

Supplementary Figure 1 a) Scheme of microfluidic device fabrication by photo and soft lithography, (a1, a2) 50nm Pd evaporated on Si wafer with 100 nm SiO₂ insulating layer and 5nm Cr as an adhesion layer (a3) A layer of SU-8 photoresist with the thickness of 4μ m forms a microfluidic channel. (a4) The device is sealed with a PDMS layer to inlet and outlet port room to insert a Ag/ AgCl electrode (b) Photograph of the bioprotonic microfluidic device; described above, insert is top view.



Supplementary Figure 2 AFM characterization of SLB formation on the Pd contact, (a) Tapping mode AFM image of a naked Pd contact after O_2 plasma cleaning (b) Zoom in of a naked Pd contact shows an almost flat substrate with a roughness of ca. 0.5 ± 0.0 nm (c) AFM image of Pd contact covered with SLB, (d) Zoom in of a Pd covered with SLB shows increase in roughness of ca. 0.7 ± 0.1 nm (e) A representative force-distance curve for rupture of SLB in liquid following the protocol from literature.⁵ The rupture occurs at F= 1 nN, F is the force between the AFM tip and the sample surface with repulsive force oriented upwards on the y- axis. There is attraction right at a distance at 16.2 nm from the Pd surface by the small dip in the graph, indicating the tip first senses the bilayer. The rupture depth occurs at 4.8 \pm 0.7 nm.



Supplementary Figure 3 I-V sweep for SLB formation, the electrical characteristics and integrity of the SLB are characterized with I-V measurements and ferrocyanide $([Fe(CN-)_6]^4)$ is used as a redox probe in a two terminal configuration where Ag/AgCl is used as a reference and counter electrode. i_{H+} max (V= 600mV) for the Pd contact covered with the SLB (black trace) is ca.11 times smaller than for the naked Pd contact (gray trace). (The data is collected from 3 different devices with the different dimensions: Pd/ SLB: 3 different devices of 2 µm × 50 µm, Naked Pd: 2 different devices of 2 µm × 50 µm and 1 device of 2 µm × 20 µm. The error bars are the root mean square (RMS) of the displacement of the data from the average value)



Supplementary Figure 4 I-V sweep of gA device, Pd / SLB has low $i_{H+} > -0.225$ mA cm⁻² across the entire voltage range, with a small oxidation peak with i_{H+} max of ca. 0.2 mA cm⁻², confirming the high polarization resistance of the SLB (black trace). Bioprotonic devices with integrated gA have higher i_{H+} max of ca. 3 mA cm⁻², with a large PdH_x oxidation peak at 50 mV ³, indicating H⁺ across the SLB membrane (green trace). (The data is collected from 3 different devices with the different dimensions: Pd/ SLB: 3 different devices of 2 μ m × 50 μ m, Pd/ SLB + gA: 2 μ m × 20 μ m, 2 μ m × 50 μ m, 2 μ m × 70 μ m. The error bars are the root mean square (RMS) of the displacement of the data from the average value).



Supplementary Figure 5 I-t plot of ALM disruption by introducing urea, Urea disrupts the ALM molecules, removing the ALM channels from the SLB². At V= -200 mV, $i_{H+} = -5.5$ mA cm⁻² for SLB+ ALM (red trace) while the channel is open and at V= 100 mV, $i_{H+} = 1.9$ mA cm⁻² for SLB+ ALM (red trace). Addition of 40 mM urea to solution disrupts the aggregation of individual ALM monomers, it brings the corresponding i_{H+} to -0.4 mA cm⁻² at V= -200 mV (purple trace) and $i_{H+} = 0$ mA cm⁻² by the follow up at V= 100mV (blue trace). (The data is collected from 3 different devices with the different dimensions, Pd/ SLB: 3 different devices of 2 μ m x 50 μ m, Pd/ SLB + ALM: 2 μ m x 20 μ m, 2 μ m x 40 μ m, 2 μ m x 50 μ m, Pd/ SLB + ALM + Urea: 2 μ m x 20 μ m. The error bars are the root mean square (RMS) of the displacement of the data from the average value)



Supplementary Figure 6 I-V Sweep of ALM voltage-gated device, Pd / SLB has a low $i_{H_+} > -0.225$ mA cm⁻² across the entire voltage range, with a small oxidation peak with i_{H_+} max of ca. 0.2 mA cm⁻², confirming the high polarization resistance of the SLB (black trace). Bioprotonic devices with integrated with ALM have similar polarization resistances to SLB at low V due to the high impedance across the SLB membrane when ALM channels are closed. Additionally, we observe a large PdH_x oxidation peak with a max i_{H_+} of ca. 45 mA cm⁻² at V= 50 mV⁻³, indicating H⁺ flow across the SLB (red trace). (The data is collected from 3 different devices with the different dimensions, Pd/ SLB: 3 different devices of 2 μ m × 50 μ m, Pd/ SLB + ALM: 2 μ m × 20 μ m, 2 μ m × 40 μ m, 2 μ m × 50 μ m. The error bars are the root mean square (RMS) of the displacement of the data from the average value).



Supplementary Figure 7 Surface tension and AFM characterization of APTES self-assembly on

Pd contact, (a) Scheme of the Pd device covered with APTES ¹(b) Molecular structure of APTES (c) Surface tension measurements of a naked Pd contact after O₂ plasma cleaning of ca. 9° (d) Surface tension measurements of Pd contact covered by APTES shows increase on the contact angle to ca. 57° (e) AFM image in tapping mode of a naked Pd contact after plasma cleaning, shows an almost flat substrate with a roughness of ca. 0.6 ± 0.1 nm (f) AFM image in tapping mode of the Pd contact covered by ATPES shows a decrease in roughness to ca. 0.3 ± 0.1 nm. (n= 3, for the device with dimension of 2 μ m × 50 μ m).



Supplementary Figure 8 (a, b, c) Schematics of bioprotonic gA devices with the self-assembly of APTES. (a) Pd contact coated with APTES / SLB. The APTES / SLB inhibits the flux of H⁺ from the bulk solution to the Pd/solution interface. (b) Pd contact with APTES / SLB incorporating gA is semipermeable to H⁺, with gA channels facilitating the rapid flow of H⁺ to the Pd solution interface (c) Addition of 1 mM Ca²⁺ to the bulk solution, blocks gA and prevents the flow of H⁺ to the Pd / APTES / solution interface (d) *i*_{*H*+}-t plot for V = -200 mV and V = 0 mV for APTES / SLB (black trace) and APTES / SLB + gA (green trace) and APTES / SLB + gA + Ca²⁺ (blue trace) on a Pd contact. (The data is collected from 3 different devices with the different dimensions, Pd/ APTES / SLB: 2 µm × 50 µm, and 2 different device of 2 µm × 85 µm, Pd/ APTES / SLB + gA: 2 µm × 50 µm and 2 different devices of 2 µm × 80 µm. The error bars are the root mean square (RMS) of the displacement of the data from the average value).



Supplementary Figure 9 I-V sweep of gA device, Pd / APTES / SLB has $i_{H+} > -4$ mA cm⁻² across the entire voltage range, without oxidation peak, confirming the high polarization resistance of the APTES / SLB (black trace). Bioprotonic devices with integrated gA have higher i_{H+} max of ca. 4 mA cm⁻², with a large PdH_x oxidation peak at 45 mV ³, indicating H⁺ across the APTES / SLB membrane (green trace). (The data is collected from 3 different devices with the different dimensions: Pd/ SLB: 3 different devices of 2 µm x 85 µm, Pd/ SLB + gA: 2 µm x 40 µm, 2 µm x 50 µm, 2 µm x 70 µm. The error bars are the root mean square (RMS) of the displacement of the data from the average value).



Supplementary Figure 10 (a) I-V sweep of bioprotonic device with electrochemical redox reaction of H⁺ for the Pd contact with SLB (black trace), Pd contact with SLB + gA (green trace), and Pd contact with SLB + ALM (red trace). (b) Tafel fitting parameters are applied for the low polarization (linear) regime yielding the H⁺ exchange current at the respective electrodes. (The data is collected from the devices with dimension of 2 μ m

Parameter	SLB On Pd contact
$\beta_{a \text{ -bare Pd}}$	0.452 ± 0.093
$eta_{a ext{-SLB}}$	0.254 ± 0.025
К [°] _{LB} (cm/s)	$(6.82 \pm 2.45) \times 10^{-4}$
$\mathbf{K}^{0}_{SLB}/\mathbf{K}^{0}_{bare Pd}$	0.93
I /I bare Pd (600 mV)	0.12 ± 0.03

Supplementary Table 1 Tafel parameter on SLB formation by I-V characterization, the Tafel electrode parameters extracted from the low polarization and exponential regimes. These values show similar ionic insulation as compared with reported electrodes with integrated SLB.⁴

Exponential

Linear

Ln(I) = ln(FAk⁰C⁰) +
$$f\beta$$
 (V - V⁰)
 β_a = anodic transfer coefficient
 k^0 = reaction rate constant
 C^0 = initial concentration
 f = F / RT

 V^{0} = standard reaction potential of redox probe

$$\mathbf{I} = \mathbf{I}^{\mathsf{O}} f (\mathbf{V} - \mathbf{V}^{\mathsf{O}})$$

 I^0 = exchange current density

Supplementary Note 1

The AFM measurements on SLB formation were performed in liquid using the Bruker liquid cell holder using Bruker's Peak Force method. We used Scan-Asyst fluid cantilevers for all measurements. For topography images, the Bruker's Peak Force method was used with forces always under F = 1 nN, in order to prevent damage to the surface. F is the force between the AFM tip and the sample surface with repulsive force oriented upwards on the y- axis and it is calculated by the AFM software based on the cantilever deflection and cantilever spring constant. Distance is measured by Piezo element in the head. For the force-distance measurements, each data (n= 20, for the device with dimension of 2 μ m × 50 μ m) was collected at a single point on the film, with forces high enough to penetrate the film.

Supplementary Note 2

Equation 1 to Equation 5:

$$\beta = -2.303([H^+]IL + [OH^-]_{IL} + \sum_{j=0}^{n} \frac{C_{IL}K_{a,j}[H^+]_{IL}}{(K_{a,j} + [H^+]_{IL})^2}) \qquad ^{6}$$

Where the ($[OH_{lL}]_{lL}$ is the hydroxide concentration in the IL ($[OH_{lL}]_{lL} = Kw/[H_{lL}]$), C_{lL} is the concentration of the corresponding buffering acid (KH_2PO_4 or H_3PO_4), K_a is the acid dissociation constant of the buffering acid, and β is the buffer capacity.

We use a standard buffer solution of 5mM PBS and 100mM KCl, resulting in the following pH response:

$$[K_{2}HPO_{4}] + [KH_{2}PO_{4}] + [H_{3}PO_{4}] = 5 \times 10^{-3} M$$
(2)
$$pH = pK_{a} + \log \frac{[KA]}{[HA]}$$
(3)

Where [HA] and [KA] are the conjugate acid and base concentrations in the isolation layer, respectively. We use equation 2 and equation 3 to solve a system of equations that results in equations to calculate each acid concentration as a function of pH:

$$[KH_2PO_4] = \frac{5*10^{-3}}{10^{pH-6.8} + 10^{2.148-pH} + 1} M$$
(4)

$$[H_3PO_4] = \frac{5*10^{pH-5.148}}{10^{pH-6.8}+10^{2.148-pH}+1} M$$
(5)

Supplementary References

- 1 Leonenko, Z., Carnini, A. & Cramb, D. Supported planar bilayer formation by vesicle fusion: the interaction of phospholipid vesicles with surfaces and the effect of gramicidin on bilayer properties using atomic force microscopy. Biochimica et Biophysica Acta (BBA)-Biomembranes 1509, 131-147 (2000).
- 2 Mathew, M. K., Nagaraj, R. & Balaram, P. Membrane channel-forming polypeptides. Aqueous phase aggregation and membrane-modifying activity of synthetic fluorescent alamethicin fragments. Journal of Biological Chemistry 257, 2170-2176 (1982).
- 3 Chevillot, J.-P., Farcy, J., Hinnen, C. & Rousseau, A. Electrochemical study of hydrogen interaction with palladium and platinum. Journal of Electroanalytical Chemistry and Interfacial Electrochemistry 64, 39-62 (1975).
- 4 Misra, N. et al. Bioelectronic silicon nanowire devices using functional membrane proteins. Proceedings of the National Academy of Sciences 106, 13780-13784 (2009).
- 5 Attwood, S. J., Choi, Y. & Leonenko, Z. Preparation of DOPC and DPPC supported planar lipid bilayers for Atomic Force Microscopy and Atomic Force Spectroscopy. International journal of molecular sciences 14, 3514-3539 (2013).
- 6 The Melanin Binding of Drugs and Its Implications. Drug Metabolism Reviews 15, 1183-1212, doi:10.3109/03602538409033561 (1984).