

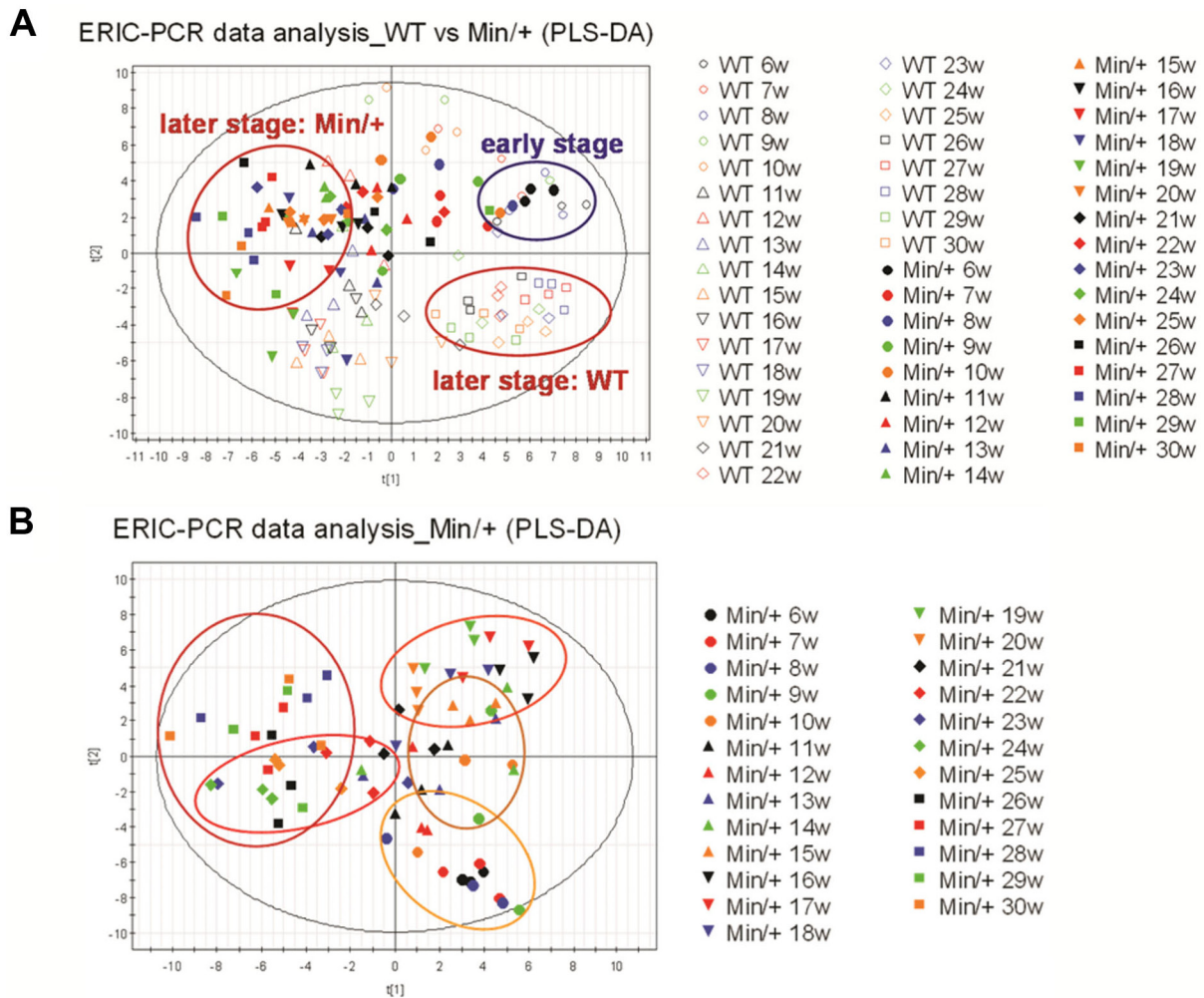
## SUPPLEMENTARY DATA

## REFERENCES

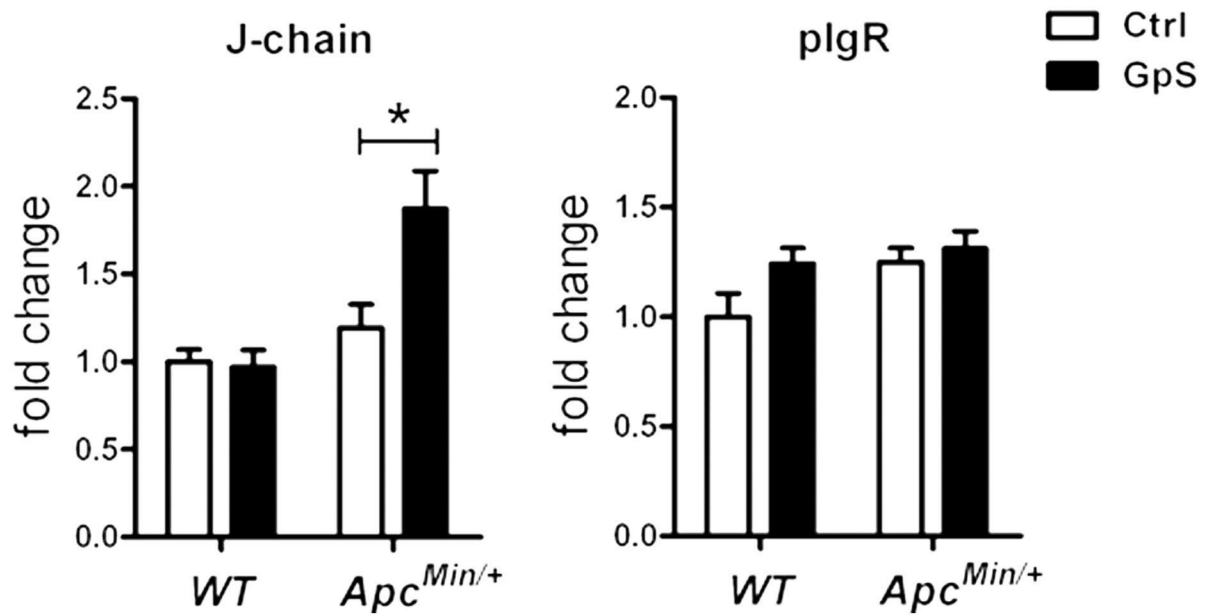
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Supplementary Figure S1: A bar chart of the relative abundance of bacterial genera in the individual mice of different treatment groups.



**Supplementary Figure S2: The time course PLS-DA plots of ERIC-PCR DNA profile.** **A.** *Apc*<sup>Min/+</sup> mice versus WT. Open symbols: WT; Solid symbols: *Apc*<sup>Min/+</sup> mice. **B.** *Apc*<sup>Min/+</sup> mice at different disease stages. Different symbol shapes represented for different disease stages. The microbial profiles of *Apc*<sup>Min/+</sup> mice and their WT littermates were characterized from 6 weeks to 30 weeks of age for 25 consecutive weeks. The fecal samples were prepared for genomic DNA and subjected for ERIC-PCR. Gel pictures were digitized by Image Lab 3.0 system (Bio-Rad). Based on the distance and the intensity of each DNA band, SIMCA-P 12.0 tool was applied to obtain the PLS-DA score plots. WT: n=3; *Apc*<sup>Min/+</sup> mice: n=3.



**Supplementary Figure S3: The relative mRNA expression of J-chain and plgR.** J-chain is known to be critical for polymeric immunoglobulin receptor (pIgR)-mediated epithelial transport of IgA. Data were normalized to the expression of reference gene, and expressed as fold change of the WT control group. Data is presented as the mean  $\pm$  SEM (\* P < 0.05, GpS versus control samples); n=6/group.

Supplementary Table S1: Primers for Quantitative Real-Time PCR

Target gene	Nucleotide sequence of primers (5' to 3')		References
	Forward	Reverse	
<i>β-actin</i>	TGTTACCAACTGGGACGACA	CTGGGTCACTTTTCACGGT	[1]
<i>Hprt1</i>	TCAGTCAACGGGGACATAAA	GGGGCTGTACTGCTTAACCAG	[2]
<i>dsrA</i>	CTGCGAATATGCCTGCTACA	TGGTCGARCTTGATGTCGTC	[3]
<i>PanCrp</i>	GGTGATCATCAGACCCCAGCATCAGT	AAGAGACTAAAAGTGGAGCAGC	[4]
<i>Lyz 1</i> ( <i>P-lysozyme</i> )	GCCAAGGTCTACAATCGTTGTGAGTTG	CAGTCAGCCAGCTTGACACCACG	[5]
<i>Lyz 2</i> ( <i>M-lysozyme</i> )	GGCTGGCTACTATGGAGTCAGCCTG	GCATTCACAGCTCTGGGGTTTTG	[5]
<i>MUC2</i>	CCCAGAAGGGACTGTGTATG	TTGTGTTGCTCTTGGTCAG	[6]
<i>MUC4</i>	GTCTCCCATCACGGTTCAGT	TGTCATTCCACACTCCCAGA	[6]
<i>J-chain</i>	GAACCTTTGTATAACCATTTGTCAGACG	CTGGGTGGCAGTAACAACCT	[6]
<i>pIgR</i>	AGTAACCGAGGCCTGTCCTT	GTCACTCGGCAACTCAGGA	[6]
<i>iNOS</i>	GTTCTCAGCCCAACAATACAAGA	GTGGACGGGTCGATGTCAC	[7]
<i>CXCL10</i>	CCAAGTGCTGCCGTCATTTTC	GGCTCGCAGGGATGATTTCAA	[7]
<i>Arginase 1</i>	TGGCTTGCGAGACGTAGAC	GCTCAGGTGAATCGGCCTTTT	[7]
<i>Ym1</i>	TTATCCTGAGTGACCCTTCTAAG	TCATTACCCTGATAGGCATAGG	[7]
<i>Trem2</i>	CTGGAACCGTCACCATCACTC	CGAAACTCGATGACTCCTCGG	[7]
<i>MR</i>	GCTGAATCCCAGAAATTCCGC	ATCACAGGCATACAGGGTGAC	[7]
<i>IFN-γ</i>	ATGAACGCTACACACTGCATC	CCATCCTTTTGCCAGTTCCTC	[7]
<i>TNF-α</i>	CAAATGGCCTCCCTCTCAT	CTCCTCCACTTGGTGGTTTG	[8]
<i>IL-12</i>	ACTCTGCGCCAGAAACCTC	CACCCTGTTGATGGTCACGAC	[7]
<i>IL-4</i>	GGTCTCAACCCCCAGCTAGT	GCCGATGATCTCTCTCAAGTGAT	[7]
<i>IL-10</i>	GCTCTTACTGACTGGCATGAG	CGCAGCTCTAGGAGCATGTG	[7]
<i>IL-13</i>	GGATATTGCATGGCCTCTGTAAC	AACAGTTGCTTTGTGTAGCTGA	[7]
<i>IL-1β</i>	GCTGAAAGCTCTCCACCTCA	GGCCACAGGTATTTTGTTCGT	[8]
<i>IL-6</i>	CTTCCATCCAGTTGCCTTCTTG	AATTAAGCCTCCGACTTGTGAAG	[9]
<i>IL-18</i>	CCTGACATCTTCTGCAACCT	TTCCGTATTACTGCGGTTGT	[10]
<i>IL-23</i>	GCACCTGCTTGACTCTGACA	ATCCTCTGGCTGGAGGAGTT	[8]
<i>Foxp3</i>	CCCATCCCCAGGAGTCTTG	ACCATGACTAGGGGCACTGTA	[11]
<i>IL-17</i>	TTAACTCCCTTGGCGCAAAA	CTTCCCTCCGATTGACAC	[11]