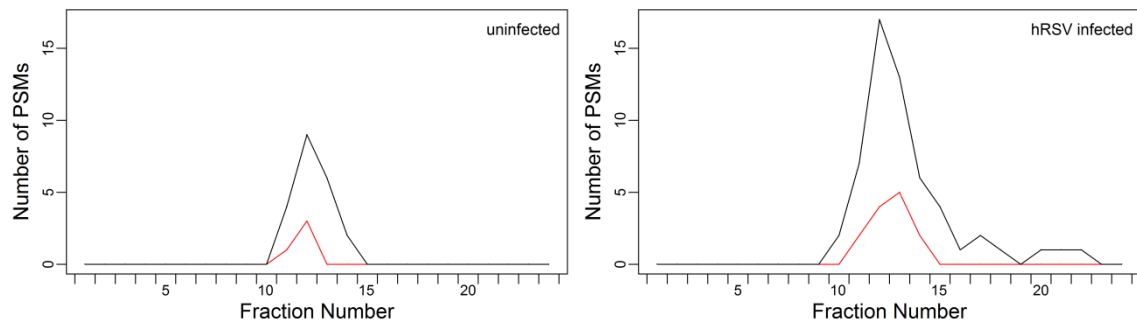
Traces for peptides assigned to the PSMB10 protein group in uninfected and hRSV-infected lysates (left and right, respectively). The **RED** traces represent PSMs for the N-terminal peptide that exclusively matched the precursor proteoform (peptide sequence MLKPALEPR exclusively matching P40306). The **BLACK** traces represent PSMs that matched both the precursor and processed PSMB10 proteoforms (P40306 and P40306 40-273 respectively).

B) STAT1

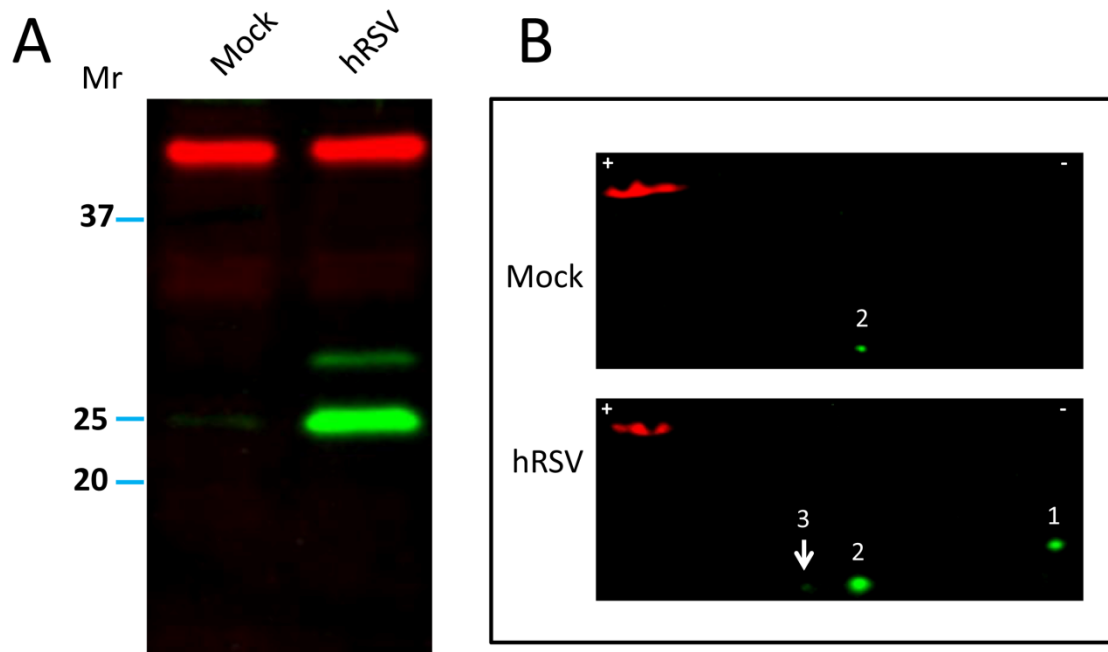


Traces for peptides assigned to the STAT1 protein group in uninfected and hRSV-infected lysates (left and right, respectively). The **RED** traces represent PSMs exclusively matching the Stat1- α proteoform (peptide sequences IVGSVEFDSMMNTV, LQTTDNLPLMSPEEFDEVSR and TELISVSEVHPSR exclusively matching proteoform sequence P42224) and the **BLUE** traces represent PSMs for the C-terminal peptide of Stat1- β (peptide sequence TELISVSEV matching Stat1- β proteoform sequences P42224-2 and J3KPM9). The **BLACK** traces represent an abundant peptide that matches both Stat1- α and Stat1- β proteoforms (peptide sequence FLEQVHQLYDDSPMEIR matching proteoform sequences P42224, D2KFR9, P42224-2 and J3KPM9).

C) WARS



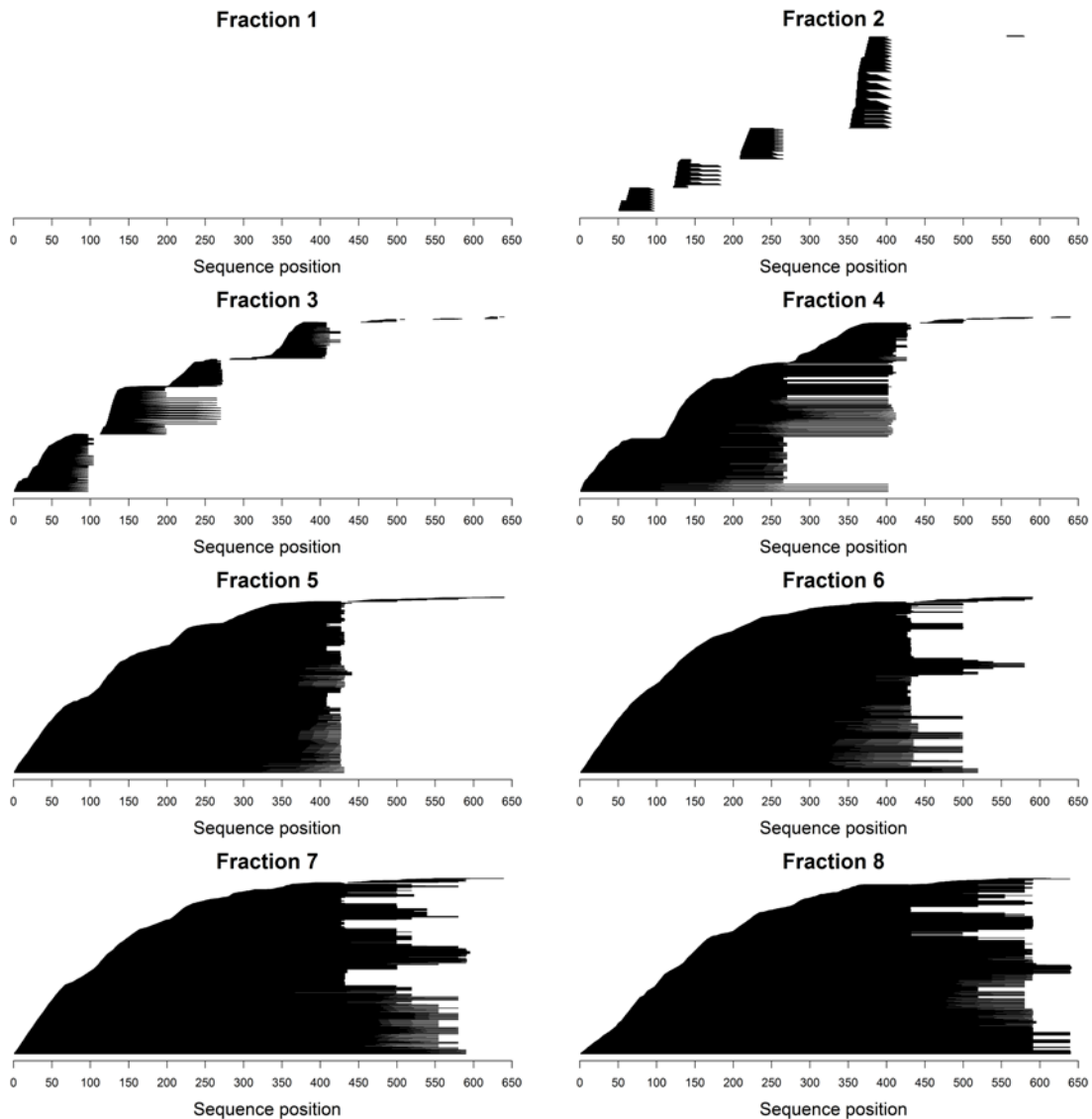
Traces for peptides assigned to the WARS protein group in uninfected and hRSV-infected lysates (left and right, respectively). The **RED** traces represent PSMs that exclusively matched the full length TrpRS sequence (peptide sequence AGNASKDEIDSAVK exclusively matching proteoform sequence P23381). The **BLACK** traces represent PSMs for an abundant shared peptide (ALIEVLQPLIAEHQAR) that matched the full length TrpRS proteoform (P23381), processed proteoforms T1-TrpRS and T2-TrpRS (P23381 71-471 and P23381 94-471, respectively) and the alternative splice form mini-TrpRS (P23381-2).



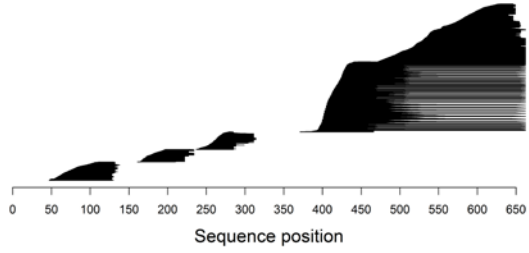
Supplemental Fig. 3. **Western blot characterization of PSMB10 proteoforms induced by hRSV infection of A549 cells.**

(A) One-dimensional-Western blot uninfected (Mock) and hRSV-infected (hRSV) A549 whole cell lysates (10 μ g). Green bands represent PSMB10 specific reactivity and the red band is β -actin specific reactivity. (B) Two-dimensional-Western blots of equal quantities (20 μ g) of protein from lysates of Mock (top panel) and hRSV (bottom panel) infected A549 cells. Green spots correspond to the precursor (spot 1), activated (spot 2) and activated with an putative acidic PTM (arrowed spot 3) proteoforms of PSMB10. The red area of reactivity, corresponding to approximately 42 kDa, represents β -actin used as a loading control.

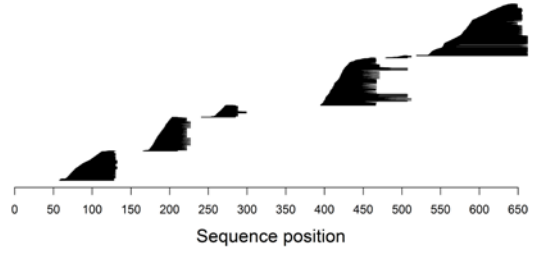
Supplemental Fig. 4. **Predicted focusing of MxA fragments.** Focusing is predicted for all MxA fragments generated by unspecific cleavage of the full length proteoform sequence P20591. Short fragments that do not contain at least one tryptic peptide were excluded from the analysis. Each black line represents an MxA fragment predicted to focus to the respective OGE fraction. For each OGE fraction, fragments are aligned to the full length MxA proteoform sequence (P20591) and are stacked according to the index of the N-terminal residue in the full length sequence, then according to the fragment length. MxA fragments containing the N-terminal region of MxA are predicted to focus to approximately fraction 4 to 9 whereas fragments containing the C-terminal region of MxA are predicted to focus to approximately fraction 14 to 21. The full length MxA proteoform sequence is predicted to focus to fraction 11.



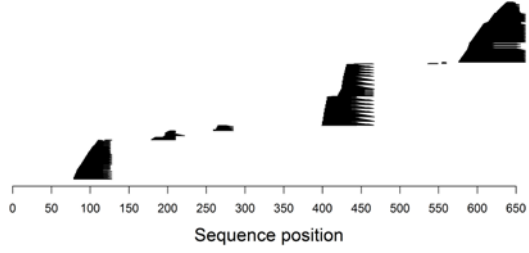
Fraction 21



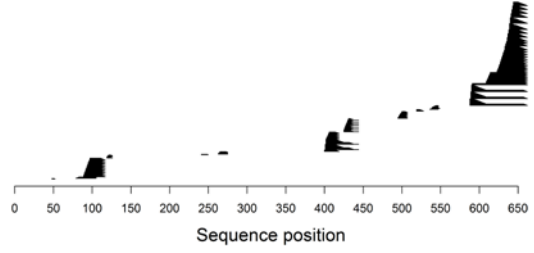
Fraction 22

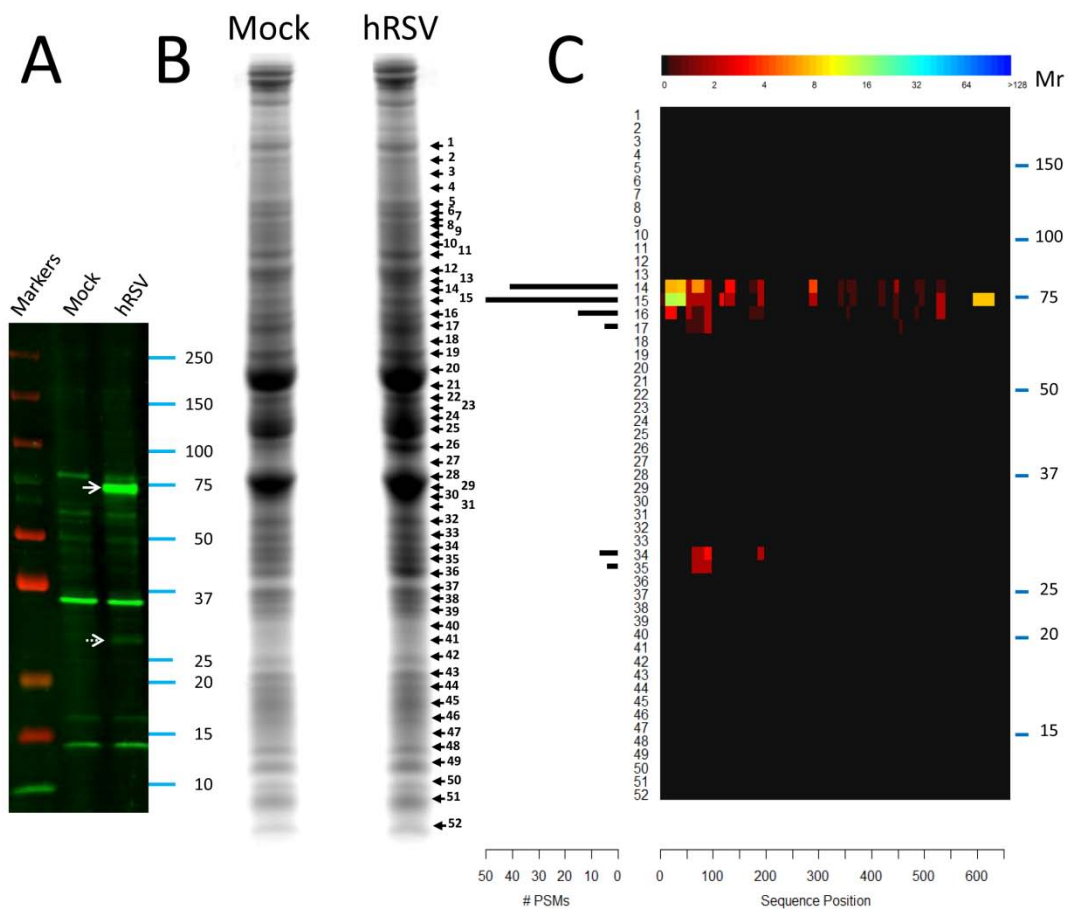


Fraction 23

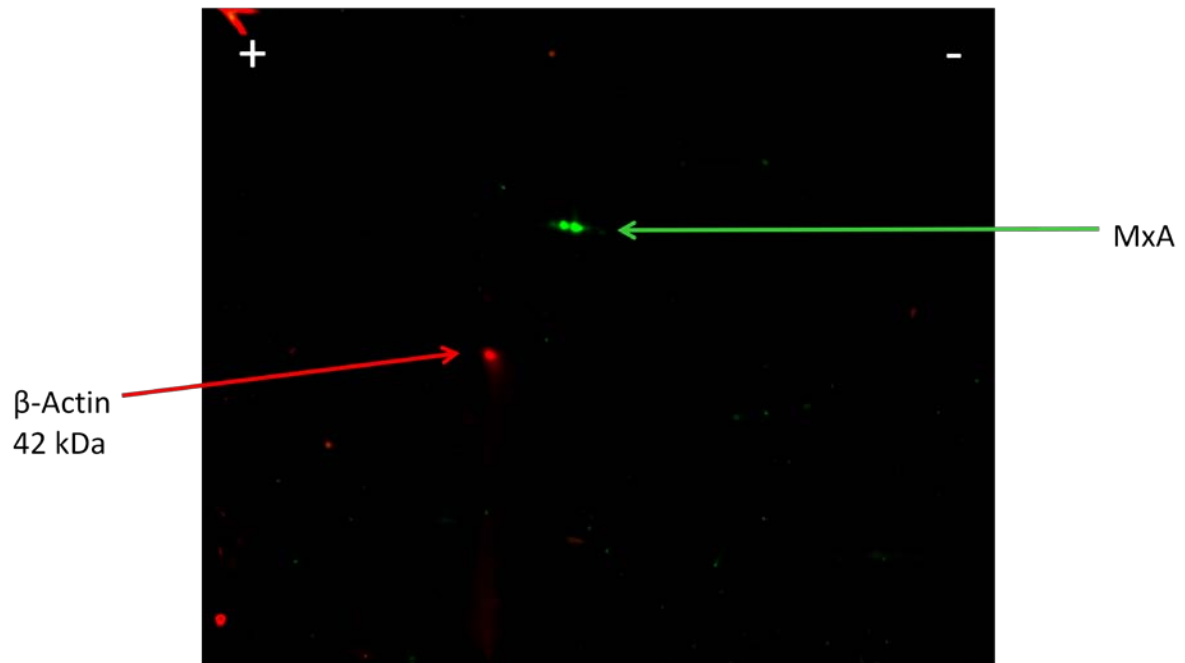


Fraction 24

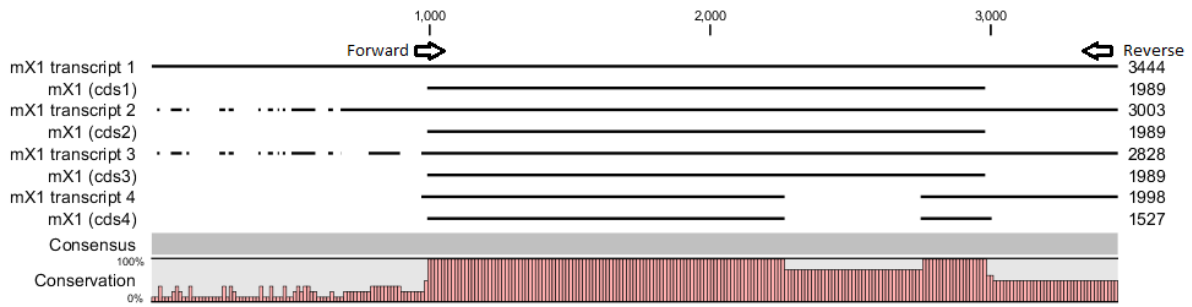




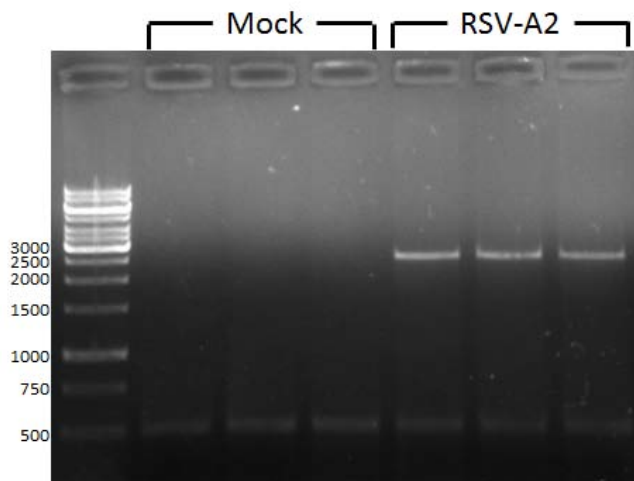
Supplemental Fig. 5. **Western blot and Protomap analyses of MxA proteoforms in uninfected A549 whole cell lysates.** (A) One-dimensional-Western blot of uninfected (Mock) and hRSV-infected (hRSV) A549 whole cell lysates. The solid white arrow indicates the mobility of the full-length canonical MxA proteoform. The broken white arrow indicates the mobility of the 29 kDa MxA proteoform. (B) Stained 1D-SDS-PAGE gels of uninfected (Mock) and hRSV-infected (hRSV) A549 cell lysates which were sliced according to the numbered arrows. (C) Whereas in-gel digests of hRSV-infected lysate slices revealed PSMs for MxA indicated in the PSM histograms, uninfected lysates produced no PSMs for MxA. The sequence coverage heat map (right) represents the alignment of all PSMs identified in the in-gel slices at each position of the canonical MxA proteoform sequence (P20591).



Supplemental Fig. 6. **Complete 2D--Western blot of an hRSV-Infected A549 cell lysate.** This blot was probed with an antibody to the N-terminal common to full length MxA and VarMxA.



Supplemental Fig. 7. **Alignment of potential MX1 transcripts.** Reported transcript variants with accession numbers, NM_001144925.2, NM_002462.4, NM_001178046.2, NM_001282920.1, and their respective coding sequences (cds) were aligned. Oligonucleotide primers were designed to cover the entire cds indicated by arrows.



Supplemental Fig. 8. **PCR analysis of MX1 mRNA transcripts.** mRNA was extracted from mock and A2 hRSV infected A549 cells at 24hrs post infection. mRNA was reverse transcribed to cDNA and PCR was performed with MX1 specific primers.

LOCUS mX1_(A549_RSV_induced)_CDS 2410 bp DNA linear UNA
19-Sep-2014
DEFINITION
FEATURES Location/Qualifiers
CDS 20..2005
/label=mX1
ORIGIN
1 TACTTTGCAA AGAAGGAAGA TGGTTGTTTC CGAAGTGGAC ATCGCAAAAAG CTGATCCAGC
61 TGCTGCATCC CACCCTCTAT TACTGAATGG AGATGCTACT GTGGCCCAGA AAAATCCAGG
121 CTCGGTGGCT GAGAACAACC TGTGCAGCCA GTATGAGGAG AAGGTGCGCC CCTGCATCGA
181 CCTCATTGAC TCCCTGCGGG CTCTAGGTGT GGAGCAGGAC CTGGCCCTGC CAGCCATCGC
241 CGTCATCGGG GACCAGAGCT CGGGCAAGAG CTCCGTGTTG GAGGCACTGT CAGGAGTTGC
301 CCTTCCCAGA GGCAGCGGGA TCGTGACCAG ATGCCCGCTG GTGCTGAAAC TGAAGAAACT
361 TGTGAACGAA GATAAGTGGA GAGGCAAGGT CAGTTACCAG GACTACGAGA TTGAGATTTT
421 GGATGCTTCA GAGGTAGAAA AGGAAATTA TAAAGCCCAG AATGCCATCA CCGGGGAAGG
481 AATGGGAATC AGTCATGAGC TAATCACCCCT GGAGATCAGC TCCCGAGATG TCCCGGATCT
541 GACTCTAATA GACCTTCCTG GCATAACCAG AGTGGCTGTG GGCAATCAGC CTGCTGACAT
601 TGGGTATAAG ATCAAGACAC TCATCAAGAA GTACATCCAG AGGCAGGAGA CAATCAGCCT
661 GGTGGTGGTC CCCAGTAATG TGGACATTGC CACCACAGAG GCTCTCAGCA TGGCCCAGGA
721 GGTGGACCCC GAGGGAGACA GGACCATCGG AATCTTGACG AAGCCTGATC TGGTGGACAA
781 AGGAACTGAA GACAAGGTTG TGGACGTGGT GCGGAACCTC GTGTTCCACC TGAAGAAGGG
841 TTACATGATT GTCAAGTGCC GGGGCCAGCA GGAGATCCAG GACCAGCTGA GCCTGTCCGA
901 AGCCCTGCAG AGAGAGAAGA TCTTCTTTGA GAACCACCCA TATTTAGGG ATCTGCTGGA
961 GGAAGGAAAG GCCACGGTTC CCTGCCTGGC AGAAAACTT ACCAGCGAGC TCATCACACA
1021 TATCTGTAAG TCTCTGCCCC TGTTAGAAAA TCAAATCAAG GAGACTCACC AGAGAATAAC
1081 AGAGGAGCTA CAAAAGTATG GTGTCGACAT ACCGGAAGAC GAAAATGAAA AAATGTTCTT
1141 CCTGATAGAT AAAATTAATG CCTTTAATCA GGACATCACT GCTCTCATGC AAGGAGAGGA
1201 AACTGTAGGG GAGGAAGACA TTCGGCTGTT TACCAGACTC CGACACGAGT TCCACAAATG
1261 GAGTACAATA ATTGAAAACA ATTTTCAAGA AGGCCATAAA ATTTTGAAGTA GAAAAATCCA
1321 GAAATTTGAA AATCAGTATC GTGGTAGAGA GCTGCCAGGC TTTGTGAATT ACAGGACATT
1381 TGAGACAATC GTGAAACAGC AAATCAAGGC ACTGGAAGAG CCGGCTGTGG ATATGCTACA
1441 CACCGTGACG GATATGGTCC GGCTTGCTTT CACAGATGTT TCGATAAAAA ATTTTGAAGA
1501 GTTTTTTAAAC CTCCACAGAA CCGCCAAGTC CAAAATTGAA GACATTAGAG CAGAACAAGA
1561 GAGAGAAGGT GAGAAGCTGA TCCGCTCCA CTTCAGATG GAACAGATTG TCTACTGCCA
1621 GGACCAGGTA TACAGGGGTG CATTGCAGAA GGTCCAGAGG AAGGAGCTGG AAGAAGAAAA
1681 GAAGAAGAAA TCCTGGGATT TTGGGGCTTT CCAATCCAGC TCGGCAACAG ACTCTTCCAT
1741 GGAGGAGATC TTTTCAGACC TGATGGCCTA TCACCAGGAG GCCAGCAAGC GCATCTCCAG
1801 CCACATCCCT TTGATCATCC AGTTCTTCAT GCTCCAGACG TACGGCCAGC AGCTTCAGAA
1861 GGCCATGCTG CAGCTCCTGC AGGACAAGGA CACCTACAGC TGGCTCCTGA AGGAGCGGAG
1921 CGACACCAGC GACAAGCGGA AGTTCCTGAA GGAGCGGCTT GCACGGCTGA CGCAGGCTCG
1981 GCGCCGGCTT GCCCAGTTCC CCGGTTAACC ACACTCTGTC CAGCCCCGTA GACGTGCACG
2041 CACACTGTCT GCCCCGTTT CCGGGTAGCC ACTGGACTGA CGACTTGAGT GCTCAGTAGT
2101 CAGACTGGAT AGTCCGTCTC TGCTTATCCG TTAGCCGTGG TGATTTAGCA GGAAGCTGTG
2161 AGAGCAGTTT GGTTTCTAGC ATGAAGACAG AGCCCCACCC TCAAATGCAC ATGAGCTGGC
2221 GGGGATGAAG GATGCTGTCT TCGTACTGGG AAAGGGATTT TCAGCCCTCA GAATCGCTCC
2281 ACCTTGACG TCTCCCCTTC TCTGTATTCC TAGAAACTGA CACATGCTGA ACATCACAGC
2341 TTATTTCTCT ATTTTTATAA TGTCCCTTCA CAAACCCAGT GTTTTAGGAG CATGAGTGCC
2401 GTGTGTGTGC

Supplemental Fig. 9. DNA sequence of the PCR product as described in Supplemental Figs. 7 and 8.


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LOCUS      mX1_(A549_RSV_induced)_CDS_protein      662 aa
linear     UNA
FEATURES             Location/Qualifiers
ORIGIN
   1 MVVSEVDIAK ADPAAASHPL LLNGDATVAQ KNPGSVAENN LCSQYEEKVR PCIDLIDSLR
  61 ALGVEQDLAL PAIAVIGDQS SGKSSVLEAL SGVALPRGSG IVTRCPLVLK LKKLVNEDKW
121 RGKVSQDYDYE IEISDASEVE KEINKAQNAI TGEGMGISHE LITLEISSRD VPDLTLLIDL
181 GITRVAVGNO PADIGYKIKT LIKKYIQRQE TISLVVPSN VDIATTEALS MAQEVDPEDG
241 RTIGILTKPD LVDKGTEDKV VDVVRNLVFN LKKGYMIVKC RGQQEIQDQL SLSEALQREK
301 IFFENHPYFR DLLEEGKATV PCLAEKLTSE LITHICKSLP LLENQIKETH QRITEELQKY
361 GVDIPEDENE KMFFLIDKIN AFNQDITALM QGEETVGEED IRLFTRLRHE FHKWSTIEN
421 NFQEGHKILS RKIQKFENQY RGRELPGFVN YRTFETIVKQ QIKALEEPAV DMLHTVTDMV
481 RLAFTDVSIK NFEEFFNLHR TAKSKIEDIR AEQEREGEKL IRLHFQMEQI VYQDQVYRG
541 ALQKVREKEL EEEKKKKSVD FGAFQSSSAT DSSMEEIFQH LMAYHQEASK RISSHIPLII
601 QFFMLQTYGQ QLQKAMLQLL QDKDTYSWLL KERSDTSDKR KFLKERLARL TQARRRLAQF
661 PG

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Supplemental Fig.10. **MxA protein sequence**. Complete amino acid sequence translated from the DNA sequence presented in Supplemental Fig. 9.