

## 1 **Repository Text**

## 2 **Methods**

### 3 **Cell lines and Transfections**

4 IL-6R expressing Cos-7 (Origene) cells and U3A cells (provided by G. Stark, Cleveland  
5 Clinic, Ohio), were used. The cells were maintained in complete Dulbecco modified Eagle  
6 medium (DMEM; Life technologies, NY) supplemented with 10% FCS, 2mM L-glutamine,  
7 HEPES and antibiotics. Transient transfections were carried out using either electroporation  
8 (Lonza, Germany) or Lipofectamine 2000® transfection reagent (Life technologies, NY)  
9 following manufacturers' protocol.

### 10 **Plasmids and constructs**

11 Plasmid encoding Myc-DDK tagged wild type STAT3 transcript variant 1 was purchased  
12 from Origene Technologies, MD, USA (RC215836). Point mutations were introduced in the wild  
13 type sequences using point mutagenesis PCR and the resulting PCR products were cloned into  
14 the pCM6-XL4 background (BioInnovatise, Rockville, MD). Endotoxin-free plasmid  
15 preparations were made using Endo Free plasmid kits purchased from Qiagen (Germany).

### 16 **STAT3 phosphorylation assay**

17 Cells transfected with plasmids encoding wild type or mutant *STAT3* were stimulated with  
18 oncostatin M (OSM; 50ng/ml) or IL-6 in increasing concentrations (20ng, 100ng, 200ng,  
19 1,000ng) for 30 min at 37°C. Cells were then fixed with 4% paraformaldehyde (Electron  
20 Microscopy Sciences, PA), permeabilized with 100% methanol and stained for STAT3 (BD  
21 APC) and phosphoSTAT3 (BD AF 488, pY705). Fold induction of phosphoSTAT3 (Y705) in  
22 OSM or IL-6 stimulated cells over non-stimulated cells was calculated using the geometric mean

23 of the fluorescent values and the fluorescent values are denoted as MFI values (Mean  
24 Fluorescent Intensity).

25 All data were acquired on either FACS Calibur or BD LSR Fortessa (BD Biosciences). A  
26 minimum of 50,000 counts were acquired within the live cell population gate for each tube. Data  
27 were analyzed using FlowJo version 9 (Tree Star Inc.).

### 28 **Protein Models**

29 The 3D models of STAT3 were generated using I-TASSER server. The electrostatic potential  
30 was calculated with APBS and displayed using UCSF Chimera.

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32 **References**

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77 **Supplementary figure legend**

78 **Figure E1**

79 Previously reported *STAT3* mutations by *STAT3* domain. Hypomorphic dominant negative  
80 mutations found in HIES (Job's) patients are indicated in purple; hypermorphic gain of function  
81 mutations found in leukemia or lymphoma patients are in green; hypermorphic gain of function  
82 mutations found in autoimmune disorders are in orange.

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**Figure E1**

