

Supplementary Figure 1 a) Scheme of microfluidic device fabrication by photo and soft lithography, (a1, a2) 50nm Pd evaporated on Si wafer with 100 nm SiO₂ insulating layer and 5nm Cr as an adhesion layer (a3) A layer of SU-8 photoresist with the thickness of 4μm forms a microfluidic channel. (a4) The device is sealed with a PDMS layer to inlet and outlet port room to insert a Ag/ AgCl electrode (b) Photograph of the bioprotonic microfluidic device; described above, insert is top view.

Supplementary Figure 2 AFM characterization of SLB formation on the Pd contact, (a) Tapping mode AFM image of a naked Pd contact after O₂ plasma cleaning (b) Zoom in of a naked Pd contact shows an almost flat substrate with a roughness of ca. 0.5 ± 0.0 nm (c) AFM image of Pd contact covered with SLB, (d) Zoom in of a Pd covered with SLB shows increase in roughness of ca. 0.7 ± 0.1 nm (e) A representative force-distance curve for rupture of SLB in liquid following the protocol from literature.⁵ The rupture occurs at F= 1 nN, F is the force between the AFM tip and the sample surface with repulsive force oriented upwards on the y- axis. There is attraction right at a distance at 16.2 nm from the Pd surface by the small dip in the graph, indicating the tip first senses the bilayer. The rupture depth occurs at 4.8 ± 0.7 nm.

Supplementary Figure 3 I-V sweep for SLB formation, the electrical characteristics and integrity of the SLB are characterized with I-V measurements and ferrocyanide ([Fe(CN-)₆]⁴) is used as a redox probe in a two terminal configuration where Ag/AgCl is used as a reference and counter electrode. i_{H+} max (V= 600mV) for the Pd contact covered with the SLB (black trace) is ca.11 times smaller than for the naked Pd contact (gray trace). (The data is collected from 3 different devices with the different dimensions: Pd/ SLB: 3 different devices of 2 μm \times 50 μm, Naked Pd: 2 different devices of 2 μm \times 50 μm and 1 device of 2 μm \times 20 μm. The error bars are the root mean square (RMS) of the displacement of the data from the average value)

Supplementary Figure 4 I-V sweep of gA device, Pd / SLB has low i_{H+} > -0.225 mA cm⁻² across the entire voltage range, with a small oxidation peak with i_{H+} max of ca. 0.2 mA cm⁻², confirming the high polarization resistance of the SLB (black trace). Bioprotonic devices with integrated gA have higher i_{H_1} max of ca. 3 mA cm⁻², with a large PdH_x oxidation peak at 50 mV 3 , indicating H⁺ across the SLB membrane (green trace). (The data is collected from 3 different devices with the different dimensions: Pd/ SLB: 3 different devices of 2 μm x 50 μm, Pd/ SLB + gA: 2 μm x 20 μm, 2 μm x 50 μm, 2 μm x 70 μm. The error bars are the root mean square (RMS) of the displacement of the data from the average value).

Supplementary Figure 5 I-t plot of ALM disruption by introducing urea, Urea disrupts the ALM molecules, removing the ALM channels from the SLB ². At V= -200 mV, i_{H+} = -5.5 mA cm⁻² for SLB+ ALM (red trace) while the channel is open and at V= 100 mV, i_{H+} = 1.9 mA cm⁻² for SLB+ ALM (red trace). Addition of 40 mM urea to solution disrupts the aggregation of individual ALM monomers, it brings the corresponding i_{H+} to -0.4 mA cm⁻² at V= -200 mV (purple trace) and $i_{H+} = 0$ mA cm⁻² by the follow up at V= 100mV (blue trace). (The data is collected from 3 different devices with the different dimensions, Pd/ SLB: 3 different devices of 2 μm × 50 μm, Pd/ SLB + ALM: 2 μm × 20 μm, 2 μm × 40 μm, 2 μm × 50 μm, Pd/ SLB + ALM + Urea: 2 μm × 20 μm, 2 μm × 40 μm, 2 μm × 50 μm. The error bars are the root mean square (RMS) of the displacement of the data from the average value)

Supplementary Figure 6 I-V Sweep of ALM voltage-gated device, Pd / SLB has a low i_{H+} > -0.225 mA cm⁻² across the entire voltage range, with a small oxidation peak with i_{H+} max of ca. 0.2 mA cm⁻², confirming the high polarization resistance of the SLB (black trace). Bioprotonic devices with integrated with ALM have similar polarization resistances to SLB at low V due to the high impedance across the SLB membrane when ALM channels are closed. Additionally, we observe a large PdH_x oxidation peak with a max i_{H+} of ca. 45 mA cm⁻² at V= 50 mV³, indicating H⁺ flow across the SLB (red trace). (The data is collected from 3 different devices with the different dimensions, Pd/ SLB: 3 different devices of 2 μm × 50 μm, Pd/ SLB + ALM: 2 μm × 20 μm, 2 μm × 40 μm, 2 μm × 50 μm. The error bars are the root mean square (RMS) of the displacement of the data from the average value).

Supplementary Figure 7 Surface tension and AFM characterization of APTES self-assembly on

Pd contact, (a) Scheme of the Pd device covered with APTES ¹(b) Molecular structure of APTES (c) Surface tension measurements of a naked Pd contact after $O₂$ plasma cleaning of ca. 9° (d) Surface tension measurements of Pd contact covered by APTES shows increase on the contact angle to ca. 57° (e) AFM image in tapping mode of a naked Pd contact after plasma cleaning, shows an almost flat substrate with a roughness of ca. 0.6 ± 0.1 nm (f) AFM image in tapping mode of the Pd contact covered by ATPES shows a decrease in roughness to ca. 0.3 ± 0.1 nm. (n= 3, for the device with dimension of 2 μ m \times 50 μ m).

Supplementary Figure 8 (a, b, c) Schematics of bioprotonic gA devices with the self-assembly of APTES. (a) Pd contact coated with APTES / SLB. The APTES / SLB inhibits the flux of H⁺ from the bulk solution to the Pd/solution interface. (b) Pd contact with APTES / SLB incorporating gA is semipermeable to H⁺, with gA channels facilitating the rapid flow of H⁺ to the Pd solution interface (c) Addition of 1 mM Ca²⁺ to the bulk solution, blocks gA and prevents the flow of H⁺ to the Pd / APTES/ solution interface (d) i_{H+} -t plot for V = -200 mV and V = 0 mV for APTES / SLB (black trace) and APTES / SLB + gA (green trace) and APTES / SLB+ gA + Ca²⁺ (blue trace) on a Pd contact. (The data is collected from 3 different devices with the different dimensions, Pd/ APTES/SLB: $2 \mu m \times 50 \mu m$, and 2 different device of $2 \mu m \times$ 85 μm, Pd/ APTES / SLB + gA: 2 μm × 50 μm and 2 different devices of 2 μm × 80 μm, Pd/ APTES/ SLB+ gA+ Ca²⁺ 2 μm x 50 μm and 2 different devices of 2 μm x 80 μm. The error bars are the root mean square (RMS) of the displacement of the data from the average value).

Supplementary Figure 9 I-V sweep of gA device, Pd / APTES / SLB has i_{H+} > -4mA cm⁻² across the entire voltage range, without oxidation peak, confirming the high polarization resistance of the APTES / SLB (black trace). Bioprotonic devices with integrated gA have higher i_{H+} max of ca. 4 mA cm⁻², with a large PdH_x oxidation peak at 45 mV 3 , indicating H⁺ across the APTES / SLB membrane (green trace). (The data is collected from 3 different devices with the different dimensions: Pd/ SLB: 3 different devices of 2 μm × 85 μm, Pd/ SLB + gA: 2 μm × 40 μm, 2 μm × 50 μm, 2 μm × 70 μm. The error bars are the root mean square (RMS) of the displacement of the data from the average value).

Supplementary Figure 10 (a) I-V sweep of bioprotonic device with electrochemical redox reaction of H⁺ for the Pd contact with SLB (black trace), Pd contact with SLB + gA (green trace), and Pd contact with SLB + ALM (red trace). (b) Tafel fitting parameters are applied for the low polarization (linear) regime yielding the H⁺ exchange current at the respective electrodes. (The data is collected from the devices with dimension of 2 μm

Parameter	SLB On Pd contact
β a -bare Pd	0.452 ± 0.093
β_{a-SLB}	0.254 ± 0.025
$K^{0}_{ \textrm{LB}}$ (cm/s)	-4 $(6.82 \pm 2.45) \times 10$
$\overline{\mathbf{0}}$ 0 K_{SLB}/K_{bare} bare Pd	0.93
$(600 \, \text{mV})$ I / I SLB bare Pd	0.12 ± 0.03

Supplementary Table 1 Tafel parameter on SLB formation by I-V characterization, the Tafel electrode parameters extracted from the low polarization and exponential regimes. These values show similar ionic insulation as compared with reported electrodes with integrated SLB.⁴

Exponential

Linear

Ln(I) = ln(FAk^oC^o) +
$$
f \beta
$$
 (V - V^o)
\n β_a = anodic transfer coefficient
\n k^o = reaction rate constant
\nC^o = initial concentration
\n $f = F / RT$

 V^{0} = standard reaction potential of redox probe

$$
I = I^0 f (V - V^0)
$$

 I^0 = exchange current density

Supplementary Note 1

The AFM measurements on SLB formation were performed in liquid using the Bruker liquid cell holder using Bruker's Peak Force method. We used Scan-Asyst fluid cantilevers for all measurements. For topography images, the Bruker's Peak Force method was used with forces always under $F = 1$ nN, in order to prevent damage to the surface. F is the force between the AFM tip and the sample surface with repulsive force oriented upwards on the y- axis and it is calculated by the AFM software based on the cantilever deflection and cantilever spring constant. Distance is measured by Piezo element in the head. For the force-distance measurements, each data ($n= 20$, for the device with dimension of 2 μ m \times 50 μ m) was collected at a single point on the film, with forces high enough to penetrate the film.

Supplementary Note 2

Equation 1 to Equation 5:

$$
\beta = -2.303([H^+]IL + [OH^-]_{IL} + \sum_{j=0}^{n} \frac{c_{IL}K_{a,j}[H^+]_{IL}}{(K_{a,j}+[H^+]_{IL})^2})
$$
 6

Where the ([OH]_{IL} is the hydroxide concentration in the IL ([OH]_{IL} = Kw/[H⁺]_{IL}), C_{IL} is the concentration of the corresponding buffering acid (KH_2PO_4 or H_3PO_4), K_a is the acid dissociation constant of the buffering acid, and β is the buffer capacity.

We use a standard buffer solution of 5mM PBS and 100mM KCl, resulting in the following pH response:

$$
[K_2HPO_4] + [KH_2PO_4] + [H_3PO_4] = 5 \times 10^{-3} M
$$
 (2)

$$
pH = pK_a + \log \frac{[KA]}{[HA]}
$$
 (3)

Where [HA] and [KA] are the conjugate acid and base concentrations in the isolation layer, respectively. We use equation 2 and equation 3 to solve a system of equations that results in equations to calculate each acid concentration as a function of pH:

$$
[KH_2PO_4] = \frac{5 \times 10^{-3}}{10^{pH - 6.8} + 10^{2.148 - pH} + 1} M
$$
 (4)

$$
[H_3PO_4] = \frac{5*10^{pH-5.148}}{10^{pH-6.8}+10^{2.148-pH}+1} M
$$
 (5)

Supplementary References

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