

Supporting Information

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Acid-Responsive Absorption and Emission of 5-N-Arylaminothiazoles: Emission of White Light from a Single Fluorescent Dye and a Lewis Acid

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General Remarks.

¹H and ¹³C NMR spectra were recorded in CDCl₃ or CD₃OH. Chemical shifts of protons are reported in δ values referenced to tetramethylsilane as an internal standard in CDCl₃, and the following abbreviations are used: s, singlet; d, doublet; t, triplet; m, multiplet. All spectra were acquired in the proton-decoupled mode. HRMS were recorded on a double-focusing mass spectrometer (EI). IR spectra were obtained using KBr pellets or neat films. UV–vis absorption, fluorescence, and absolute fluorescence quantum yields were obtained on the respective spectrometers. Toluene was distilled from sodium metal. All other chemicals were used without further purification. Column chromatography was performed on silica gel 60 N (Spherical Neutral, 100–210 µm). All manipulations of synthesis were carried out under an argon atmosphere. All measurements were carried out under atmosphere at 23 °C.

Materials.

2,6-Diphenylisonicotinic acid, Lawesson's reagent and deuterated solvents were purchased from Aldrich Chemical Company, Inc. Dichloromethane (for synthesis), ethyl acetate, hexane, iodine, methanol, THF and silica gel 60N (Spherical Neutral, 100–210 μ m) were purchased from Kanto Chemical Co., Inc. Acetonitrile, benzylamine, dichloromethane (for measurements), diethyl ether, Na₂S₂O₃, MgSO₄, toluene and 3 Å molecular sieve were purchased from Nakarai Tesque Inc. *n*-BuLi was purchased from Mitsuwa Chemical, Ltd. The compounds **1**, **2**, thioamides and thioformamides were prepared according to literature procedures.^[1]

Instruments.

 1 H and 13 C NMR spectra were measured on a JNM- α 400 spectrometer. The mass spectra (MS) and high resolution mass spectra (HRMS) were taken on JMS-700 mass spectrometers. The IR spectra were obtained on a JASCO-FT-IR 410 spectrophotometer. UV-vis spectra were measured on a HITACHI U-4100 spectrophotometer. Fluorescence spectra were measured on a HORIBA FluoroMax-4. Excited and fluorescence spectra were measured on a JASCO spectrofluorometer FP-8600. Fluorescent quantum yields were measured on a HAMAMATSU absolute PL quantum yield spectrometer C11347-01. Fluorescence lifetimes were measured on a HAMAMATSU compact fluorescence lifetime spectrometer C11367.

Synthesis of thioamide

N-Phenylmethyl-3,5-diphenyl-4-pyridincarboamide



The 2,6-diphenylisonicotinic acid (0.826 g, 3.0 mmol) and 3 Å MS (0.27 g) was added benzylamine (0.33 mL, 3.0 mmol) at room temperature, and the mixture was stirred for 46.5 h at 160 °C. The resulting mixture was filtered, and extracted with methanol. The residue was purified by column chromatography (SiO₂, CH₂Cl₂) and

recrystallized from CH₂Cl₂/hexane to give amide (0.922 g, 84%) as a white solid. (m.p. 178–180 °C); IR (KBr) 3285, 3031, 1638, 1537, 1495, 1455, 1404, 1371, 1344, 1274, 1244, 1179, 1072, 918, 784, 683

cm⁻¹; ¹H NMR (400 MHz, CD₃OH) δ 4.64 (d, *J* = 8.0 Hz, 2H, CH₂), 6.57 (br, 1H, NH), 7.28–7.45 (m, 12H, Ar), 7.82 (s, 2H, Py), 8.09–8.11 (m, 3H, Ar); ¹³C NMR (100 MHz, CD₃COCD₃) δ 44.2 (CH₂), 115.9, 127.1, 127.8, 128.0, 128.7, 128.8, 128.9, 129.5, 129.9, 137.0, 137.6, 138.6, 143.3, 157.8, 166.2 (Ar), ; MS (EI) *m*/*z* 364 (M⁺); HRMS (EI) Calcd for C₂₅H₂₀N₂O : 364.1576; found : 364.1588.

N-Phenylmethyl-3,5-diphenyl-4-pyridincarbothioamide



To a solution of *N*-phenylmethyl-3,5-diphenyl-4-pyridincarboamide (0.364 g, 1.0 mmol) in toluene (5.0 mL) was added Lawesson's reagent (0.809 g, 2.0 mmol) at room temperature, and the mixture was stirred for 6 h at 115 °C. The resulting mixture was concentrated in vacuo. The residue was purified by column

chromatography (SiO₂, CH₂Cl₂, methanol and hexane : EtOAc = 5 : 1) to give thioamide (0.269 g, 71%) as a yellow solid. (m.p. 166–167 °C); IR (KBr) 3177, 3030, 2916, 1603, 1549, 1495, 1391, 1329, 1259, 1197, 1077, 992, 962, 660 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 5.02 (d, *J* = 4.0 Hz, 2H, CH₂), 7.38–7.50 (m, 11H, Ar), 7.89 (m, 3H, NH, Py), 8.12 (d, *J* = 8.0 Hz, 4H, Ar); ¹³C NMR (100 MHz, CDCl₃) δ 51.1 (CH₂), 115.7, 127.2, 128.6, 128.6, 128.9, 129.3, 129.6, 135.8, 138.7, 150.3, 157.7 (Ar), 197.5 (S=C); MS (EI) *m/z* 380 (M⁺); HRMS (EI) Calcd for C₂₅H₂₀N₂S : 380.1347; found : 380.1350.

General Procedure for the Preparation of Thiazolines.



To a solution of thioamide (1 equiv) in THF was added slowly a 1.25 M solution of *n*-butyllithium in *n*-hexane (2 equiv) at -80–0 °C, and the mixture was stirred for 10 min at this temperature. To this was added thioformamide (1 equiv) at -80–0 °C, and the mixture was stirred for 20–30 min at this temperature. To this was added iodine (3 equiv) at -80–0 °C, and the mixture was stirred for 3 h at -80 °C to room temperature. The resulting mixture was poured into a saturated aqueous solution of Na₂S₂O₃, and extracted with Et₂O. The organic layer was dried over MgSO₄ and concentrated in vacuo. The residue was purified by column chromatography (SiO₂) to give the corresponding thiazolines.

General Procedure for the Preparation of Thiazoles.



To a solution of thiazoline (1 equiv) in THF was added iodine (2 equiv) at room temperature, and the mixture was stirred for 14–42 h at room temperature. The resulting mixture was poured into a saturated

aqueous solution of $Na_2S_2O_3$, and extracted with Et_2O . The organic layer was dried over MgSO₄ and concentrated in vacuo. The residue was purified by column chromatography (SiO₂) to give the corresponding thiazoles.

(4R*,5S*)-4,5-Dihydro-2-(4-pyridyl)-4-phenyl-N,N-bis(4-methylphenyl)-5-thiazolamine



According to the general procedure for thiazolines, the crude material was purified by column chromatography (SiO₂, hexane : EtOAc = 5:1) to give the corresponding thiazoline (0.156 g, 72%) as a yellow oil. : IR (KBr) 3028, 2920, 2859, 1592, 1508, 1407, 1237, 1034, 962, 824, 730, 700 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 2.27 (s, 6H, Me), 6.00 (d, J = 4.0 Hz, 1H, SCH), 6.34 (d,

J = 4 Hz, 1H, NCH), 6.91 (d, J = 8 Hz, 4H, Ar), 7.06 (d, J = 8 Hz, 4H, Ar), 7.28–7.35 (m, 5H, Ar), 7.63–7.64 (m, 2H, Ar), 7.67–7.68 (m, 2H, Ar); ¹³C NMR (100 MHz, CDCl₃) δ 23.6, 85.4, 85.5, 124.9, 126.2, 129.2, 131.0, 131.8, 132.9, 136.4, 142.4, 143.5, 146.1, 153.2 169.9; MS (EI) m/z 435 (M⁺); HRMS (EI) Calcd for C₂₈H₂₅N₃OS : 435.1769; found : 435.1761.

2-(4-Pyridyl)-4-phenyl-*N*,*N*-bis(4-methylphenyl)-5-thiazolamine (3)



According to the general procedure for thiazoles, compound **3** was prepared. The crude material was purified by column chromatography (SiO₂, hexane : EtOAc = 3 : 1) to give thiazole **3** (0.09 g, 63%) as an orange oil.: IR (KBr) 3027, 2919, 1595, 1505, 1444, 1345, 1286, 995, 909, 812, 727, 694 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 2.26 (s, 6H, OMe), 6.97–7.03 (m, 7H, Ar),

7.21–7.29 (m, 4H, Ar), 7.79–7.80 (m, 2H, Ar), 7.92–7.94 (m, 2H, Ar), 8.66–8.68 (m, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 23.6 (Me), 122.8, 123.3, 124.5, 130.3, 131.0, 131.1, 132.8, 135.9, 139.6, 143.7, 147.1, 151.5, 153.3 (Ar), 162.3 (SC=N); MS (EI) *m*/*z* 433 (M⁺); HRMS (EI) Calcd for C₂₈H₂₃N₃S : 433.1613; found : 433.1611.

(4R*,5S*)-4,5-Dihydro-2-(4-pyridyl)-4-phenyl-N,N-bis(4-methoxyphenyl)-5-thiazolamine



According to the general procedure for thiazolines, the crude material was purified by column chromatography (SiO₂, hexane : EtOAc = 10 : 1 and 5 : 1) to give the corresponding thiazoline (0.189 g, 81%) as a yellow oil. : IR (KBr) 3037, 2998, 2930, 2832, 1591, 1506, 1242, 1032, 961, 823, 700 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 3.74 (s, 6H, OMe), 6.00 (d, *J* = 4.0 Hz, 1H,

SCH), 6.31 (d, J = 4 Hz, 1H, NCH), 6.79–6.82 (m, 4H, Ar), 6.95–6.98 (m, 4H, Ar), 7.31–7.35 (m, 5H, Ar), 7.64–7.66 (m, 2H, Ar), 7.68 (d, 4.0 Hz, 2H, Ar); ¹³C NMR (100 MHz, CDCl₃) δ 58.3, 58.4, 85.6, 86.3, 117.6, 125.1, 127.6, 129.1, 129.3, 131.1, 131.6, 142.1, 152.9, 153.0, 159.1, 169.7; MS (EI) m/z 465 (M⁺, -2H); HRMS (EI) Calcd for C₂₈H₂₅N₃O₂S : 467.1667; found : 467.1646.

2-(4-Pyridyl)-4-phenyl-*N*,*N*-bis(4-methoxyphenyl)-5-thiazolamine (4)



According to the general procedure for thiazoles, compound **4** was prepared. The crude material was purified by column chromatography (SiO₂, hexane : EtOAc = 3 : 1) to give thiazole **4** (0.05 g, 44%) as a brown oil.: IR (KBr) 2929, 1593, 1504, 1443, 1241, 1779, 1033, 823, 696, 669 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 3.76 (s, 6H, OMe), 6.77–6.79 (m, 4H, Ar),

7.01–7.03 (m, 4H, Ar), 7.21–7.29 (m, 3H, Ar), 7.79–7.81 (m, 2H, Ar), 7.93–7.95 (m, 2H, Ar), 8.68–8.69 (m, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 55.7, 55.9 (OMe), 114.8, 114.9, 120.1, 123.1, 123.4, 127.7, 128.4, 133.4, 140.8, 141.1, 143.9, 148.0, 150.7, 150.9, 156.1 (Ar), 159.0 (SC=N); MS (EI) *m*/*z* 465 (M⁺); HRMS (EI) Calcd for C₂₈H₂₃N₃O₂S : 465.1511; found : 465.1530.

(4R*,5S*)-4,5-Dihydro-2-(3,5-diphenyl-4-pyridyl)-4-phenyl-*N*,*N*-diphenyl-5-thiazolamine



According to the general procedure for thiazolines, the crude material was purified by column chromatography (SiO₂, hexane : EtOAc = 20 : 1) to give the corresponding thiazoline (0.136 g, 49%) as a pale yellow solid. (m.p. 87–90 °C) : IR (KBr) 3429, 3033, 1592, 1578, 1551, 1492, 1449, 1405, 1352, 1248, 1206, 1179,

1032, 906, 692 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 6.11 (d, *J* = 4.0 Hz, 1H, SCH), 6.41 (d, *J* = 4.0 Hz, 1H, NCH), 7.07–7.12 (m, 6H, Ar), 7.28–7.38 (m, 8H, Ar), 7.42–7.52 (m, 7H, Ar), 8.05 (s, 2H, Py), 8.19 (d, *J* = 4.0 Hz, 4H, Ar); ¹³C NMR (100 MHz, CDCl₃) δ 82.0, 82.7, 117.1, 123.5, 124.1, 126.4, 127.1, 128.2, 128.7, 129.0, 129.3, 129.5, 138.8, 139.3, 142.2, 145.5, 157.6 (Ar), 167.5 (SC=N); MS (EI) *m*/*z* 559 (M⁺); HRMS (EI) Calcd for C₃₈H₂₉N₃S : 559.2082; found : 559.2077.

2-(3,5-Diphenyl-4-Pyridyl)-4-phenyl-*N*,*N*-diphenyl-5-thiazolamine (6)



According to the general procedure for thiazoles, compound **6** was prepared. The crude material was purified by column chromatography (SiO₂, hexane : EtOAc = 20 : 1) and GPC to give thiazole **6** (0.05 g, 65%) as a yellow solid. (m.p. 240–242 °C): IR (KBr) 3435, 1595, 152, 1524, 1488, 1443, 1352, 1306, 1271,

1154, 1072, 908, 777, 693 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.02 (t, *J* = 4.0 Hz, 2H, Ar), 7.14–7.16 (m, 4H, Ar), 7.23–7.29 (m, 7H, Ar), 7.45–7.47 (m, 2H, Ar), 7.49–7.53 (m, 4H, Ar), 7.94–7.96 (m, 2H, Ar), 8.20–8.23 (m, 6H, Ar); ¹³C NMR (100 MHz, CDCl₃) δ 115.2, 121.8, 123.6, 127.2, 127.6, 128.4, 128.8, 129.5, 133.1, 139.1, 142.0, 142.5, 146.5, 148.9, 149.2, 157.9, 159.4 (Ar), 160.8 (SC=N); MS (EI) *m/z* 557 (M⁺); HRMS (EI) Calcd for C₃₈H₂₇N₃S : 557.1926; found : 557.1923.

2-(4-Pyridyl)-4-phenyl-*N*,*N*-bis(4-hydroxyphenyl)-5-thiazolamine (5)



To a 5-*N*,*N*-bis(4-methoxyphenyl)thiazole **4** (0.028 g, 0.06 mmol) was added HBr (0.1 mL) at room temperature, and the mixture was stirred for 13 h at 100 °C. The resulting mixture was poured into a saturated aqueous solution of K₂CO₃, and extracted with EtOAc. The organic layer was dried over Na₂SO₄

and concentrated in vacuo. The residue was purified by column chromatography (SiO₂, 5–7% MeOH in CH₂Cl₂) to give thiazole **5** (0.023 g, 90%) as a yellow solid. (m.p. 272–274 °C); IR (KBr) 3391, 3233, 3043, 3025, 2796, 1602, 1507, 1441, 1242, 1213, 830, 695 cm⁻¹; ¹H NMR (400 MHz, CD₃OH) δ 6.62–6.65 (m, 4H, Ar), 6.85–6.87 (m, 4H, Ar), 7.18–7.23 (m, 3H, Ar), 7.82–7.84 (m, 2H, Ar), 7.88–7.90 (m, 2H, Ar), 8.57–8.59 (m, 2H, Ar) The protons due to O<u>H</u> were not observed.; ¹³C NMR (100 MHz, CD₃COCD₃) δ 116.6, 117.4, 120.9, 124.7, 128.8, 129.3, 129.5, 134.9, 141.3, 148.0, 152.1, 155.2, 159.5, 165.2; MS (EI) *m/z* 437 (M⁺); HRMS (EI) Calcd for C₂₆H₁₉N₃O₂S : 437.1198; found : 437.1212.

Photophysical properties and spectra (The addition of Brønsted acids)



Spectra of thiazole **3–6** and **8–11** were measured in Et_2O (Table 2). Fluorescent spectra of **8–10** were not observed because of their weak fluorescent intensities.







The addition of HCl to thiazole **2**, shifted its wavelengths of absorption and emission to longer wavelengths by 83 nm and 108 nm, respectively (Table S1, entry 1). However, **2** did not show any changes when adding TFA and CH₃COOH (entries 2 and 3).

Table S1. Photophysical properties of thiazole 2 and 12 ^a							
N		Ph N Ph Ph	Acids			N 12	Ph I N Ph Ph
entry	l _{abs} (nm)	l _{em} (nm) ^b	acid	IP ^c (nm)		l _{abs} (nm)	l _{em} (nm) ^t
1	382	472	HCI	412	12	465	580
2	382	472	TFA	-	-	-	
3	382	472	CH₃COOH	-	-	-	

 ^{a}In Et_2O, c = 10 5 M. $^{b}Excited$ at wavelength of maximum absorption. ^{c}IP stands for isosbestic point.





Photophysical properties and spectra (The addition of Lewis acids)









Photophysical properties of the mixture of thiazole **2** and Lewis acids were measured in CH_2Cl_2 . The reaction of **2** and $GaCl_3$ passed through two steps (0.25–0.5 equiv. and 0.75–3 equiv. of $GaCl_3$). The red arrow indicates the equivalents of $GaCl_3$ from 0.25 to 0.5 equiv. While, the blue arrow indicates the equivalents of $GaCl_3$ from 0.75 to 3 equiv.



Fig. S26 Fluorescent spectra of 2·B(C6F5)3 - 0.25 0.5 0.75 - 1 - 1.25 Intensity - 1.5 - 1.75 Wavelength / nm



The spectra of absorption and emission of 2 and 6 changed by adding $B(C_6F_5)_3$.

Wavelength / nm

CIE coordinates



CIE coordinates of solution **2** and $B(C_6F_5)_3$ were plotted from the emission spectra (Figure 3 (b), Table S2). The color of the solution changed more drastically than that of **6**. The solution **2** showed bluish white emission when 0.5 equivalents of $B(C_6F_5)_3$ were added.



Table S3. CIE data of 6 with 0-100 equivalents of $B(C_6F_5)_3{}^a$



CIE calculated from emission spectra of solution **6** and $B(C_6F_5)_3$ (Figure S31, Table S3). The emission color of solution **6** and $B(C_6F_5)_3$ gradually changed from blue to white and orange.

DFT calculations

All calculations were carried out using the GAUSSIAN 09 program.^[2] Geometry optimization was performed with hybrid density functional theory (DFT) at the B3LYP^[3] level, by using the 6-31+G(d,p) basis set for all atoms. The UV/Vis absorption spectra of optimized geometries were calculated with the time dependent (TD) DFT method at CAM-B3LYP^[4] level. Solvent effects were taken into account by means of polarized continuum model (PCM)^[5] calculations. The results for **2** and **7** are shown in Figures S31 and S32 respectively, and the energy levels of their HOMO and LUMO, and NBO charges are summarized in Table S4. XYZ Coordinates are uploaded as a separate Supporting Information.



Table S4. Summary of energy and energy gaps ofHOMO and LUMO of 2 and $7 \cdot H_2O$

	HOMO (eV)	LUMO (eV)	HOMO-LUMO gap (eV)
2	-6.85	-1.14	5.71
7•H₂O	-7.05 (0.2 down)	-2.13 (0.99 down)	4.92

The calculated UV-vis absorption spectra for **2** and **7** at CAM-B3LYP level show a red shift of 70 nm in absorption maximum from 364 nm in **2** to 434 nm in **7** after protonation, which is consistent with the experimental results in Table 1 exhibiting a substantial red shift of 93 nm from 382 nm in **2** to 475 nm in protonated **7**. To find possible origins of such remarkably red-shifted absorption of **7**, we investigated their electronic properties in details. TD-DFT results indicated that the maximum absorption is mainly assigned to HOMO-LUMO transition for both of **2** and **7**, as shown in Fig. S32 and S33. Interestingly, LUMO orbital has a great contribution from the pyridyl group, whereas HOMO orbital is basically from aminothiazole fragment and almost none from the pyridyl group. It is noticed that protonation of **2** leads to lowering of both HOMO and LUMO orbitals. But LUMO orbital is much more strongly lowered by 0.99 eV in energy, compared with HOMO orbital with lowering of 0.2 eV, as shown in Table S4. Consequently, the HOMO-LUMO electronic band gap significantly drops down by 0.79 eV from 5.71 eV to 4.92 eV, which is probably the origin for the intense red-shifted absorption in **7**.

X-ray structure analyses

The X-ray quality crystals were obtained by slow diffusion of hexane into CH_2Cl_2 solutions of 2 and 7. The crystal was cut from the grown crystals and was mounted on a glass fiber. Intensity data were collected on a RIGAKU Saturn70 CCD system with VariMax Mo Optic using MoK α radiation ($\lambda = 0.71075$ Å). X-ray absorption was corrected by multi-scan methods.^{[6], [7]} The structures were solved by direct method using SHELXS-97 and refined by the full-matrix least-squares method on F^2 using SHLEXL-97.^[8] Structure determination was carried out using the free GUI software of Yadokari-XG 2009.^[9] Crystal data and structure refinement are summarized in the Tables S5 and S6.

Table S5. Crystal data and structur	re refinement for 2		
Empirical formula	C ₂₆ H ₁₉ N ₃ S		
Formula weight	405.50		
Temperature	103(2) K		
Wavelength	0.71075 Å		
Crystal system	Orthorhombic		
Space group	Pbca		
Unit cell dimensions	a = 7.4437(4) Å	a=90°	
	b = 20.0110(11) Å	b=90°	
	c = 26.7323(16) Å	g= 90°	
Volume	3981.9(4) Å ³		
Z	8		
Density (calculated)	1.353 Mg/m ³		
Absorption coefficient	0.181 mm ⁻¹		
<i>F</i> (000)	1696		
Crystal size	0.08 x 0.02 x 0.01 mm ³		
Theta range for data collection	1.52 to 25.50°		
Index ranges	-8<=h<=9, -24<=k<=23, -32<=l<=32		
Reflection collected	50863		
Independent reflections	3703 [<i>R</i> (int) = 0.2451]		
Completeness to theta = 27.50°	100.0%		
Max. min. transmission	0.9982 and 0.9857		
Refinement method	Full-matrix least-squares on F ²		
Data / restraints / parameters	3703 / 0 / 271		
Goodness-of-fit on F^2	1.046		
Final R indices [<i>I</i> >2sigma(<i>I</i>)]	$R_1 = 0.0739, wR_2 = 0.1698$		
R indices (all data)	$R_1 = 0.1386$, w $R_2 = 0.2148$		
Largest diff. peak and hole	0.524 and -0.640 e.Å ⁻³		

Crystal data and structure refinement

,			
Empirical formula	$C_{27}H_{22}F_3N_3O_4S_2$		
Formula weight	573.60		
Temperature	103(2) K		
Wavelength	0.71075 Å		
Crystal system	Monoclinic		
Space group	<i>P</i> 2 ₁ /n		
Unit cell dimensions	a = 11.8558(6) Å	a=90°	
	b = 6.8270(3) Å	b=90.959°	
	c = 31.3378(18) Å	g= 90°	
Volume	2536.1(2) Å ³		
Z	4		
Density (calculated)	1.502 Mg/m ³		
Absorption coefficient	0.273 mm ⁻¹		
F (000)	1184		
Crystal size	0.08 x 0.04 x 0.01 mm ³		
Theta range for data collection	1.83 to 25.50°		
Index ranges	-14<=h<=14, -8<=k<=8, -37<=l<=37		
Reflection collected	22829		
Independent reflections	4583 [<i>R</i> (int) = 0.1152]		
Completeness to theta = 27.50°	97.0%		
Max. min. transmission	0.9973 and 0.9785		
Refinement method	Full-matrix least-squares on F ²		
Data / restraints / parameters	4583 / 4 / 358		
Goodness-of-fit on F^2	1.018		
<pre>Final R indices [l>2sigma(l)]</pre>	$R_1 = 0.0720, w R_2 = 0.157$	7	
R indices (all data)	$R_1 = 0.1624, wR_2 = 0.2070$		
Largest diff. peak and hole	0.977 and -0.368 e.Å ⁻³		

Table S6. Crystal data and structure refinement for 7

Planes for the calculations of dihedral angles A-2, A-4, and A-5 of 2 and 7 are shown in Figures S33–S38.

Figure. S33 Two planes for the calcuation of dihedorak angle A-2 of 2

the second



Figure. S34 Two planes for the calcuation of dihedorak angle A-4 of 2



Figure. S37 Two planes for the calcuation of dihedorak angle A-4 of 7



Figure. S38 Two planes for the calcuation of dihedorak angle A-5 of 7

Figure. S35 Two planes for the calcuation of dihedorak angle A-5 of 2





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¹H and ¹³C NMR spectra

















