

## **Whole Chromosome Instability induces senescence and promotes SASP**

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**Supplementary information**

**Supplementary Tables:**

**Table S1. Distribution of ploidy changes observed during replicative senescence and total number of cells analyzed for each cell line.**

	<b>Total %Aneuploid</b>	<b>%4n</b>	<b>%3n</b>	<b>%1n</b>	<b>%&gt;5n</b>	<b>%2n</b>	<b>%Non-diploid</b>	<b>Cells counted</b>
<b>HPF PD42</b>	4.0	4.5	0.5	0.2	0.7	90.1	9.9	554
<b>HPF PD56</b>	10.0	7.5	1.1	0.4	1.4	79.6	20.4	279
<b>HPF PD74</b>	22.9	26.6	1.5	0.0	11.1	37.8	62.2	458
<b>MEF PD04-1</b>	5.1	30.3	0.0	0.0	0.0	64.6	35.4	99
<b>MEF PD04-2</b>	2.0	42.6	0.0	0.0	1.0	54.5	45.5	101
<b>MEF PD10-1</b>	19.6	36.1	2.1	0.0	9.3	33.0	67.0	97
<b>MEF-PD10-2</b>	16.1	36.6	0.0	0.0	6.5	40.9	59.1	93

**Table S2. Distribution of ploidy changes detected in each cell line per biological replicate (1, 2 and 3), and frequency BrdU positive cells as detected by immunofluorescence.**

	<b>%Aneuploid</b>	<b>%4n</b>	<b>%3n</b>	<b>%1n</b>	<b>%&gt;5n</b>	<b>%2n</b>	<b>%Non-diploid</b>	<b>% BrdU</b>
<b>EV1</b>	1.9	2.9	0.5	0.0	0.5	94.2	5.8	83.1
<b>EV2</b>	0.0	3.0	0.0	0.0	0.0	97.0	3.0	82.1
<b>EV3</b>	5.0	5.0	0.0	0.0	1.0	88.9	11.1	72.4
<b>shB1-1</b>	6.3	8.9	0.0	1.6	1.0	82.3	17.7	72.9
<b>shB1-2</b>	1.5	10.4	0.0	0.0	0.0	88.1	11.9	72.3
<b>shB1-3</b>	18.1	4.5	1.0	0.0	0.5	75.9	24.1	81.9
<b>shB2-1</b>	6.5	9.7	0.0	0.0	0.0	83.9	16.1	61.3
<b>shB2-2</b>	12.4	9.4	0.5	0.0	3.0	74.8	25.2	64.4
<b>shB2-3</b>	3.6	14.3	0.5	0.0	2.0	79.6	20.4	68.4
<b>shS1-1</b>	18.6	11.3	2.0	0.5	1.5	66.2	33.8	51.0
<b>shS1-2</b>	17.9	18.4	2.0	0.0	3.5	58.2	41.8	52.5
<b>shS1-3</b>	4.0	17.4	0.5	0.0	1.0	77.1	22.9	49.8
<b>shS2-1</b>	22.3	11.4	2.6	0.5	3.1	60.1	39.9	37.3
<b>shS2-1</b>	20.9	18.4	2.5	0.0	2.5	55.7	44.3	38.5
<b>shS2-3</b>	10.0	23.0	1.0	0.0	1.0	65.0	35.0	39.0
<b>SEN1</b>	9.2	19.5	0.5	0.0	1.5	69.2	30.8	19.8
<b>SEN2</b>	10.1	23.1	0.5	0.5	1.9	63.9	36.1	21.6



**Table S4. Values of the SASP factors secreted 2 fold or more by shS2 cells relative to EV detected in the preliminary SASP screen. For each sample, the average and standard deviation (s.d.) concentration values for each component (pg/ml) are shown. The results are listed in order of fold difference expression, indicating the *p* values for each SASP component (generated from a t-test analysis between EV and shS2 values – n=2). Official gene symbol is followed by previous gene symbol and spelled full names within brackets.**

Gene	EV Average (pg/ml)	EV (s.d.)	shS2 Average (pg/ml)	shS2 (s.d.)	Fold difference	Significant? (alpha<0.05)	<i>p</i> value (two-tailed)
<i>CXCL8/IL-8</i> (C-X-C motif chemokine ligand 8)	7.0	9.8	1917.9	1170.9	273.3	No	0.147
<i>CXCL10/IP-10</i> (C-X-C motif chemokine ligand 10)	0	0	96.0	55.6	96.0	No	0.135
<i>CCL2/MCP-1</i> (C-C motif chemokine ligand 2)	4.9	6.9	156.6	26.6	32.2	<u>Yes</u>	<u>0.016</u>
<i>IL1B/IL-1<math>\beta</math></i> (Interleukin 1 beta)	0.1	0	0.9	0.2	16.7	<u>Yes</u>	<u>0.030</u>
<i>TNF/TNF-<math>\alpha</math></i> (Tumor necrosis factor)	0.6	0.1	8.8	0.8	14.8	<u>Yes</u>	<u>0.005</u>
<i>CXCL1/GRO<math>\alpha</math></i> (C-X-C motif chemokine ligand 1)	10.1	14.3	121.9	172.5	12.1	No	0.457
<i>CSF2/GM-CSF</i> (Colony stimulating factor 2)	2.1	3.0	20.2	4.4	9.6	<u>Yes</u>	<u>0.041</u>
<i>IL6/IL-6</i> (Interleukin 6)	2.5	3.1	23.1	10.3	9.4	No	0.114
<i>CCL27/CTACK</i> (C-C motif chemokine ligand 27)	0.9	1.3	7.1	4.6	7.8	No	0.208
<i>CXCL12/SDF-1<math>\alpha</math></i> (C-X-C motif chemokine ligand 12)	12.2	17.3	68.2	11.3	5.6	No	0.062
<i>IL15/IL-15</i> (Interleukin 15)	0.3	0	1.1	0.8	4.0	No	0.293
<i>IL12B</i> (Interleukin 12B)	4.1	5.5	13.7	2.4	3.3	No	0.152
<i>MIF</i> (Macrophage migration inhibitory factor)	89.2	11.8	292.1	228.1	3.3	No	0.336

<i>NGF/b-NGF</i> (Nerve growth factor)	1.4	0.2	4.5	1.6	3.2	No	0.113
<i>CLEC11A/SCGF-b</i> (C-type lectin domain family 11 member A)	135.8	91.1	422.0	147.6	3.1	No	0.145
<i>IL10/IL-10</i> (Interleukin 10)	1.1	1.6	3.0	4.2	2.6	No	0.611
<i>KITLG/SCF</i> (KIT ligand)	1.1	0.9	2.5	0.1	2.2	No	0.160

**Table S5. Validation of W-CIN-induced SASP components. The secreted factors that were tested are listed in alphabetical order and indicated as the concentration (pg/ml) measured in the conditioned medium of EV, Bub1-, SMC1A-depleted and SEN cells, respectively. For each sample, the average and standard deviation (s.d.) concentration values for each component are shown. The fold differences relative to control EV cells, as well as the *p* value generated from One-way ANOVA analysis comparing all samples to EV are also shown.**

		Gene	EV Average (pg/ml)	EV (s.d.)	shB1 Average (pg/ml)	shB1 (s.d.)	Fold difference relative to EV	Significant? (alpha<0.05)	<i>p</i> value (two-tailed)
		<b>BUB1-depleted</b>		<b>shB1</b>					
<i>CCL2 (MCP-1)</i>	4.402			1.471	37.002	17.269	8.4	No	0.4819
<i>CCL27 (CTACK)</i>	0.467			0.121	1.632	0.527	3.5	No	0.3736
<i>CLEC11A (SCGF-b)</i>	376.129			45.857	835.251	162.340	2.2	No	0.5198
<i>CSF2 (GM-CSF)</i>	0.631			0.212	0.901	0.061	1.4	No	0.763
<i>CXCL1 (GRO<math>\alpha</math>)</i>	1.215			0.645	0.531	0.132	0.4	No	0.8221
<i>CXCL10 (IP-10)</i>	0.049			0.012	0.231	0.291	4.8	No	0.8554
<i>CXCL12 (SDF-1<math>\alpha</math>)</i>	4.014			0.960	6.037	3.008	1.5	No	0.5837
<i>CXCL8 (IL8)</i>	0.744			0.236	5.215	3.821	7.0	No	0.9427
<i>IL10</i>	0.074			0.080	0.173	0.074	2.3	No	0.841
<i>IL1B</i>	0.001			0.000	0.002	0.001	1.4	No	0.994
<i>IL6</i>	2.734			1.551	23.598	16.623	8.6	No	0.1891
<i>MIF</i>	3.055			0.854	9.938	3.504	3.3	No	0.4388
<i>TNF</i>	0.053	0.021	0.161	0.046	3.1	No	0.2379		
<b>shB2</b>		Gene	EV Average (pg/ml)	EV (s.d.)	shB2 Average (pg/ml)	shB2 (s.d.)	Fold difference relative to EV	Significant? (alpha<0.05)	<i>p</i> value (two-tailed)
		<i>CCL2 (MCP-1)</i>	4.402	1.471	35.866	15.600	8.1	No	0.497
		<i>CCL27 (CTACK)</i>	0.467	0.121	2.565	1.973	5.5	No	0.2086
		<i>CLEC11A (SCGF-b)</i>	376.129	45.857	1817.459	570.667	4.8	No	0.0786
		<i>CSF2 (GM-CSF)</i>	0.631	0.212	0.937	0.921	1.5	No	0.6293

	<i>CXCL1 (GRO<math>\alpha</math>)</i>	1.215	0.645	0.920	0.786	0.8	No	0.9155
	<i>CXCL10 (IP-10)</i>	0.049	0.012	0.193	0.235	4.0	No	0.8821
	<i>CXCL12 (SDF-1<math>\alpha</math>)</i>	4.014	0.960	4.672	4.409	1.2	No	0.8523
	<i>CXCL8 (IL8)</i>	0.744	0.236	3.320	2.300	4.5	No	0.9761
	<i>IL10</i>	0.074	0.080	0.531	0.720	7.2	No	0.4666
	<i>IL1B</i>	0.001	0.000	0.001	0.001	1.1	No	0.9989
	<i>IL6</i>	2.734	1.551	9.155	5.749	3.3	No	0.693
	<i>MIF</i>	3.055	0.854	11.115	1.574	3.6	No	0.4058
	<i>TNF</i>	0.053	0.021	0.319	0.149	6.0	<u>Yes</u>	<u>0.0201</u>



		Gene	EV Average (pg/ml)	EV (s.d.)	shS1 Average (pg/ml)	shS1 (s.d.)	Fold difference relative to EV	Significant? (alpha<0.05)	p value (two-tailed)		
		SMC1A -depleted		shS1		<i>CCL2 (MCP-1)</i>	4.402	1.471	86.744	54.639	19.7
<i>CCL27 (CTACK)</i>	0.467			0.121	2850.731	1827.076	6108.9	No	0.308		
<i>CLEC11A (SCGF-b)</i>	376.129			45.857	2.143	3.027	0.0	<u>Yes</u>	0.0132		
<i>CSF2 (GM-CSF)</i>	0.631			0.212	0.038	0.050	0.1	No	0.5099		
<i>CXCL1 (GRO<math>\alpha</math>)</i>	1.215			0.645	1.235	1.755	1.0	No	0.9944		
<i>CXCL10 (IP-10)</i>	0.049			0.012	3.903	4.450	80.2	<u>Yes</u>	0.0079		
<i>CXCL12 (SDF-1<math>\alpha</math>)</i>	4.014			0.960	9.450	12.376	2.4	No	0.2625		
<i>CXCL8 (IL8)</i>	0.744			0.236	24.331	30.952	32.7	No	0.7324		
<i>IL10</i>	0.074			0.080	0.219	0.374	3.0	No	0.7939		
<i>IL1B</i>	0.001			0.000	0.002	0.002	1.5	No	0.993		
<i>IL6</i>	2.734			1.551	18.527	15.946	6.8	No	0.3743		
<i>MIF</i>	3.055			0.854	23.326	7.588	7.6	<u>Yes</u>	0.0499		
<i>TNF</i>	0.053			0.021	0.089	0.015	1.7	No	0.7494		
shS2		Gene	EV Average (pg/ml)	EV (s.d.)	shS2 Average (pg/ml)	shS2 (s.d.)	Fold difference relative to EV	Significant? (alpha<0.05)	p value (two-tailed)		
		<i>CCL2 (MCP-1)</i>	4.402	1.471	245.152	166.934	55.7	<u>Yes</u>	0.0002		
		<i>CCL27 (CTACK)</i>	0.467	0.121	5.090	4.568	10.9	<u>Yes</u>	0.0142		
		<i>CLEC11A (SCGF-b)</i>	376.129	45.857	3092.486	1422.508	8.2	<u>Yes</u>	0.0042		
		<i>CSF2 (GM-CSF)</i>	0.631	0.212	3.202	2.416	5.1	<u>Yes</u>	0.0044		
		<i>CXCL1 (GRO<math>\alpha</math>)</i>	1.215	0.645	17.205	12.003	14.2	<u>Yes</u>	0.0001		
		<i>CXCL10 (IP-10)</i>	0.049	0.012	6.090	1.767	125.1	<u>Yes</u>	0.0003		
		<i>CXCL12 (SDF-1<math>\alpha</math>)</i>	4.014	0.960	54.133	10.738	13.5	<u>Yes</u>	< 0.0001		
		<i>CXCL8 (IL8)</i>	0.744	0.236	310.829	273.745	418.0	<u>Yes</u>	0.0008		

		<i>IL10</i>	0.074	0.080	1.470	1.881	19.9	<u>Yes</u>	0.0432
		<i>IL1B</i>	0.001	0.000	0.699	0.593	523.0	<u>Yes</u>	< 0.0001
		<i>IL6</i>	2.734	1.551	63.525	51.904	23.2	<u>Yes</u>	0.0033
		<i>MIF</i>	3.055	0.854	60.870	31.504	19.9	<u>Yes</u>	< 0.0001
		<i>TNF</i>	0.053	0.021	0.484	0.272	9.2	<u>Yes</u>	0.001

	Gene	EV Average (pg/ml)	EV (s.d.)	SEN Average (pg/ml)	SEN (s.d.)	Fold difference relative to EV	Significant? (alpha<0.05)	p value (two-tailed)
<b>SEN</b>	<i>CCL2 (MCP-1)</i>	4.402	1.471	277.703	192.630	63.090	<u>Yes</u>	0.0009
	<i>CCL27 (CTACK)</i>	0.467	0.121	2.315	1.160	4.961	No	0.1349
	<i>CLEC11A (SCGF-b)</i>	376.129	45.857	1443.024	300.854	3.837	No	0.0815
	<i>CSF2 (GM-CSF)</i>	0.631	0.212	6.406	1.129	10.148	<u>Yes</u>	< 0.0001
	<i>CXCL1 (GRO<math>\alpha</math>)</i>	1.215	0.645	125.527	90.238	103.336	<u>Yes</u>	0.0001
	<i>CXCL10 (IP-10)</i>	0.049	0.012	26.977	15.297	554.233	<u>Yes</u>	< 0.0001
	<i>CXCL12 (SDF-1a)</i>	4.014	0.960	64.369	38.901	16.035	<u>Yes</u>	0.0002
	<i>CXCL8 (IL8)</i>	0.744	0.236	386.169	164.381	519.378	<u>Yes</u>	< 0.0001
	<i>IL10</i>	0.074	0.080	0.482	0.242	6.519	No	0.3463
	<i>IL1B</i>	0.001	0.000	5.190	2.873	3882.364	<u>Yes</u>	< 0.0001
	<i>IL6</i>	2.734	1.551	1643.687	746.793	601.275	<u>Yes</u>	< 0.0001
	<i>MIF</i>	3.055	0.854	24.642	8.639	8.065	<u>Yes</u>	0.0111
	<i>TNF</i>	0.053	0.021	0.743	0.448	14.106	<u>Yes</u>	0.0008

**Table S6. Correlation analysis between the percentages of not 2n cells and secretion levels of individual SASP components (CLEC11A, CCL2, CCL27 and MIF) including the Pearson correlation coefficient (*r*) and *p* values that were found statistically significant. The average percentage of not 2n cells in each sample (X axis) is plotted against the average levels of each factor analyzed (Y axis).**

Sample	X axis		Y axis							
	% Average Not 2n cells	% Not 2n cells (s.d.)	Average CLEC11A (pg/ml)	CLEC11A (s.d.)	Average CCL2 (pg/ml)	CCL2 (s.d.)	Average CCL27 (pg/ml)	CCL27 (s.d.)	Average MIF (pg/ml)	MIF (s.d.)
EV	6.6	4.1	376.1	45.9	4.4	1.5	0.5	0.1	3.1	0.9
shB1	17.9	6.1	835.3	162.3	37.0	17.3	1.6	0.5	9.9	3.5
shB2	20.6	4.6	1817.5	570.7	35.9	15.6	2.6	2.0	11.1	1.6
shS1	32.8	9.5	2850.7	1827.1	86.7	54.6	2.1	3.0	23.3	7.6
shS2	39.7	4.6	3092.5	1422.5	245.2	166.9	5.1	4.6	60.9	31.5
SEN	33.4	3.7	1443.0	300.9	277.7	192.6	2.3	1.2	24.6	8.6
<b>Pearson correlation coefficient (<i>r</i>)</b>			<b>0.8619</b>		<b>0.8318</b>		<b>0.8221</b>		<b>0.8534</b>	
<b><i>p</i> value (two-tailed)</b>			<b>0.0273</b>		<b>0.0401</b>		<b>0.0447</b>		<b>0.0306</b>	
<b>Significant? alpha&lt;0.05</b>			<b>Yes</b>		<b>Yes</b>		<b>Yes</b>		<b>Yes</b>	

**Table S7. shRNA sequences of the targets used in this study.**

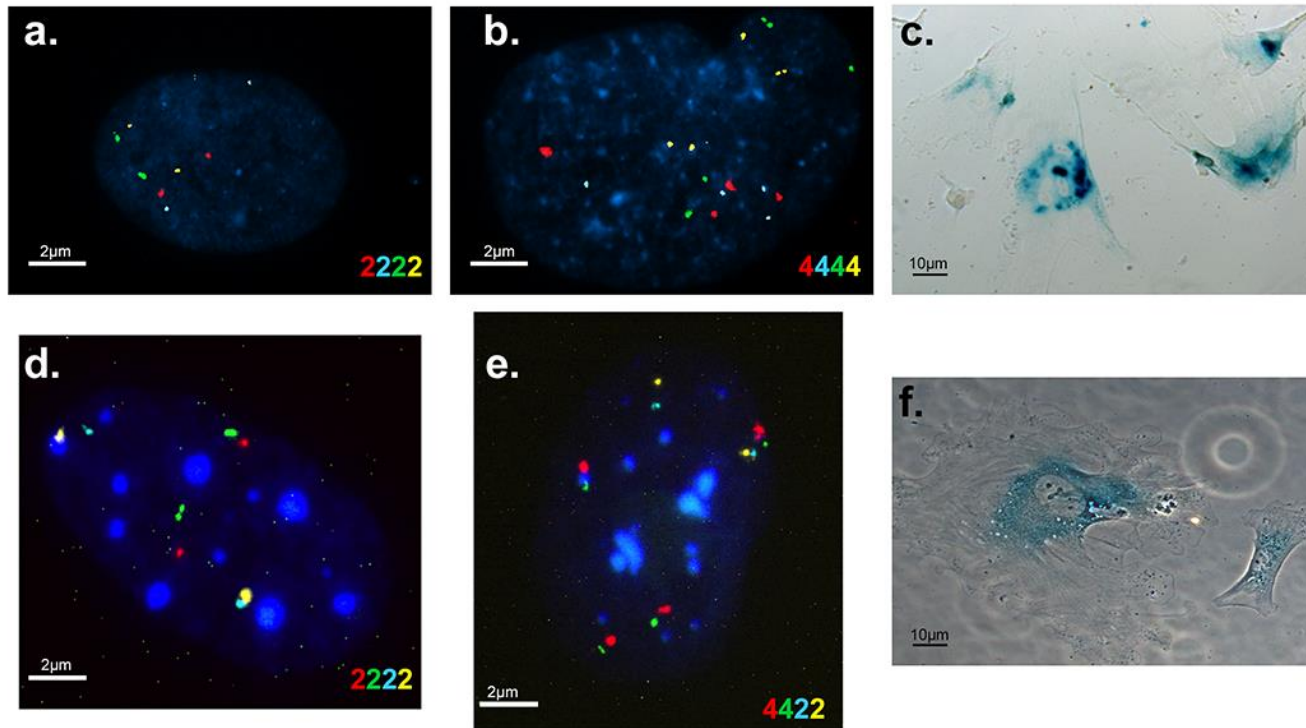
<b>Target gene</b>	<b>shRNA ID</b>	<b>Sense sequence 5' - 3'</b>
BUB1	shB1	CCTGGGTCAGAGTATAGATAT
BUB2	shB2	TACAACAGTGACCTCCATCAA
BUB3	shB3	CGAGGTTAATCCAGCACGTAT
BUB4	shB4	AGAAATACAATCAACGGAGAA
SMC1A	shS1	GCCGGGACTGTATTCAGTATA
SMC1A	shS2	CCAACATTGATGAGATCTATA
SMC1A	shS3	CCAACAAGGAAATGACCCATT
SMC1A	shS4	CGACAGATTATCGGACCATTT

**Table S8. Sequences of the human primers used for qRT-PCR.**

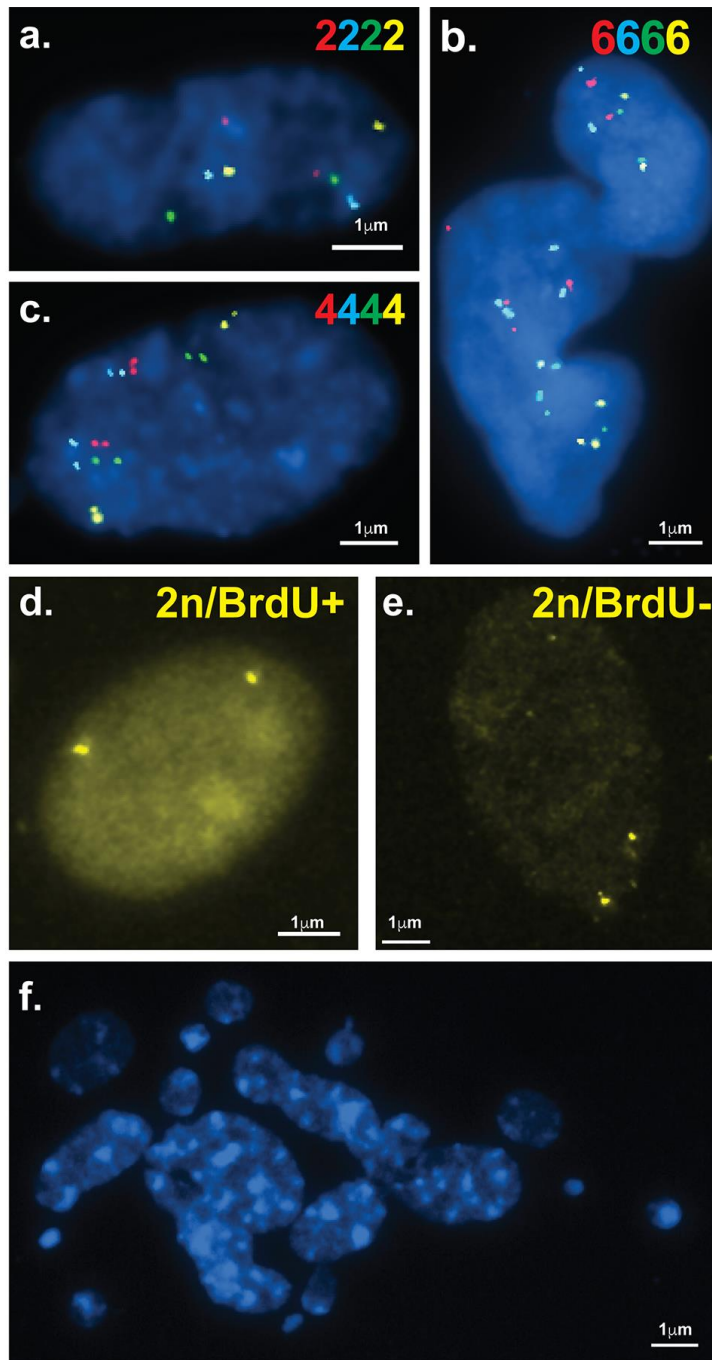
<b>Gene</b>	<b>Fwd primer 5'-3'</b>	<b>Rev primer 5'-3'</b>	<b>TaqMan probe</b>
<b>BUB1</b>	GGTGGTGCCTTCAAGGGATGG	GCAGCAGGCTGGCTCAGACG	SYBR green detection
<b>SMC1A</b>	AGGCCCTCATTGAGATTGAC	GCAATACGCTGAAGCACACT	SYBR green detection
<b>BUB3</b>	CGCATGCCTGGAGTGGAGGA	GGGGCATCATGGGTCCCAAC	SYBR green detection
<b>BUB1B</b>	CGTGGCAATACAGCTTCACT	AGGCTTTCTGGTGCTTAGGA	SYBR green detection
<b>SOD1</b>	TCAGGAGACCATTGCATCAT	GAATGTTTATTGGGCGATCC	SYBR green detection
<b>GPX1</b>	TTGACATCGAGCCTGACATC	ACTGGGATCAACAGGACCAG	SYBR green detection
<b>GAPDH</b>	TCAAGAAGGTGGTGAAGCAG	CGCTGTTGAAGTCAGAGGAG	CCTCAAGGGCATCCTGGGCT
<b>CLEC11A</b>	GGCTAGTGACCTGCACACAG	GCACCCAAAGTTCCAGATGT	CTCGGCAGTTCCAGAGGCC

## Supplementary figures

**Fig S1. Primary mammalian cells *in vitro* show accumulation of ploidy changes as they approach SEN. (a, b, d and e)** Representative images of four-color interphase-FISH in HPFs (a and b) and MEFs (d and e) showing diploid (a and d), tetraploid (b) and aneuploid nuclei (e). (c and f) Bright field representative images of senescent HPF (c) and MEFs (f) stained for SA- $\beta$ gal.

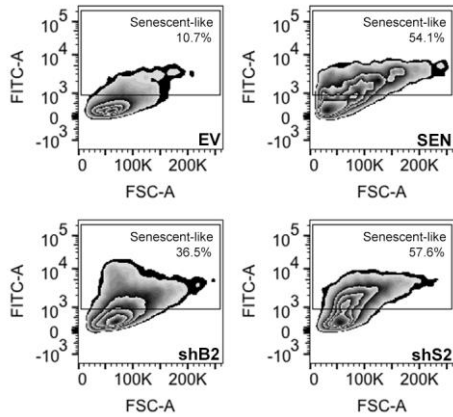


**Fig S2. Depletion of *BUB1* or *SMC1A* results in W-CIN, reduction of DNA replication and negligible apoptosis in human fibroblasts.** (a-c) Representative interphase FISH images showing the hybridization signals that were detected. The numbers of signals for each chromosome are indicated with the correspondent color of fluorophore used for staining. (a) Diploid nuclei (2222). (b) Nuclei containing 6 copies of each chromosome (6666), and morphology suggestive of abnormal mitosis. (c) Tetraploid nuclei (4444). Representative images of BrdU positive (d), and negative (e) nuclei containing 2 hybridization signals for chromosome 9 in the Cy5 channel. (f) Representative image of nuclei with fragmented appearance, suggestive of apoptotic bodies.

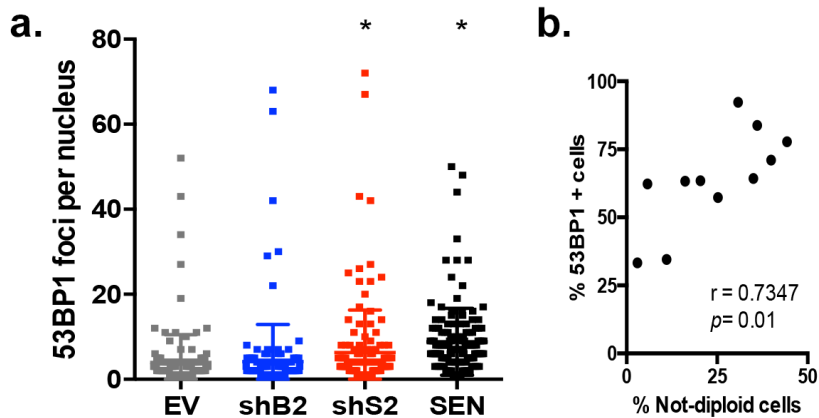




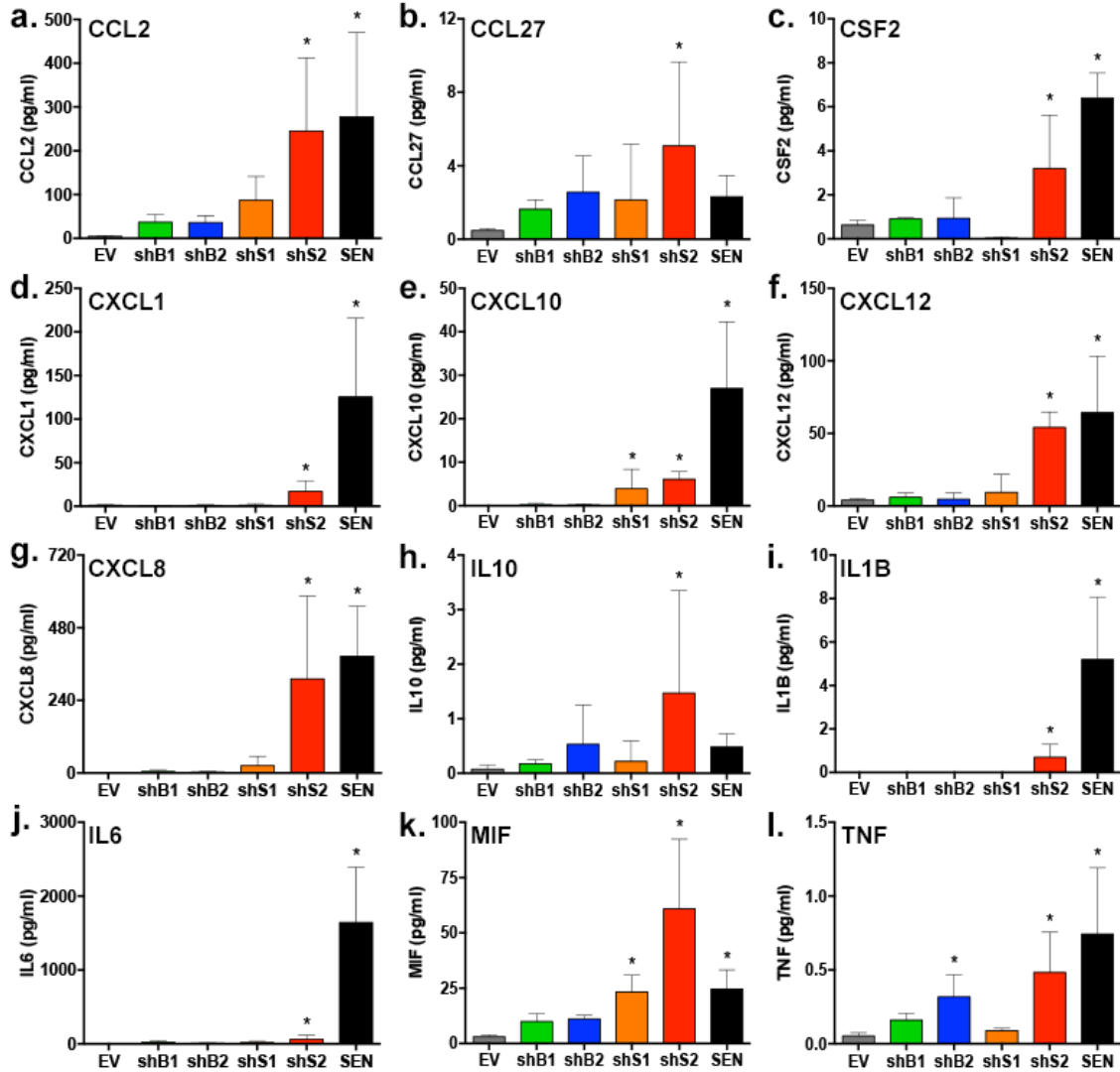
**Fig S3. Down-regulation of BUB1 and SMC1A shifts cells towards a senescent-like phenotype.** Representative FACS diagrams depicting the senescent-like population (i.e. high size and/or high AF) and respective percentage of these cells in each culture. Forward scatter channel (FSC-A) in X-axis depicts cell size and autofluorescence channel (FITC-A) in Y-axis depicts lipofuscin content. EV=empty vector; SEN= replicative senescence; shB2= BUB1-depleted; shS2= SMC1-depleted.



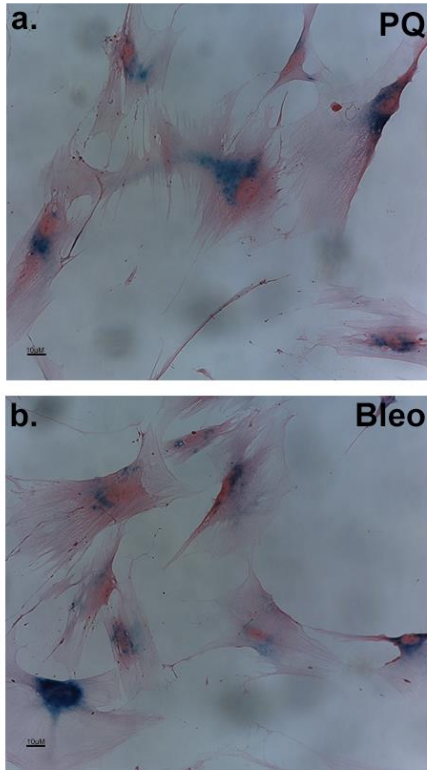
**Fig S4. W-CIN-induced senescence is associated with generation of DNA DSBs.** (a) Quantification of number of 53BP1 foci per nucleus in each culture. (b) Correlation plot between the percentages of not-diploid and 53BP1 foci containing cells. (\*) Indicates significant differences ( $p < 0.05$ ) from EV cells by One-way ANOVA. Data are mean  $\pm$  SD ( $n=3$ ).



**Fig S5. SASP analysis of W-CIN cells.** Plots depicting the amount of each of the SASP factors tested (pg/ml) in the CM of all cell lines. (\*) Indicates significant differences ( $p < 0.05$ ) from EV cells by One-way ANOVA. Data are mean  $\pm$  SD (n=6, n=5 for SEN).



**Fig S6. PQ and Bleo-induced senescence do not up-regulate CLEC11A.** Bright field representative images of (a) PQ- and (b) Bleo-induced HPF senescent fibroblasts stained for SA- $\beta$ gal.



**Fig S7. Efficiency of lentiviral transduction.** Plot representing the percentage of cells infected by lentivirus at the three multiplicity of infection tested (MOI)(n=2).

