Appendix 2: Supplementary tables [posted as supplied by author]

Akkoc et al 2010[1] Andersen et al 2013[2] Black et al 2001[3] Black et al 2002[4]	Open, randomized, single centre trial Randomised, control trial, Samples at 6-11 weeks (early sample group, n=124) or5-9 months (late sample group, n=221). Randomised, controlled trial, single region	Healthytermnewborns (n=19)Infants in Guinea- Bissau, n=357 (345 included in analysis)Healthy individuals fromHealthy individuals fromNorthern Malawi with no prior history of BCG vaccinationHealthyyoung	BCG vaccination at birth (n=10) compared to BCG vaccination at 2 months (n=9) BCG revaccination at 19 months, n=158 or nothing, n=187 BCG compared to placebo (dextran matrix of BCG vaccine), single dose.	Cytokine responses IFN-γ, IL-13, TNF- α and IL-10 stimulated with LPS, PPD, or PHA. Skin tests and IFN-γ responses to Mycobacterial spp.	IFN-γ and IL-10 responses to PBMCs stimulated with PHA IFN-γ, IL-13, TNF- α and IL-10 stimulated with LPS or PHA. IFN-γ responses to control samples in lymphocyte cultures in	pg/ml Geometric mean/geometric mean ratio pg/ml	Non- significant(NS) between groups. NS for IFN-γ and IL-13 NS	Part of the REVAC trial which was halted prematurely due to excess mortality. Study carried out in the context of the Karonga Prevention Study, a large vaccine trial and
2013[2] Black <i>et al</i> 2001[3] Black <i>et al</i>	trial, Samples at 6-11 weeks (early sample group, n=124) or5-9 months (late sample group, n=221). Randomised, controlled trial, single region Randomised,	Bissau, n=357 (345 included in analysis) Healthy individuals from Northern Malawi with no prior history of BCG vaccination Healthy young	months, n=158 or nothing, n=187 BCG compared to placebo (dextran matrix of BCG vaccine), single dose.	and IL-10 stimulated with LPS, PPD, or PHA. Skin tests and IFN-γ responses to	and IL-10 stimulated with LPS or PHA.	mean/geometric mean ratio	IL-13	which was halted prematurely due to excess mortality. Study carried out in the context of the Karonga Prevention Study, a large vaccine trial and
2001[3] Black <i>et al</i>	controlled trial, single region Randomised,	from Northern Malawi with no prior history of BCG vaccination Healthy young	placebo (dextran matrix of BCG vaccine), single dose.	responses to	control samples in	pg/ml	NS	context of the Karonga Prevention Study, a large vaccine trial and
				1				epidemiological study of tuberculosis and leprosy in the Karonga district.
	multiple regions	adults from Northern Malawi (n= 633, mean age 19 years) and East London and Essex, United Kingdom (n=424, mean age 14 years)	BCG vaccination compared to placebo	IFN-γ responses to control antigens and PPD, and skin testing to mycobacterial antigens	IFN-γ responses to control antigens	pg/ml	NS	Study carried out in the context of the Karonga Prevention Study, a large vaccine trial and epidemiological study of tuberculosis and leprosy in the Karonga district.
Burl <i>et al</i> Jul. 2010[5]	Open, randomised, single centre trial	Healthy Gambian newborns (n=103)	BCG vaccination at birth (n=53) compared to BCG vaccination at 4.5 months (n=50)	Cytokine responses and cell phenotyping	Whole blood phenotyping FN-γ, IL-13, IL-6, IL-17 ind IL-10 production by whole blood stimulated with control antigens	% CD4+CD25+ T cells and % CD4+CD25+FOXP 3+ T cells Log10pg/ml	NS	
Burl <i>et al</i> Aug. 2010[6]	Open, randomised, single centre trial	Healthy Gambian newborns (n=103)	BCG vaccination at birth compared to BCG vaccination at 4.5 months	Immune responses to tuberculin skin tests, cytokine studies and cellular phenotyping	Controls (SEB) used for whole blood IFN-γ and IL-10 responses (not reported) T cell phenotype	pg/ml %CD4+CD25+ T cells and %CD4+CD25+FOX 3+ T cells	Non-specific data not reported therefore significance unknown	

[7]	single centre trial	newborns (n=103)	compared to BCG vaccination at 4.5 months	cellular phenotyping	and IFN-γ) from whole blood cultures stimulated with SEB. Cellular phenotying	%CD4+CD25+ T cells and %CD4+CD25+FOX 3+ T cells	production at 4.5 months in SEB stimulated cultures of vaccinated compared to unvaccinated (p<0.0421).	which two other included publications were based
Djuardi <i>et al</i> 2010[8]	Birth cohort from two regions in Indonesia	Newborns (n=147, mean age at vaccination 5 weeks, IQR 2-8.5)	Infant vaccination program comprising, BCG, Hepatitis B, DTP, OPV and measles vaccines.	Cytokine responses	IFN-γ, IL-5, TNF-α, IL- 10 and IL-13 responses from whole blood cultures stimulated with PHA, LPS and media only	pg/ml and median with IQR	Increased IFN- γ production to PHA at 24 months vs baseline (p<0.001). Decreased IL-5 to PHA at 5 months vs baseline (p<0.05). Decreased TNF- α and IL-10 to LPS at 24 months vs baseline (p<0.001)	
Elliot <i>et al</i> 2011[9] (BCG and TT)	Observational analysis of newborns after a randomised, double blinded, controlled trial of anti-helminth therapy in pregnant women	Newborns of women recruited from a Ugandan hospital antenatal clinic (n=2345).	Infant vaccination program comprising of BCG, polio, diphtheria, pertussis, tetanus, hepatitis B, <i>H. influenzae</i> and measles vaccines.	Cytokine responses to BCG and TT	IFN-γ, IL-5, IL-13 and IL-10 responses to control stimulated cultures (not reported)	Geometric mean ratios	Not specific results/significanc e not reported	
Faustman <i>et al</i> 2012[10]	Double blinded, randomised, controlled, single centre, trial	Long term type 1 diabetic adults (n=6) and healthy non- diabetic controls	BCG vaccination (n=3 diabetics and n=3 controls) compared to placebo (n= 3 diabetics and n=3 controls)	T-cells, auto-antibodies and C-peptide	Autoreactive T cells Insulin autoantibodies (GAD, IA-2A, ZnT8A) C-peptide	% Units pmol/L	Autoreactive T cells -Significance tests not reported Insulin autoantibodies- GAD:2 BCG treated subjects significant changes from baseline (one increase, one decrease p=0.0001/0.0017). ZnT8A: sig decrease in one BCG subject only Transient and significant rise in C-peptide level in BCG subjects	
Fjallbrant <i>et al</i> 2007[11]	Cohort study, all subjects immunised and blood taken before and 2 months and 1 year after vaccination.	TST negative, healthy students in Sweden either never vaccinated (n=15) or previously	BCG vaccine	Lymphocyte transformation stimulated with PPD, ConA and unstimulated.	Lymphocyte transformation stimulated with ConA and unstimulated.	СРМ	Non-specific data not reported	

Gruber et al	Prospective cohort of	vaccinated (n=16), total n=31. Part of a neonatal	Children at high risk for	Cytokine levels from supernatants, stimulated as above Atopic manifestations	Unstimulated and ConA stimulated cytokine levels Atopic manifestations	% with specified	NS	
2000[12]	vaccinated and unvaccinated children. Samples taken at birth, 12, 24, 36, 60, 72 and 84 months.	birth cohort in Germany, (MAS-90 study group), total n=1314. Included in this analysis, n=774	TB were vaccinated with BCG,median age 30days (range 1-343) n=169. Included in this analysis, n=92.	Total and specific IgE	Total and specific IgE	symptoms kU/L	NS (NB percentage of positive IgE tests from total tests performed was lower in BCG groups in first 3 years then higher at 5 and 7 years)	
Hoft <i>et al</i> 1998[13]	Double-blinded, randomized controlled study	Healthy adults (n=54, age 18-45 years)	Connaught BCG (n=18), tice BCG (n=18), placebo (n=18)	T cell proliferation to mycobacterial antigens and controls, T cell subset expansion proliferation to mycobacterial antigens and controls, T cell subsets following IL-2 and IPP stimulation	T cell proliferation responses to media and tetanus toxoid T cell subset expansion to media and tetanus toxoid T cell subsets following IL-2 and IPP stimulation	fold increase dpm of day 56 compared to day 0 mean % of cells Absolute number of CD4+, CD8+ and γδ T cells	NS Increased CD4 (p<0.01), CD8 (p=0.02) and γδ (p=0.03) T cell proliferation following IL-2 plus IPP in BCG responders.	
Hoft <i>et al</i> 1999[14]	Double-blinded, randomized controlled study and an open label study	Healthy adults (n=66, age 18-45 years)	Connaught BCG (n=12, open label, n=18 double blind), Tice BCG (n=18 double blind), placebo (n=18 double-blind)	T cell proliferation, IFN- γ and IL-4 production, cytotoxicity assays, and mycobacterial antibodies	T cell proliferation to PHA and media alone IFN-γ and IL-4 ELISpot responses from PBMCs incubated with media alone IFN-γ and IL-4 ELISAs of culture supernatants of PBMCs with media alone	dpm spot forming cells/1^4 cells pg/ml	Non-specific data not reported	
Hussey et al 2002[15]	Five groups with different vaccine timing and methodologies	Healthy newborns recruited from primary care services	Danish BCG at birth intradermally n=11, Danish BCG at 10 weeks intradermally n=11, Japanese BCG at birth intradermally n=10, Japanese BCG at birth percutaneously using	PBMC proliferation, cytokine responses and cytotoxicity	PHA and TT proliferation responses TT IFN-γ, IL-10 and IL- 5 responses	pg/ml pg/ml	NS NS	

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			Bignell tool n=10, Japanese BCG at birth					
			percutaneously using					
Kagina et al	Single centre,	Healthy newborns	Biovac tool n=20. BCG vaccination at birth	Whole blood	Whole blood	% of cytokine	NS (in	
2009[16]	randomized study	(n=46)	(n=25) or delayed until 10 weeks of age (n=21)	intracellular cytokines (IFN- γ , TNF- α and IL-2), and Lymphocyte phenotyping of BCG specific cells	intracellular cytokines (IFN-γ, TNF-α and IL-2)	positive CD4+ T cells	supplementary data)	
Kleinnijenhuis et al 2012[17]	Open label cohort study	Healthy adults aged 20-36 years scheduled to receive BCG due to travel to TB endemic regions (n=20).	BCG vaccination	Cytokines in response to TB, <i>S. aureus</i> and <i>C. albicans</i> and phenotype of circulating monocytes.	IFN-γ, TNF-α and IL-1β production from PBMC cultures stimulated with S. aureus and C. albicansCD14+ cellsTLR4andCD11b surface expression	fold induction % of total MFI	Sig increase from baseline at 2w and 3m (except S. <i>aureus</i> and IL-1 β at 2 weeks), p<0.05 or 0.01 Sig increase at 2 weeks (p<0.05) TLR4: Sig	
					IL-1 β and TNF- α mRNA expression	Fold induction	decrease at 2 weeks (p<0.05) and increase at 3m (p<0.005) CD11b sig increase NS	
Lalor <i>et al</i> 2009[18]	Combination of a case-control study in the UK and a cohort in Malawi	Healthy infants from the UK (n=117) and Malawi (n=615)	UK - BCG vaccination compared to unvaccinated age-match controls Malawi – BCG vaccination	IFN- γ responses to PPD	IFN-γ responses from whole blood cultures stimulated with control antigens	pg/ml	Statistical comparisons not reported	Unpublished data included in analysis
Lalor <i>et al</i> 2010[19]	Case-control study comparing vaccinated and unvaccinated infants 3 months after vaccination.	UK infants, n=28	Cases: BCG vaccination at 5-10 weeks (mean 7 weeks) Controls: no BCG.	IFN-γ production stimulated a by PHA, TB PPD or unstimualted. 21 cytokine and chemokines in supernatants stimulated by TB PPD or unstimulated	IFN-γ production stimulated by PHA, or unstimulated. 21 cytokines and chemokines in unstimulated supernatants. supernatants. supernatants.	pg/ml	Unstimulated control data not reported.	Infants had previously taken part in an IFN gamma study
Lalor <i>et al</i> 2011[20]	Sub-group of a larger study consisting of a combination of a case- control study in the UK and a cohort in Malawi	unvaccinated) and from Malawi (n=40 vaccinated)	UK - BCG vaccination compared to unvaccinated age-match controls Malawi – BCG vaccination	Cytokine, chemokine and growth factor responses.	Large panel of cytokine and chemokine responses (multiplex bead array) from whole blood cultures stimulated with control antigens	pg/ml	Control data not reported	
Libraty <i>et al</i> 2014[21]	Case-control study (nested into a dengue virus study)	Infants who had not received BCG in the first 2 weeks of life	BCG in first two weeks or delayed until after first vaccination with	IFN-γ ELISpot. Flow cytometry	IFN-γ ELISpot to TT, Polio, HBsAg and PHA. T cell intracellular	SFC/10 ⁶ cells % cells	BCG vaccinated infants had increased IFN-γ	

		(n=13) and age-and sex match infants who did receive BCG in the first two weeks (n=38)	DTP		cytokine staining for; IFN-γ+/TNF- α+/CD4+/CD4RO+/- FoxP3+TNF- α/CD4+/CD4RO+/-		ELISpot to TT (p=0.046) and IFN- γ +/TNF- α +/CD4+/CD4RO + T cells (p=0.018)	
Lowry <i>et al</i> 1998[22]	Randomised, placebo- controlled trial comparing different doses of BCG with saline. Blood samples before, 2 and 16 weeks and 1 year after vaccination.	Healthy adults aged 18-44 years born in the USA, not previously vaccinated with BCG, n=76	Very low dose BCG, low dose BCG, standard dose BCG or high dose BCG or saline placebo. 5 subjects from very low, low and placebo groups were revaccinated at 18 weeks.	Adverse events. PPD skin tests. Lymphopriliferation to M TB, Ag85, BCG, M. avium and ConA. IL-4 and IFN-γ stimulated as above IgG sero-reactivity to BCG-SA antigen	Lymphoproliferation to Con A IL-4 and IFN-γ to ConA (not reported)	CPM Not reported	NS (p value not reported) Not reported	
Marchant <i>et al</i> 1999[23]	Single centre, randomised, controlled trial	Healthy newborns (n=137)	BCG vaccination given at birth, 2 months or 4 months.	Whole blood and PBMC proliferation and cytokine responses	lymphoproliferation IFN-γ, IL-5, IL-13 and IL-4 responses following culture with PHA	Stimulation index pg/ml	NS NS	
Marks et al 2003[24]	Retrospective case- control study across two regions of Sydney, Australia	Children from a region where BCG vaccination was routine $(n=309, age 9.5+/-2.4 years)$ and another region where it was not $(n=442, age 9.7+/-2.0)$.	BCG vaccination compared to unvaccinated	Cytokine responses, Clinical components of allergy	IL-4, IL-5, IL-10 and IFN- γ secreted in response to house dust mite stimulation of PBMCs Serum total IgE	Stimulation index, median with bow and whisker plots Geometric mean kIU/L	IL-10 SI significantly lower for BCG recipients, p<0.0001. NS for other cytokines Comparison not reported	
Miles <i>et al</i> 2008[25]	Cohort study, all infants vaccinated and part of a larger CMV cohort study.	Male and female infants in the Gambia taking part in a CMV study, all infected with CMV, n=133 (though not all sampled at all time-points). Children aged 4-5 years in the Gambia, n=25 and adult females aged 21-31 years who had given birth 18 months ago , n=11.	Infants were immunised at birth with BCG. All children were recorded to have received BCG. Adults assumed to have had BCG.	IFN-γ, IL-2, CD154 responses of CD4 T cells stimulated with PPD or normal human dermal fibroblast lysate (NHDF as required by CMV study)	IFN-γ IL-2, CD154 responses of CD4 T cells stimulated NHDF (not reported).	% of CD4 cells expressing the cytokine, median with interquartile range.	Not reported	Part of a larger CMV study and all infants had CMV infection.
Miles <i>et al</i> 2010[26]	Single centre, case- control study examining maternal HIV status on newborns response to BCG	Newborns of HIV positive (n=16) and negative (n=21) Malawian women	BCG vaccination at birth (OPV also given).	IFN- γ responses to BCG and PPD, cellular phenotypes. PHA controls.	CD4 counts in HIV negative infants born to HIV negative mothers CCR7 and CD45RA expression	Median with IQR Cells/ml blood x 10^5	Significance reported between groups (based on HIV status of mother), all of whom were vaccinated.	

Ota <i>et al</i> 2002[27]	Single centre, randomised trial	Healthy newborns (n=151)	BCG vaccination given at birth, 2 months or 4.5 months. Routine vaccine schedule (OPV, DTP and Hepatitis B vaccine) also given	Proliferation, cytokine and vaccine specific antibody responses	CD27 and CD28 expression PD-1+ and CD57+, CD4+ T cells IFN-γ release by PBMC s PBMC proliferation to HBsAg, TT and PHA IL-5, IL-13 and IFN-γ PBMC responses to HBsAg, TT and PHA	Cells/ml blood x 10^5 % Spot forming units per 10^6 PBMCs Stimulation index pg/ml	Significant between groups Significant between groups	
					Antibody responses to HBsAg, TT, PV1, DT	mIU/ml	Significant between groups	
Smith <i>et al</i> 2012[28]	Two case-control studies: one in infants the other in adolescents	UK born BCG vaccinated infants (n=21 mean age 11.5, SD 6.8 weeks) and unvaccinated (n=18, mean age 13, SD 3.4). Adolescents vaccinated with BCG (n=16) and unvaccinated (n=20)	BCG vaccinated compared to unvaccinated	Cytokine and chemokine responses to PPD and Heparin-binding haemagglutinin	PHA used as positive control (reported in supplemental data).	Pg/ml Median with interquartile range	NS	
Soares <i>et al</i> 2013ar[29]	Cohort study	Healthy newborns (n=90)	BCG vaccination at birth	Intracellular cytokine expression of T cells.	PMA, TT, PHA stimulated cytokine responses	Data not reported	Not reported	
Steenhuis et al 2008[30]	Randomized, prospective, single blinded (researcher) trial to determine effect of BCG on allergic disease.	Mainly Caucasian newborns with mother or father and sibling with allergic disease (n=121)	BCG vaccination or placebo (normal saline) at 6 weeks of age	Prevalence of allergic disease at 18 months.	Prevalence of allergic disease at 4 and 18months. Leucocytes and eosinophils in peripheral blood	Questionnaire, SCORAD scoring, lung function Mean leucocytes and eosinophils ±SD	NS	19 participants had BCG repeated at 4 months as they had no scar and negative skin test.
Tastan <i>et al</i> 2005[31]	Randomised trial comparing vaccination at birth or 2 months. Blood taken at 48 hours (group 1 only) 2 months and 4 months (group 2 only) of age.	Healthy, male and female, term newborns in Turkey, n=40.	Group 1: BCG vaccine at birth, n=20 Group 2: BCG vaccine at 2 months, n=20 NB all received routine immunisations including DTP and polio at 2, 4 and 6 months	Total lymphocytes	Total lymphocytes, αβ+ T cells and γδ+ T cells	Cells/µl, median value with quartiles.	$\begin{array}{llllllllllllllllllllllllllllllllllll$	
van den Biggelaar <i>et al</i> 2009[32]	Case-control	Healthy newborns from PNG (n=50) and Australia (n=50). Healthy Australian adults as	BCG vaccination in PNG newborns. No BCG vaccination in Australian participants	Mononuclear cell culture cytokine production. mRNA expression. Lymphocyte phenotyping	Cytokine (IL-10, IFN-γ, TNF-α, IL-5) responses to PHA	Data not reported	Not reported	

		controls (n=15)						
Vargas <i>et al</i> 2004[33]	Randomised, placebo controlled trial investigating effect of BCG on asthma. Samples taken before and 12 months after vaccination.	Asthmatic schoolchildren, male and female, in Mexico, n=82 (67 completed study)	BCG, n=33 or placebo (saline), n=34. NB all children were treated for their asthma for 12 months prior to vaccination	Symptoms questionnaire Leukocyte count, eosinophil count, IgE, esosinophils in nasal mucus and parasites in stools. Subset (17 vaccinated and 18 placebo) had IL-4 and IFN-γ from phorbol myristate acetate and ionomycin stimulated PBMCs	Symptoms questionnaire Leukocyte count, eosinophil count, IgE, esosinophils in nasal mucus and parasites in stools. IL-4 and IFN-γ from phorbol myristate acetate and ionomycin stimulated PBMCs. Stimulated PBMCs.	Cells/mm3, IU/ml Pg/ml	Within group sig change over time for controls but not for vaccinated for IgE, IL-4 and IFN-γ. Within group sig decrease in monocytes, eosinophiles and % eosinophils in nasal cytology for BCG.	
Vekemans <i>et al</i> 2004[34]	Single centre randomized study in	Healthy newborns (n= 29)	BCG at birth compared to 2 months of age	PBMC IFN-γ responses to mycobacterial	PBMC IFN-γ production to PHA and TT (not	pg/ml	Betweengroupcomparisonsnotdone.Not reported	
200101	newborns and a case- control study in adults	Adults with tuberculosis (n=33), TB exposed (n=21), living in the vicinity of TB subjects (n=22) and heavily TB exposed health care workers (n=23)	Adults were not vaccinated	antigens and controls PBMC IL-13 production (in adults)	reported for newborns)			
Vijaya Lakshmi V, <i>et</i> <i>al</i> 2005[35]	Case-control study (vaccination was performed retrospectively)	Children aged 5-7 years who had been vaccinated with BCG (n=45), unvaccinated (n=31) or had active TB (n=31)	BCG vaccination compared to unvaccinated and active TB patients (vaccination was retrospective)	Lymphoproliferation and cytokine responses	IL-2 dependent cell line proliferation supplemented with supernatant from lymphocyte culture stimulated with antigens IFN-γ lymphocyte cultures supernatant of lymphocyte cultures stimulated with control antigens	Stimulation index	Higher stimulation index of lymphocyte transformation for vaccinated vs unvaccinated healthy children (p<0.05) Increased IFN-γ in vaccinated children (p<0.01)	
Weir <i>et al</i> 2004[36]	Randomised, controlled trial, multiple regions	Healthy young adults from Northern Malawi (n= 633, mean age 19 years) and East London and Essex, United Kingdom (n=424, mean age 14 years)	BCG vaccination compared to placebo	Cytokine responses to mycobacterial antigens.	Cytokines (TNF-α, IL- 10, IL-1β) from whole blood culture supernatants	Median responses pg/ml	TNF-α- NS IL-10 decrease from baseline in Malawi subjects for <i>M.</i> <i>scrofulaceum</i> and <i>M. vaccae</i> only (p=0.015 and 0.02)	Study carried out in the context of the Karonga Prevention Study, a large vaccine trial and epidemiological study of tuberculosis and leprosy in the Karonga district.
Weir <i>et al</i> Oct. 2008[37]	Single region case- control study	Healthy young adults from East London and Essex,	BCG vaccination compared to unvaccinated	IFN-γ responses to mycobacterial antigens.	IFN-γ responses to control antigens	%>63pg/ml and %>250pg/ml	NS	

		United Kingdom (n=424, mean age 14 years)						
2008[38]	Follow-on study of three separate, open label randomized trials	UK adolescents. Unvaccinated 12-14 year olds n=213 Vaccinated 3 years prior, 17-18 year olds n=20 Vaccinated in first year of life, 12-14 year olds n=43 with 212 similar aged children who were unvaccinated as controls	BCG vaccination compared to unvaccinated	IFN-γ from supernatants of whole blood assays stimulated with mycobacterial antigens and controls	Controls used for IFN-γ from supernatants of whole blood assays	pg/ml	NS	

Table B. Characteristics of included measles vaccine studies

Author	Methods	Participants	Interventions	Outcomes	Non-specific outcomes	Method of reporting non- specific outcomes	Difference in NSE outcome	Notes
Bertley <i>et al</i> 2004[39]	Follow on to a randomised, controlled trial. Samples taken before vaccination, at 9 months and at 5 years.	5 year old children in Sudan who had previously taken part in a measles vaccine RCT. Recruited from 6/14 of the communities in the original trial.	At 5 months of age: Connaught high titre measles vaccine, n=61 or meningococcal A+C vaccine as control, n=59. All received standard titre Schwarz vaccine at 9 months. A third group from the original trial (single high dose EZ vaccine) were not included.	Neutralizing antibodies. Lymphoproliferation to measles virus (at 5 years only)	Lymphoproliferation to Vero control stimulus.	CPM	NS (comparable across groups, details not given)	Follow-on study
Gans <i>et al</i> 1999[40]	Cohort study, all infants vaccinated and blood taken before and 12 weeks after immunisation.	Healthy infants in the USA aged: 6months (N=60), 9 months (N=46) 12 months (N=56) And Healthy adults aged 20-40 years who had previously had at least one measles vaccine (N=29).	6 and 9 month old infants: Monovalent Edmonson strain measles vaccine (Attenuvax) 12month old infants: MMR Adults: no vaccination given	Measles specific T cell proliferation, IL-2 and IFN- γ production. Effect of passive measles antibody and rhIL-12 on above. T cell proliferation to PHA (not fully reported). Measles antibody titres.	IL-12, IFN-γ, and T cell proliferation to PHA (not fully reported).	Mean CPM with standard error and pg/ml.	NS (p value not given)	Post vaccine samples available for 134/162 infants but not all tested for all assays. Non-specific effects not fully reported.
Gans <i>et al</i> 2004[41]	Case control study. Cases received measles vaccine at 6 or 9 months then MMR at 12 months. Blood taken before and 12 weeks after first vaccination then 24 weeks after MMR. Controls enrolled at 12 months. Bloods taken before and 24 weeks after MMR.	Cases: Infants receiving well-child care in California aged 6 (N=32) or 9 (M=23) months. Controls: 12month infants (N=83)	Cases: Monovalent Measles vaccine (Attenuvax) at 6 or 9 months followed by MMR at 12 months Controls: MMR at 12 months	Measles antibody, (passive and non) after 1 and 2 doses. T cell proliferation to measles antigen and PHA (not reported).	T cell proliferation to PHA (not reported).	Results not reported	Not reported	Not all sample sizes allowed for full immunological evaluation. Non-specific effects not reported.
Hennino et al 2007[42]	Prospective, double- blinded, randomized, placebo-controlled	10-14month-old males and female infants in the USA with atopic dermatitis, N=12,	Single Schwartz strain measles vaccination at 10-14months or placebo (physiological serum)	Severity of atopic dermatitis, seroconversion, serum levels of CCL18 and E- selectin 1 month post vaccination.	Severity of atopic dermatitis, seroconversion, serum levels of CCL18 and E- selectin 1 month post vaccination	Individual SCORAD eczema severity score for 6 randomly chosen infants. Serum levels of CCL18 and E- selectin in ng/ml for six participants before and after treatment.	NS CCL18 decrease in 2 individual measles treated subjects compared to baseline (p=0.0183 and 0.0011)	All children had atopic dermatitis
Hussey <i>et al</i> 1996[43]	Cohort study with 3 groups (different	Healthy infants attending 2	Group 1: EZ vaccine (medium titre) at 6	Measles antibody responses. Proliferation,	Proliferation, IL-2 receptor, CD4, CD8, β2	Median CPM, U/ml (sIL-2r, CD4, CD8),	Decrease in PHA proliferation in	StudycommencedbeforeWHO

	vaccine and ages). Blood taken before, 2 weeks and 3 months after vaccination.	immunisation clinics in Cape Town, SA (one in high measles prevalence area and one in low measles area) Group 1: (high prevalence) 6 months old, N=38. Group 2: (high prevalence) 6 months old, N=26. Group 3: (low prevalence) 9 months old, N=24.	months Group 2: Schwarz vaccine (medium titre) at 6 and 9 months Group 3: Schwarz vaccine at 9 months.	IL-2 receptor, CD4, CD8, β2 microglobulin and Neopterin in response to PHA. Lymphocyte subsets. Comparison between males and females.	microglobulin and Neopterin in response to PHA. Lymphocyte subsets.	mg/L (β2 microglobulin) and nmol/L (Neopterin).	Schwarz strain groups at 3 months post immunization (p=0.013, 0.002). Decrease in PHA proliferation at 2 months post immunisation in Schwarz strain at 9 months group (p<0.001). Increase in soluble CD8 (p=0.02) and $\beta 2$ microglobulin (p=0.04) from baseline in Schwarz strain at 9 months group. Difference between all groups for soluble IL-2 receptor (p<0.001), soluble CD4 (p=0.015) and absolute CD8 count (p=0.008).	recommendation to withdraw EZ vaccine
Jaye <i>et al</i> 2014[44]	Case-control study (disease versus vaccinated)	Children and adults with acute measles. Healthy newborns (n=22)	Measles vaccination at 9 months of healthy children recruited at birth	Cytotoxic T cell assays	Controls used for cytotoxic T cell assays	Data not reported	Not reported	
Liguori et al 1998[45]	Cohort study, All children vaccinated and bloods taken before, 5 and 15 days after vaccination.	Healthy children in Italy, males and females, aged 5-9 years with no history of measles, N=20.	Single Schwarz strain measles vaccine (Rimevax) at baseline.	Measles IgG and IgM. Serum TNF–α, IFN-α and IFN-γ.	Serum TNF $-\alpha$, IFN- α and IFN- γ .	pg/ml for each individual subject.	NS	Funding source not reported
Lisse <i>et al</i> 1994[46]	Case-control, follow- on study of 2 previous trials.	All participants from capital of Guinea- Bissau Cases: From trial 1- all children who had received medium titre EZ vaccine at 4months, now aged 5 years. From trial 2-children who had received either medium or high dose EZ vaccine at 4 months, now aged 3-4years. Controls: Children from previous trials who received standard dose	Follow-on study, no new interventions	Total white cells and lymphocyte subsets (presented for each trial and by sex).	Total white cells and lymphocyte subsets (presented for each trial and by sex).	Median cell counts/% cells with interquartile range.	Lower percentage of lymphocytes and CD4:CD8 ratio and higher percentage and total CD8 cells in females receiving high titre EZ strain (p<0.05).	330 of the 854 originally recruited provided samples for this follow- on study. Mortality data is presented elsewhere in Aaby <i>et al</i> 1993

		Schwarz vaccine at 9months.						
Njie-Jobe <i>et al</i> 2012[47]	Randomised trial comparing children vaccinated at 4 and 9 months or just 9 months. Bloods at baseline, 4, 9 or 9.5 (randomized to tests for memory or effector response), 18, 36, 36.5 and 48 months.	Gambian infants, male and female, initially recruited at 4 months of age, N=132	Group 1 (N=64): Measles vaccine at 9 and 36 months Group 2 (n=68): EZ measles vaccine at 4,9 and 36 months NB both groups also received routine EPI vaccines including DTP, HepB, Hib, oral polio and at 9 months, yellow fever.	MeaslesHAI.IFN γELISpoteffectorresponsestomeaslestomeaslesfusionpeptidesandPHA.FOXP3mRNA expression.rpression.γ,IL-10,IL-2Rα andMP-1β levels.CD8 andCD4cellsexpressingIFN γ and/or CD69.NBNotallassaysperformedatalltime-points.	ELISpot IFN-γ response to uninfected Vero cells and PHA (data not reported). Plasma IFN γ, IL-10, IL- 2Rα and MIP-1β levels.	Median cytokines level in pg/ml with interquartile range.	NS	
Okada <i>et al</i> 2001[48]	Case-control comparing age- matched measles patients with 1-2 year- old children immunized with measles vaccine.	Cases: uncomplicated, acute measles patients in Tokyo area, aged 1-2 years. N=147. Controls: healthy children aged 1-2 years who received measles vaccine. N=32.	Controls received measles vaccine, either AIK-C strain or Schwarz strain, further details not reported.	Total lymphocyte count, CD4, CD8 and B cells. Evidence of apoptosis of PBMC (through analysis of DNA fragmentation). NK cells (data not reported). Measles antibody responses.	Total lymphocyte count, CD4, CD8 and B cells. Evidence of apoptosis of PBMC (through analysis of DNA fragmentation). NK cells (data not reported).	Cells per µL, each point representing average of 3-15 samples. Mean % apoptosis and cell surface expression of Fas, FasL or TRAIL-Rs. Mean cytokine levels in pg/ml (ng/ml for Sol- FasL).	NS	NB for purposes of this SR, only the results for the vaccinated cohort have been included
Ovsyannikova et al 2003[49]	Cohort study with two groups of different ages. Randomly assigned to undergo single phlebotomy on one day 0-40 days post intervention.	Healthy infants/children in the USA, male and female. Group 1: 12- 15month-old, N=15. Group 2: 4-12year- old, N=42	Single Edmonson strain measles vaccination	PHA stimulated cytokine production (IL-2, IL-4, IL-6, IFN-γ and TNF-α), plasma cytokine levels, measles antibody titres.	PHA stimulated cytokine production (IL-2, IL-4, IL-6, IFN-γ and TNF-α), plasma cytokine levels.	Median cytokine concentration in pg/ml (different children at each time-point) with interquartile range.	In children decrease in IFN- γ to PHA at day 20. Higher overall median IFN- γ to PHA in children compared to infants Decrease in median plasma IFN- γ (p=0.0027), TNF- α (p=0.0001), and sIL2-R (p=0.0001) in children compared to infants.	Each subject had single blood draw therefore results at each time point are different children. Day 0 children (N=2 and 7) did not receive vaccine before blood draw.
Pabst <i>et al</i> 1999[50]	Case-control comparing children of mothers who either had natural measles or were vaccinated against measles plus separate cohort vaccinated with	Group 1: 6 month- old infants whose mothers were assumed to have had measles, N=61. Group 2a: 6 month- old infants whose mother were known	Group 1 and 2a: Single dose of standard titre AIK-C measles vaccine at 26-32 weeks Group 2b: Single dose of standard titre Connaught (CLL) measles vaccine at 26-32 weeks	Reactogenicity. Measles specific antibody. Blast transformation to measles HA antigen. Production of IFN-y and IL-10 stimulated by measles antigen, TT, non-measles infected	Production of IFN-γ and IL-10 stimulated by PHA. Blast transformation to TT, non-measles infected Vero cells and Candida.	Pg/ml, mean ± standard deviation. Mean CPM ± standard deviation.	NS	Measles vaccine was given immediately after the third dose of DTP- Polio-Hib. Part funding provided by Merck-Frost, Canada

	different measles	to be vaccinated		Vero cells, PHA and				
	vaccine. All children vaccinated, bloods	against measles, N=119.		Candida				
	taken before and 8	Group 2b: 6 month-						
	weeks after	old infants whose						
	vaccination.	mothers were known to be vaccinated						
		against measles,						
		N=120						
Samb <i>et al</i> 1995[51]	Case-control study.	Rural Senegalese children who had received EZ vaccine at 5 months (n=73) and children who had received placebo at 5 months followed by Schwarz vaccine	Rabies vaccine	Immunogenicity to measles, yellow fever and rabies vaccines. Skin tests. Haematological parameters.	Rabies and Yellow fever immune responses. CD3, CD4, and CD8	YF – Log2 neutraliazation antibodies Rabies – Log10 reduction neutraliazation	NS Females who received EZ had a higher rabies neutralization (p= 0.012) and ELISA	
		(n=70)			lymphocyte counts	x10^6 cells/L	(p=0.03) antibody.	
Schnorr et al 2001[52]	Quasi-RCT comparing unmatched controls with those immunised at 4 and 9 months or 9 months only. Comparison of Blood at recruitment for controls or 7 days after vaccinated groups and at 6 and 24 weeks for all groups.	Bangladeshi infants aged 6 and 9 months of age. NB unmatched controls median age 10.1 months (range 6- 18m). Total n= 78	Group V6: Standard titre measles vaccine (2/3 had standard titre EZ and 1/3 had Schwarz) at 4 and 9 months, n=24. Group V9: As above at 9 months only, n=25 Controls: no vaccination, n=29. All received Vitamin A on recruitment.	Measles antibody levels. Delayed hypersensitivity skin test. Cytokine responses in response to stimulation with PHA.	Delayed hypersensitivity skin test. Cytokine responses in response to stimulation with PHA.	Geometric mean cytokine production I pg/ml	Increased anergy to candida in DTH assay (p=0.015 for 6 month group) and p=0.04 for 9 month group) Difference in CD71 (p=0.04), and CD30 (p=0.004) expression between groups at baseline. Difference in expression of CD25 (p=0.02), CD69 (p=0.04), CD71 (p=0.04), and CD30 (p=0.006) between groups from week 0 to week 6. Difference in expression of NK (p=0.009) and CD69 (p=0.03) between groups at week 24. Increased IL-2 (p=0.07) at 6 weeks and IL-10 (p=0.04) at 24 weeks between groups.	All received vitamin A on recruitment

Table C. Characteristics of included MMR vaccine studies

Author	Methods	Participants	Interventions	Outcomes	Non-specific outcomes	Method of reporting non- specific outcomes	Difference in NSE outcome	Notes
Nakayama <i>et al</i> 1990[53]	Cohort study. All children vaccinated and blood taken before and 6 weeks after vaccination. A subset of children also had bloods at 3,7,10 days and 2,3,5 and 10 weeks after vaccination.	Healthy children who visited outpatients in Tokyo aged 1-5years, N=229.	MMR vaccine containing measles AIK- C strain, Mumps Hoshino strain and Rubella Takahashi strain, single dose at baseline.	IFN- α responses to measles, mumps and rubella- subset of 11 children only. Measles and Rubella serum antibodies (paired serum for all subjects).	Non-stimulated controls (not reported).	Results not reported.	Not reported	No non-specific effects are reported and only non-stimulated control experiments were performed. Funding source not reported
Pabst <i>et al</i> 1997[54]	Cohort study, all infants vaccinated and blood taken before and either 14 (N=32), 22 (N=32), 30 (N=27) or 38 (N=33) days after vaccination.	12 month infants from Edmonton, Canada, N=124	MMR given within 4 weeks of first birthday	Blast transformation of PBMC to measles antigen, Vero cell control antigen, TT and Candida antigen. Production of sIL-2r, IFN- γ , IL-4 and IL-10 stimulated by PHA. CD4, CD8 and NK cells.	Blast transformation of PBMC to Vero cell control antigen, TT and Candida antigen. Production of sIL-2r, IFN-γ, IL-4 and IL-10 stimulated by PHA. CD4, CD8 and NK cells.	CPM, mean and standard error of mean. pg/ml, mean and standard error of mean. % of PBMC, mean and standard error of mean.	Significant change over time Significant change over time Significant change over time	Not all children assessed at each time point. Part funding provided by Merck-Frost, Canada
Rager-Zisman et al 2003[55]	Cohort study, all children vaccinated and blood taken before and 30 days after vaccination.	Healthy Israeli children; age 6.14 (±0.35) years with previous MMR vaccination in infancy. N=38	Single MMR vaccination (MMRII) Note: subjects had previously received MMR in infancy and had recently had a tetanus booster	Measles, mumps and rubella IgG, total IgM, IgG and IgE. White cell count, CD8, CD4 and CD4:CD8. Lymphoproliferative responses to PHA and TT. NK cells and NK specific activity.	White cell count, CD8, CD4 and CD4:CD8. Lymphoproliferative responses to PHA and TT. NK cells and NK specific activity.	Median white blood cell count (x1000/µl), % CD4, %CD8 and CD4:CD8 ratio with interquartile range before and after vaccination. Mean and standard error of the mean CPM. Mean and standard error of mean %CD56 cells.	Sig decrease in total leukocytes (p<0.0001), %CD4 (p=0.028) and %CD8 cells (p=0.041) before and after immunisation. Response to TT sig increase (p=0.006) Sig increase in CD56+ cells (p=0.01)	Number of children tested varies by assay (N=28-38)

Table D. Characteristics of included tetanus toxoid vaccine studies

Author	Methods	Participants	Interventions	Outcomes	Non-specific outcomes	Method of reporting non- specific outcomes	Difference in NSE outcome	Notes
Armitage et al 1993[56]	Case-control, comparing responses in elderly to young adults	Healthy elderly (n= 23, mean age 75) and young adults (n=23, mean age 27)	Tetanus toxoid vaccination depending on prior vaccination history	Immunogenicity, proliferation and blastogenesis	Lymphoproliferation to PHA and ConA	cpm	Decrease in ConA and PHA blastogenesis in elderly vs young adults (p=0.01 and p=0.001)	
Borut <i>et al</i> 1980[57]	Case-control, comparing agammaglobulinaemic subjects to immunised and unimmunised subjects. Retrospective vaccination	Agammaglobulinae mic (n=7, aged 12- 27 years), unimmunised (n=9, aged 4-20 weeks), and immunised (n=64, age 8 weeks - 50 years)	Vaccination status determined by history (any individual who received TT within 10 years prior to the study was considered immunised)	Skin testing, proliferation, monocyte chemotaxis and immunogenicity	Monocyte chemotaxis	Number of cells per high powered field	Higher monocyte chemotaxis in those with positive Tetanus toxoid skin test, p<0.01.	
Chollet <i>et al</i> 1979[58]	Case-control	Healthy adults (n=24 in the vaccine arm and n=51 controls)	Boosting with tetanus toxoid compared to no boosting	Lymphocyte proliferation, phenotyping (by electrophoresis), cytotoxicity, and rosettes	B, T1 and T2 cell phenotypes by electrphoresis Lymphoproliferation to PHA, PWM, ConA	% Stimulation index	Significant increase in T2 population over time in 6 of the vaccinated group.	
Chui <i>et al</i> 2004[59]	Case-control	Healthy adults cases (n=6, 39.8±7.2 years) and controls (n=6, 36.8±5.4 years)	Flt3-ligand and tetanus toxoid vaccination compared to tetanus toxoid vaccination only	Skin tests, proliferation, IFN- γ and immunogenicity.	Lymphoproliferation to PHA IFN-γ ELISpot to PHA, HBsAg and media alone	Stimulation index Spots per 100 000 cells (not reported)	Not reported	
Cooper <i>et al</i> 1998[60]	Case-control study conducted within two Ecuadorian communities	Onchocera volvulus infected (n=19, median age 36, range 15-75 years) and uninfected (n=20, median age 36, range 17-67 years) adults	Tetanus toxoid vaccination (2 doses 1 month apart) in both cohorts	Proliferation, cytokines and immunogenicity	Positive and negative controls of PBMC proliferation assays (data not reported in manuscript). PBMC cytokine production	Antigen stimulated assays were reported as geometric mean stimulation index with 95% CIs Median (pg/ml) and 95% CIs	NS	
Di Genova <i>et al</i> 2006[61] (Also BCG)	Cohort study	Healthy adults (n=12, age range 31- 44 years), all had previously received TT vaccination over 5 years prior. All subjects had also received BCG vaccination	Tetanus toxoid vaccination	PBMC and T cell proliferation and cytokine responses	Memory T cell proliferative responses to <i>Candida albicans</i> IFN-γ, IL-2 and IL-13 responses to <i>C. albicans</i> , and controls	Mean stimulation index ± SD Number of spots/2x10^5 PBMC	Individual results given, overall significance not reported.	
Fernandez <i>et al</i> 1994[62] Fevrier <i>et al</i>	Cohort study Cohort study	Healthy males (n=3, age range 25-40 years)	Tetanustoxoidvaccinationinsubjects(1subjects(1receivedTT2subjectsprior, and 2subjectsreceivedTT8-9yearsprior)Tetanustoxoid	Immunogenicity, proliferation, cytokine production Lymphoproliferation	Lymphoproliferation to PPD PBMC production of TNF- β and IL-2 in the presence of media \pm IL-2 Positive and negative	CPM Number of positive cells/100 000 PBMCs Mean cpm ± SD	Not reported	

1977[63]		range 30-40 years)	vaccination. 2 subjects	(including B and T cell	control lymphocyte			
1977[03]		lange 50 40 years)	never previously	subsets)	(non-separated, B and T			
	<u> </u>		vaccinated.	DDMC 11	cell) proliferation			
Gentile <i>et al</i> 2006[64]	Case-control study	Adults with (n=15, mean age 27 years) and without (n=15, mean age 27 years) allergic rhinitis	Tetanus toxoid vaccination in all subjects	PBMC cytokine responses, skin testing to allergens and TT immunogenicity	IFN-γ and IL-13 responses to PHA	Mean (pg/ml)	PHA and TT induced IFN- γ were increased on days 7 and 14 for the non-allergic rhinitis vs allergic rhinitis group (p<0.05). PHA induced IL- 13 was increased on day 7 for the non-allergic rhinitis vs allergic rhinitis group (p<0.05)	
Livingston <i>et al</i> 2013[65]	Single centre, cohort study	Healthy adults (n=8 mean age 33 ±10 years)	Tetanus toxoid vaccination.	PBMC proliferation, cytokine responses, cellular phenotyping and <i>in vitro</i> antibody production	PBMC proliferation to PBS CD4+CD3+ T cell Proliferation to PBS Intracellular CD4+CD3+ T cell IFN-γ, TNF-α, IL- 2, IL-17, IL-10, IL-5, IL- 13 and IL-4 produced to PBS Intracellular PBMC IFN- γ, TNF-α, IL-2, IL-17, IL-10, IL-5, IL-13 and IL-4 produced to PBS	Percentage CD25+ and CD69+, CD3+CD4+ T cells Percentage CD3+CD4+ T cells Percentage CD3+CD4+ T cells pg/ml	Significant differences between PBS and TT stimulation only reported.	
Mahalingham et al 2010[66]	Randomised, double blinded, controlled trial	Healthy females (n=108, age range 18-25 years)	Tocotrienol-rich fraction supplementation (palm oil) compared to placebo. All subjects received Tetanus toxoid vaccination	Lymphoproliferation, cytokines in culture supernatants, TT immunogenicity and plasma Vitamin E levels.	IFN-γ and IL-4 production in response to ConA IL-6 production in response to LPS	pg/ml pg/ml	Increase in ConA induced IFN- γ and IL-4 at day 56 for both groups (p<0.001). Increase in IL-6 to LPS in the intervention group at day 56 (p<0.001).	Supported by a grant from the Malaysian Palm Oil Board

Table E. Characteristics of included DTP and DT vaccine studies

Author		Methods	Participants	Interventions	Outcomes	Non-specific outcomes	Method of reporting non-	Difference in NSE outcome	Notes
Dirix <i>et</i> 2009[67]	al	Cohort study. Blood taken at 2 (before vaccination, N=42), 3 (n=28) and/or 6 (N=24) months of age and at around 13 months (N=5)	Infants in Belgium aged 2-13 months with HIV positive mothers but who were HIV negative themselves, N=63.	Infants had previously received Tetravac (tetanus, acellular pertussis, diphtheria, polio vaccine) and H influenza type B polysaccharide vaccine at 2, 3 and 4months and recombinant Hepatitis B vaccine at 3 and 4 months.	PHA, FHA and PT induced IFNγ and IL- 12p70. Spontaneous IL- 10 secretion. Effect on IFNγ secretion in presence and absence of anti-IL-10 antibody. IL- 10 polymorphisms.	PHA induced IFNγ and IL-12p70. Spontaneous IL-10 secretion.	specific outcomes Median pg/ml with interquartile range. NB PHA data reported based on spontaneous IL-10 secreting status. IL-10 data reported for individual infants.	NS/not fully reported	All children had been involved in previous studies on cellular immune responses to pertussis vaccine All had received 6 weeks of Zidovudine prophylaxis until HIV status was confirmed. Part funded by Sanofi Pasteur.
Fernandes <i>e</i> . 2010[68]	t al	Cohort study. Blood taken 7 days before, 7 and 28 days after vaccination.	Healthy males aged 18-55 years in Canada, N=20.	Single dose of tetanus/diphtheria vaccine.	Frequency of B lymphocyte subsets. IgG, IgA and tetanus/diphtheria specific antibody secretion. IgA and IgG antibody to polio virus and HSV.	Frequency of B lymphocyte subsets. IgA and IgG antibody to polio virus and HSV.	Mean absolute number or % cells ±standard error of mean. Spots/1000 PPC for 3 subjects, error bars are standard error of mean.	NS over 28 days Increase in HSV IgG (p=<0.01), IgA (p<0.01), and Polio virus IgA (p<0.005) at day 7 compared to day - 7.	Adult only study. Funding source not reported.
Fryauff <i>et</i> 1998[69]	al	Sub study of anti- malarial RCT. Subjects in anti- malarial trial were immunised while controls (not taking part in malaria trial) were not. Blood taken before, 1 and 7 months after immunisation.	Healthy Adult male transmigrate farmers of Javanese/Sundanese ethnicity, who were taking part in a malaria prophylaxis trial, N=72 and control subjects not having malaria prophylaxis, N=20.	Non-control subjects had previously randomised to weekly chloroquine, daily primaquine or placebo for 12 months. These subjects received Tetanus and diphtheria toxoid vaccine 11 months after starting the assigned drug regimen. Control subjects did not receive any intervention.	Lymphocyte proliferation in response to PHA, TT, M. TB PPD and TT subunit peptides (P2, P30 and P2P30)	Lymphocyte proliferation in response to PHAand M. TB PPD	PHA and PPD data not reported.	Not reported	Adult only study. Sub study of anti-malarial trial.
Halasa et 2008[70] He et	al	Prospective, randomised, pilot study investigating an additional dose of DTP at birth. Safety data collected after every dose and blood samples at birth, 6,7,17 and 18 months.	Infants aged 2-14 days, n=50.	Group 1: DTaP and hep B at birth Group 2: Hep B only at birth. NB Both groups received routine vaccines including DTaP at 2,4,6 and 17 months.	Adverse reactions. IgG to PT, PHA PRN and Fimbriae 2/3. PRP Hib capsule antigen. Pneumococcal capsule antigens, diphtheria and tetantus toxoids. Neutralization assays for polio and hep B surface antibodies (7 months only).	Polio neutralisation assays, pneumococcal serotypes 6, 14, 23 and Hep B antibodies at 7 and 18 month visits.	Geometric mean concentrations	Higher GMC for pneumococcal serotype 14 in controls at 7 months (p=0.035) and higher microneutralisatio n titres in controls for poliovirus 1 and 3 at 18 months (p=0.39 and 0.041)	At 7 months, 2 subjects in the experimental group and 1 control had an additional Hib dose as protective antibody titers not achieved.

1998[71]	comparing children and adults either vaccinated against pertussis or with natural pertussis infection and children/adults who did not receive pertussis vaccine. Blood taken before and 1 month after immunisation	N=8): 13 year old children (N=6) and adults (n =2, aged 26 and 60 years) with culture confirmed pertussis infection. Vaccinated cases (total N=20): Male and female children in Finland, aged 10-12years, N=17. Adult males, N=3. Controls (total N=25) : 10-12year old children who had received DT booster in last month (N=9), Adults (N=8), healthy 13 year-olds (N=6) and newborns (N=2)	diphtheria-tetanus- trivalent acellular pertussis (DTaP) vaccine given to vaccinated cases	to pertussis antigen (PT, FHA and PRN), PHA and pokeweed mitogen (latter two not reported). Cytokine mRNA expression in PBMCs for IFNγ, IL-2, IL-4 and IL- 5. IgG antibodies to PT, PHA and PRN.	to PHA and pokeweed mitogen (not reported). Cytokine mRNA responses to PHA and unstimulated were not reported	PHA and PWM. Medium only reported for individuals as [H ³]thymidine incorporation	NS (p value not given)	controls and cases came from authors department.
Heine <i>et al</i> 2011[72]	Randomized, double- blinded, placebo controlled trial comparing oral vitamin D to placebo. All subjects immunised after 9 weeks of supplementation and blood taken before and 7 days after immunisation.	Adults aged 26-34.5 years	2000IU of oral vitamin D3 oil per day or equal volume of neutral oil. All subjects received tetanus/diphtheria toxoid vaccine after 9 weeks of supplementation.	25-hydroxyvitamin D. TT specific IgG and IgA. Peripheral B and T lymphocytes. T cell activation to stimulation by Staph enterotoxin. TT specific cytokine profile (IL-2, IL-4, IL-5, IL-10, TNF- α and IFN γ). Adverse events. Leukocyte counts and IgG, IgA and IgM levels (supp. data)	T cell activation to stimulation by Staph enterotoxin and no antigen. Pre vaccination data not reported. Leukocyte counts and IgG, IgA and IgM levels (supp. data)	Medians in pg/ml with interquartile range concentrations	Not reported Monocyte count decreased in placebo group (p=0.04).	Randomized intervention is Vitamin D supplementation therefore no unimmunised controls.
Jorgensen <i>et al</i> 2013[73]	Sub-study within a larger randomised double blind controlled trial	Infants six weeks of age (n=480), either prior or after DTP vaccination	Randomised to Vitamin A supplementation or placebo	Cytokine production. Leukocyte counts.	TNF- α , IL-10, II-5, IL- 13 and IFN- γ responses in culture Lymphocytes, granulocytes, Eosinophils and monocytes	Geometric mean (pg/ml) Mean and SD	NS NS	
Lin <i>et al</i> 1997[74]	Randomized, controlled trial with three groups, including control. 15/80 subjects bled before and 1 month after vaccination.	Healthy medical personnel in Taiwan, aged 20-40 years, N=80.	Single dose of: Group 1: Td + full strength acellular pertussis vaccine Group 2: Td+half strength acellular pertussis vaccine Group 3: Td alone as	Adverse reactions. Anti- PT and anti-FHA antibodies. Lymphocyte proliferation to PT, FHA, Con-A, PHA and PWM. Cytokine production (IFN γ, IL-4, IL-5) to Con-A, PT and	Lymphocyte proliferation to Con-A, PHA and PWM. Cytokine production (IFN γ, IL-4, IL-5) to Con-A.	Stimulation index CPM, mean ±standard error. pg/ml, mean ±standard error.	NS	Adult only study. Designed to examine different pertussis vaccines but all received Td including controls.

			control	FHA.				
Rowe <i>et al</i> 2000[75]	Cohort study, all vaccinated, blood samples at 2, 3, 6 and 12 months.	Healthy infants in Australia, n=55	DTaP at 2, 4 and 6 months. Infants also received oral polio and Hib titre vaccines.	IL-4, IL-5, IL-6, IL-9, IL-10, IL13 and IFN γ stimulated by TT and PHA. IL-4 and IL-9 mRNA.	IL-4, IL-5, IL-6, IL-9, IL-10, IL13 and IFN γ stimulated by PHA.	Median pg/ml (test minus control cultures) with 10 th , 25 th , 75 th and 90 th percentiles. for positive responders	Increase in IL-5 (p=0.01) and IL- 13 (p=0.01) at 12 compared to 6 months. Increase in IL-5 (p=0.05) at 6 compared to 4 months.	
Yousfi <i>et al</i> 2005[76]	Case-control study comparing responses of elderly and young adults to mild inflammatory stress (i.e. vaccine). Blood taken 7 days before and 2 days after vaccination.	Cases: Elderly, male and female volunteers (mean age 70 \pm 4), N=7. Controls: Young, male and female volunteers (mean age 23 \pm 2).	Single dose DT-Polio and Typhim Vi vaccine.	Acute phase proteins (CRP, AGP, Fibrinogen, α1-Antitrypsin, Haptoglobulin, Albumin, Transthyretin, Transferrin). White blood cells counts. Plasma cytokine levels (TNFα, IL-6, IL-10). IL-6 6 and IL-10 production by LPS stimulated whole blood. IFN-γ production by PHA stimulated whole blood.	Acute phase proteins (CRP, AGP, Fibrinogen, α 1-Antitrypsin, Haptoglobulin, Albumin, Transthyretin, Transferrin). White blood cells counts. Plasma cytokine levels (TNF α , IL-6, IL-10). IL- 6 and IL-10 production by LPS stimulated whole blood. IFN- γ production by PHA stimulated whole blood.	Acute phase protein in mg or g/L, means ±standard error of mean. Cells/L, means ±standard error Cytokine levels ng/L or pg/ml, individual values with mean.	Increase from pre to post vaccine for CRP (p=0.003), AGP (p=0.0007), fibrinogen (p=0.004), haptoglobin (p=0.0023) and Transthyretin (p=0.01). Increase from pre to post vaccine in monocytes (p=0.007), lymphocytes (p- 0.002) and neutrophils (p=0.04) Eldery group plasma IL-6 and IL-10 increases from baseline (p<0.05). Both groups had increased IFN- γ production to PHA from baseline (p<0.05). Young adults had increased IL-6 production to LPS from baseline (p<0.05)	Adult only study. Designed to compare elderly with young adults. Vaccine used included Typhoid. Funding source not reported.
Zorzeto et al 2009[77]	Prospective, randomized, double- blinded, phase I comparative trial. Blood sample taken at 7 months of age (1 month after last vaccine).	Infants in Brazil, male and female, aged 2.1(±0.5 and 0.3)months N=247	Immunisation at 2, 4 and 6 months with either conventional whole cell pertussis vaccine or pertussis vaccine with low LPS content. Both vaccines also contained Diphtheria and tetanus toxoids.	T cell, CD3+ CD4+, CD8+ and $\gamma\delta$ + cell proliferation when stimulated by pertussis and PHA. Adverse events. IFN γ , TNF- α and IL-10 concentrations in response to pertussis and PHA. Anti-PT IgG titres. % protected against Diphtheria and tetanus.	T cell, CD3+ CD4+, CD8+ and $\gamma\delta$ + cell proliferation stimulated by PHA. IFN γ , TNF- α and IL-10 concentrations in response to PHA.	Median % of specific T cells with 95% CI. Medians of CD3+ blasts with individual points also plotted. Median pg/ml with individual points also plotted.	NS	

Table F. Characteristics of the included pertussis vaccine study

Author	Methods	Participants	Interventions	All Outcomes	Non-specific outcomes	Method of reporting non- specific outcomes	Difference in NSE outcome	Notes
Di Tommaso et	Follow-on of two	Healthy adults, male	Subjects had previously	PT neutralizing	PBMC proliferation	Stimulation index	Significance not	Authored by IRIS, the
al 1997[78]	phase 1 trials. Subjects	and female, 25-38	received two doses, six	antibodies. IgG	response to TT	for each participant	reported	Chiron-Vaccines
	bled either 6, 12, 18	years, who	weeks apart, of either a	antibodies to PT, FHA		at each time point		Immunobiology research
	and 24 months	previously	monocomponent	and 69K antigens.		for those who had		Institute
	(monocomponent	participated in one	pertussis vaccine (N=4)	Proliferation of PBMC		previously received		
	vaccine group) or 1,	of two studies, N=8.	or tricomponent	in response to PT and		monocomponent		
	2.5, 12, 18 and 54		pertussis vaccine (N=4).	TT, FHA and 69K.		vaccine only.		
	months (tricomponent							
	group) after second							
	vaccine.							

Table G. Risk of bias assessment of included BCG studies

Author		Random sequence generation (selection bias)	Allocation concealment (selection bias)	Blinding (performance bias and detection bias) All outcomes	Incomplete outcome data (attrition bias) All outcomes	Selective reporting (reporting bias)	Other bias
Akkoc et al 2010	Authors judgement	Low risk	Unclear risk	Unclear risk	Unclear risk	Unclear risk	
	Support for judgement	Full term healthy newborns from a single centre	No information on randomisation methods	Open trial	No information on management of missing data	No information on adjusting for multiple analyses	
Anderson et al 2013	Authors judgement	High risk	High risk	Unclear risk	Unclear risk	Unclear risk	Unclear risk
	Support for judgement	Subgroup of a larger trial with proportions of children taking part in subgroup trial varying by month on inclusion in the larger trial.	Randomized but no details given. Even with randomization, BCG group had significantly less hospitalisation and more often received micronutrients	Not blinded	Different children sampled at different timepoints	Multiple subgroup analyses	Part of the REVAC trial. During the study there was a national campaign to supplement with vitamin A and give missing vaccinations. Iron was also distributed to a subgroup in connection with a malaria trial.
Black et al 2001	Authors judgement	Low risk	Unclear risk	Unclear risk	Unclear risk	High risk	Unclear risk
	Support for judgement	Database was screened for eligible individuals	No details of randomisation method	No details of blinding process. Assume that participants were at least blinded to intervention	84% follow-up rate at 1 year, however variable numbers reported for each assay	Different numbers of subjects reported for each assay without explanation	Overall limited amount of information.
Black et al 2002	Authors judgement	Low risk	Low risk	Low risk	Low risk	Unclear risk	
	Support for judgement	Database was screened for eligible individuals in Malawi. In the UK recruitment was via a school vaccination program.	Randomised in blocks of six	Staff and subjects blinded to intervention	Probably a complete set of data	Those subjects who had positive skin tests were recruited in to the control groups	
Burl et al 2010	Authors judgement	Low risk	Low risk	Unclear risk	Low risk	Low risk	
	Support for judgement	Full term healthy newborns recruited at delivery from a single hospital.	Randomised in blocks of 20	Open trial	87 out of 103 completed 9 month time-point	Corrected for multiple analyses	
Burl et al (Aug) 2010	Authors judgement	Low risk	Low risk	Unclear risk	Low risk	Unclear risk	
	Support for judgement	Full term healthy newborns recruited at delivery from a single hospital.	Randomised in blocks of 20	Open trial	85/103 were followed-up at the final 20-28 month time- point	Statistical corrections made for multiple analyses Possibly more subjects excluded in group 2 due to tuberculosis exposure. i.e at 4.5 months Grp 1 n=51/53 Grp 2 n=39/50	
Burl et al 2009	Authors judgement	Low risk	Low risk	Unclear risk	Low risk	Low risk	
	Support for judgement	Full term healthy newborns recruited at delivery from a single hospital.	Randomised in blocks of 20	Open trial	87 out of 103 completed 9 month time-point	Corrected for multiple analyses	
Djuardi et al 2010	Authors judgement	High risk	High risk	Unclear risk	Unclear risk	Unclear	
	Support for judgement	Non-random. Part of a cohort to examine immune responses to helminths in children	Non-random	Open trial	98 out of 147 children were followed up at the 2 tear time-point	No information	
Elliot <i>et al</i> 2011	Authors judgement	High risk	High risk	Unclear risk	High risk	Low risk	High risk
(BCG and TT)	Support for judgement	Random sequence generation was for mother's treatment. Infants were not randomized to	Non-random	Open trial	1506 out of 2345 completed the BCG arm 1433 out of 2345 completed	Missing data described.	Mothers received various numbers of vaccinations during

		any intervention			the TT arm		pregnancy. There is no data on whether this may affect the infants subsequent response to vaccine
Faustman <i>et al</i> 2012	Authors judgement Support for judgement	Low risk Concealed allocation	Low risk Randomisation scheme prepared by the	Low risk Staff and subjects blinded to intervention	Low risk All subjects accounted for	Unclear risk A placebo treated subject developed primary EBV and was	
Fjallbrant et al 2007	Authors judgement Support for judgement	Unclear risk Subjects selected from 123 TST	hospital pharmacy High risk Non-random	High risk Non-blinded	Low risk No apparent attrition.	reported exclusively Unclear risk Non-specific data not reported	
	Support for Judgement	negative healthcare students based on matching previously vaccinated and not groups.	Non-random	Non-billided		Non-specific data not reported	
Gruber et al 2000	Authors judgement	Unclear risk	High risk	Unclear risk	High risk	Low risk	
	Support for judgement	Prospective cohort but 38% selective as high risk for atopic disease by family history	Vaccinated children had received BCG as they were at increased risk of TB and were more likely to be non- German	Unclear if blinded.	774/1314 included in this analysis as completed all scheduled examinations. Significant difference noted between these children and cohort as a whole in breastfeeding and vaccination rates.	Addresses aims, corrects for multiple analyses.	
Hoft et al 1998	Authors judgement	Unclear risk	Low risk	Low risk	Unclear risk	Unclear risk	
	Support for judgement	No information on recruitment	No information on randomisation method	Double blinded	53/54 participants completed the study. Some missing data not accounted for	Sub classified subjects in to responders and non-responders	
Hoft et al 1999	Authors judgement	Unclear risk	Unclear risk	Unclear risk	Unclear risk	Unclear risk	
	Support for judgement	No information on recruitment	No information on randomisation method	One arm was open label. Three arms were double blind	65/66 participants completed the study. Some missing data not accounted for	Most of the ELISA data is not reported.	
Hussey et al 2002	Authors judgement	High risk	High risk	Unclear risk	Unclear risk	Unclear risk	
÷	Support for judgement	Doesn't state specifically how subjects were identified	Non-random	Open trial	Data missing for some assays.	Didn't correct for multiple analyses.	
Kagina et al 2009)	Authors judgement	Low risk	Low risk	Unclear risk	Low risk	Unclear risk	
	Support for judgement	Approached mothers at antenatal clinic	Randomly allocated	Open trial	Relatively acceptable exclusions at each time-point documented	No information on a priori analysis	
Kleinnijenhuis et al	Authors judgement	High risk	High risk	Unclear risk	Low risk	Low risk	
2012	Support for judgement	Non-random	Non-random	Open trial	Small group size with no apparent attrition	No information on adjustment for multiple analyses	Small group size and no information on group subject composition
Lalor et al 2009	Authors judgement	High risk	High risk	Unclear risk	Unclear risk	Unclear risk	High risk
	Support for judgement	Recruitment in the UK through health centres. In Malawi recruitment took place at a single hospital	Non-random	Open-trial	No information on missing data	Sub-set of Malawi cohort who were vaccinated at the same time as the UK cohort were compared	Different age of vaccination in UK compared to Malawi
Lalor et al 2010	Authors judgement	High risk	Unclear risk	Unclear risk	Low risk	Unclear risk	Unclear risk
	Support for judgement	Subgroups of a larger study	Non-random	Not blinded	Single time-point an d all results appear present	Unstimulated control data not reported	Infants had previously taken part in an IFN gamma study
Lalor et al 2011	Authors judgement	High risk	High risk	Unclear risk	Unclear risk	Low risk	High risk
	Support for judgement	Subgroup of a larger study	Non-random	Open-trial	No information on missing	No information	Different age of

		where recruitment was in the UK		Γ	data		vaccination in UK
		through health centres. In			data		compared to Malawi
		Malawi recruitment took place at					compared to Malaw
		a single hospital					
Libraty et al 2014	Authors judgement	Unclear risk	Unclear risk	Unclear risk	Unclear risk	Low risk	
Liotady of an 2011	Support for judgement	Selected form a nested study due	Not random	Not blinded	No information on	All data appear to be reported	
	~~FF Jg	to specific attributes			management of missing data	· · · · · · · · · · · · · · · · · · ·	
Lowry et al 1998	Authors judgement	Unclear risk	Unclear risk	Unclear risk	Unclear risk	High risk	Unclear risk
•	Support for judgement	Doesn't state specifically how	Randomized but no	Unclear if blinded	Blood from 62/76 subjects	Non specific data not reported	15 subjects wer
		subjects were identified	details given		was tested for		revaccinated at 1
					lymphoproliferation but		weeks
					76/76 for serological studies		
Marchant et al 1999	Authors judgement	Low risk	Low risk	Unclear risk	High risk	Unclear risk	
	Support for judgement	Healthy newborns from a single	Randomised in blocks	Open trial	48 out of 137 were studied at	Adjusted for multiple analyses,	
		centre	of 6		the 1 year time-point	but no information on a priori	
N 1 10000		xx. 1 . 1	*** 1 * 1	*** 1 * 1	XX 1 1 1	analysis and ITT analysis	
Marks et al 2003	Authors judgement	High risk	High risk	High risk	Unclear risk	Low risk	
	Support for judgement	Mothers of potential participants identified by hospital records	Non-random	Retrospective open trial. No information regarding	Attempted to gather information from those who	A small proportion of the cytokine data was not included	
		Identified by hospital fecolds		masking of laboratory	didn't respond to the survey.	in analysis due to low responses	
				staff.	and the spond to the survey.	in controls.	
				Stuff.		Corrected for a wide variety of	
						confounders	
Miles et al 2008	Authors judgement	High risk	Unclear risk	Unclear risk	High risk	High risk	High risk
	Support for judgement	All samples from infants	Non-random	Not blinded	133 infants had blood taken at	NHDF results not reported	All infants were
		recruited into a larger CMV			some point but largest	1	infected with CMV
		study			number of samples at any one		
					time-point was 87.		
Miles et al 2010	Authors judgement	High risk	High risk	Unclear risk	Unclear risk	Unclear risk	High risk
	Support for judgement	Non-random	Non-random, no	Open trial	No information on	Infants that were diagnosed with	HIV positive
			unvaccinated controls		management of missing data	HIV were excluded. ELISpot	mothers and
						data classified into high and low	children born to
						responders	them all received
							anti-retroviral
							therapy
Ota et al 2002	Authors judgement	Low risk	Low risk	Unclear risk	Unclear risk	Unclear risk	
	Support for judgement	Healthy newborns from a single	Randomised in blocks	Open trial	85 out of 151 were followed-	Selective data for responses to	
	~~FF Jg	centre	of 6	• F · · · · · · ·	up at 4.5 month time-point	HBV reported	
Smith et al 2012	Authors judgement	High risk	Unclear risk	Unclear risk	Low risk	Low risk	
	Support for judgement	Doesn't state specifically how	Non-random	Open trial	Missing data accounted for.	Adjusted for multiple	
		subjects were identified		-	-	comparisons	
Soares et al 2013	Authors judgement	Low risk	Unclear risk	Unclear risk	Unclear risk	Low risk	
	Support for judgement	Recruitment antenatally, of	Not random	Not blinded	11/90 lost to follow-up and	Data appear to be complete	
		mothers attending maternity			6/90 exclusions		
a 1.1		units	** 1 1	· · ·			*** 1 * 1
Steenhuis et al 2008	Authors judgement	High risk	Unclear risk	Low risk	Low risk	Unclear risk	High risk
	Support for judgement	Newborns were recruited	Randomly allocated	Parents not blinded but	115/121 completed the study	Only 58 tested for eosinophils at	Unable to recrui
		antenatal from parents with	although no	principle researcher was.		18 months	required 200
		allergic disease	information on				participants
			randomisation method				therefore explorative
		1	1	1		1	analysis only.

							revaccinated at 4 months.
Tastan et al 2005	Authors judgement	Unclear risk	Unclear risk	Unclear risk	Low risk	Low risk	Unclear risk
	Support for judgement	Neonates were randomly chosen but not further details of selection process	Randomised but no details given	Not blinded	36/40 completed study	All results appear present	Funding source not reported
van den Biggelaar et al	Authors judgement	Unclear risk	Unclear risk	Unclear risk	Unclear risk	Unclear risk	
2009	Support for judgement	Very few details on participant selection. Significant differences at baseline between PNG and Australian newborn cohorts for caesarean delivery and head circumference	Not random	Not blinded	Some participants missing in outcome reporting with no explanation	Outcomes not clearly defined	
Vargas et al 2004	Authors judgement	Low risk	Unclear risk	Unclear risk	Low risk	Low risk	
	Support for judgement	Children recruited from several schools and diagnosis confirmed with pre-specified criteria.	Unclear if properly randomised (alternate allocation to group)	Unclear if blinded	67/82 completed study. Reasons for withdrawal given.	All results appear present	
Vekemans et al 2004	Authors judgement	Unclear risk	Low risk	Unclear risk	Unclear risk	Unclear risk	
	Support for judgement	No information on selection procedure	Randomlyallocatedalthoughnoinformationonrandomisationmethod	Open trial	No information on management of missing data	No information	
Vijaya Lakshmi V, et al	Authors judgement	High risk	High risk	High risk	Unclear risk	Unclear risk	High risk
2005	Support for judgement	Non-random	Non-random. Subjects possibly selected based on prior vaccination history	Open trial (retrospective)	No information on management of missing data	Subjects classified into responders and non-responders	Selection bias due to prior vaccine receipt
Weir et al 2004	Authors judgement	Low risk	Low risk	Low risk	Low risk	Unclear risk	
	Support for judgement	Database was screened for eligible individuals in Malawi. In the UK recruitment was via a school vaccination program.	Randomised in blocks of six	Staff and subjects blinded to intervention	Probably a complete set of data	Those subjects who had positive skin tests were recruited in to the control groups	
Weir et al 2008	Authors judgement	Low risk	High risk	Unclear risk	Unclear risk	Unclear risk	High risk
	Support for judgement	Recruitment was via a school vaccination program	Allocation dependent on response to skin testing	Open trial	No information on missing data	No information on adjustment for multiple analyses	Bias towards subjects with a Heaf grade of 2 or above being included into the control group.
Weir et al 2008	Authors judgement	Low risk	Low risk	Unclear risk	Unclear risk	Unclear risk	Unclear risk
	Support for judgement	School vaccination programs	Random allocation	Open label	Only subjects with data at each time-point were included	Not all antigens and controls reported for all cohorts	Study reports responses at different time-point by following up cohorts from three separate studies

Table H. Risk of bias assessment of included measles studies

Author		Random sequence generation (selection bias)	Allocation concealment (selection bias)	Blinding (performance bias and detection bias) All outcomes	Incomplete outcome data (attrition bias) All outcomes	Selective reporting (reporting bias)	Other bias
Bertley et al 2004	Authors judgement	Unclear risk	Unclear risk	Unclear risk	High risk	High risk	
	Support for judgement	Subjects were followed up from 6/14 of the original communities, chosen at random.	Details of previous randomisation not reported.	Not blinded	Only 37.8% of the original children were followed up at 5 years.	Results from the group who previously received single high dose EZ vaccine are not reported	
Gans et al 1999	Authors judgement	Unclear risk	Unclear risk	Unclear risk	Unclear risk	Unclear risk	High risk
	Support for judgement	Minimal details about subjects (ethinicity etc) and recruitment methods	Not randomized, adult control not matched	Not blinded	Post vaccine samples available for 134/162 infants but not all tested for all assays.	Non-specific effects not fully reported.	Unclear if previously vaccinated adults are used as controls. If so they are not suitable given age, unclear vaccination status etc.
Gans et al 2004	Authors judgement	Unclear risk	Unclear risk	Unclear risk	Unclear risk	High risk	
	Support for judgement	Minimal details about subjects (ethinicity etc).	Not randomized. Insufficient information to compare cases and controls.	Not blinded	Not all sample sizes allowed for full immunological evaluation but N is given for each assay.	PHA stimulated results not reported	
Hennino et al 2007	Authors judgement	Low risk	Low risk	Low risk	High risk	Unclear risk	High risk
	Support for judgement	All children had atopic dermatitis (AD) based on criteria of Hanifin and Rajka	Randomization file generated by computer	Double-blinded	5-6 infants out of 12 were tested for CCL18/E-selectin due to lack of serum and only 6 were followed up for 6 months for severity of AD scoring	Only 2 placebo subjects had CCL18 and E-selectin levels reported compared to 4 and 3 measles vaccine receipients	One infant seroconverted after placebo
Hussey et al 1996	Authors judgement	High risk	High risk	Unclear risk	Unclear risk	Unclear risk	High risk
	Support for judgement	Children recruited from two clinics in high and low measles prevalence areas but all children from low risk area received same vaccine and were all 9 months. No details of ethinicity.	Not randomized. Allocatted based on order of attendance. Children form low prevelance area all had same vaccine.	Not blinded	13/88 lost to follow up (2 of which excluded due to serological evidence of measles before vaccination)	Not always obvious from figures how many children have provided results.	All children aged 9 months were from a low prevalence area. Comparisons are made between groups which may not comparable.
Jaye et al 2014	Authors judgement	High risk	Unclear risk	Unclear risk	High risk	Unclear risk	
	Support for judgement	Very few demographic details	Not random	Not blinded	Initial number of recruits not stated	Not enough information	
Liguori et al 1998	Authors judgement	Unclear risk	Unclear risk	Unclear risk	High risk	Low risk	High risk
	Support for judgement	Insufficient details about recruitment and participants including previous vaccination history.	Not randomized	Not randomized	5/20 lost to follow-up- no details given	Individual results reported for each subject	Funding source not reported
Lisse et al 1994	Authors judgement	Unclear risk	Unclear risk	Unclear risk	High risk	Low risk	
	Support for judgement	Only those taking part in previous study were eligable. Only 330/854 from original study took part. Data for those not taking part not available except for those who died.	Not randomised	Not clear if those analysing samples were blinded to original group.	330/854 from original studies took part.	All results appear present.	Mortality data from this group is presented in Aaby 1993
Njie-Jobe et al 2012	Authors judgement	Unclear risk	Unclear risk	Unclear risk	Unclear risk	Unclear risk	Unclear risk
, ·····	Support for judgement	Subjects were recruited from a		Not blinded	91/132 completed study.	PHA results not reported.	Despite randomisation,

		larger CMV observational study.	not given		Similar attrition rates across groups		groups had significantly different median measles antibody titres at baseline
Okada et al 2001	Authors judgement	Unclear risk	Unclear risk	Unclear risk	Unclear risk	High risk	High risk
	Support for judgement	Insufficient demographic details (E.g ethinicity and socioeconomic status) to compare groups	Insufficient detail regarding vaccination of controls (eg when vaccinated and how many with each strain)	Not blinded	Each point on the figure represents average of 3-15 patients but unclear if numbers consistent	Control bloods were reportedly taken from 40 age-matched healthy children but no results are given.	NB for purposes of this SR, only the results for the vaccinated cohort have been included
Ovsyannikova et al	Authors judgement	Low risk	Unclear	Unclear risk	High risk	High risk	High risk
2003	Support for judgement	Subjects were randomly assigned to timing of blood draw	Not randomized, no control	Not blinded	Each subject had single blood draw therefore results at each time point are different children. Small numbers at each timepoint. Day 0 children (N=2 and 7) did not receive vaccine before blood draw.	As previous	One infant had positive measles antibody titres pre vaccination.
Pabst et al 1999	Authors judgement	Low risk	Unclear risk	Unclear risk	Unclear risk	Low risk	High risk
	Support for judgement	300 subjects enrolled over a 20 month period	Subjects in group 2 were apparently randomized to one of two vaccines but all those in group 1 received same vaccine, therefore not all groups comparable	Not clear if blinded	Samples not available for 2/300 at baseline and for 3/300 at 8 weeks. Not all assays performed on every sample- numbers given in paper.	All results appear present	Measles vaccine was given immediately after the third dose of DTP- Polio-Hib therefore any non-specific effects cannot be attributed to measles vaccine alone. Group 1 mothers were assumed to have had measles but this was not confirmed (e.g. through testing mothers for antibody).
Samb et al 1995	Authors judgement	Unclear risk	Unclear risk	Unclear risk	Unclear risk	Unclear risk	
	Support for judgement	Very few details on participant demographics	Randomly selected but no details of methods	Not blinded	Very few details on participants that were not included or missing data	all results appear present, however no evidence of an a priori analysis	
Schnorr et al 2001	Authors judgement	Unclear risk	High risk	Unclear risk	Unclear risk	High risk	
	Support for judgement	Recruited from larger measles trial	Not randomised. Controls more likely to have acute malnutrition and significantly less likely to have had 3 doses of DTP.	No information on blinding	Higher loss to follow up in control group	Non-significant data not shown	

Table I. Risk of bias assessment of included MMR vaccine studies

Nakayama et al 1990	Authors judgement	Unclear risk	Unclear risk	Unclear risk	Low risk	High risk	Unclear risk
	Support for judgement	Minimal details about subjects	Not randomized	Not blinded	Paired sera obtained for all	Non-specific effects not	Only 11/229 children
		(ethnicity, mean age etc.)			subjects (NB only 11 took	reported.	had blood taken for IFN
					part in IFN subset)		assays.
Pabst et al 1997	Authors judgement	Unclear risk	Unclear risk	Unclear risk	Unclear risk	Low risk	
	Support for judgement	Minimal details about subjects	Not randomized	Not blinded	Not all children assay at each	All results appear present	
		(ethnicity, mean age etc.)			time point. Not clear if all		
					children provided two		
					samples.		
Rager-Zisman et al	Authors judgement	Unclear risk	Unclear risk	Unclear risk	Unclear risk	Unclear risk	High risk
2003	Support for judgement	Healthy Israeli children but	Not randomized, no	Not blinded	Number of children tested	As previous.	13 children had received
		limited information about	control		varies by assay (N=28-38).		two doses of MMR
		subjects (ethnicity etc) and 13			Due is some cases to lack of		previously due to an
		had received 2 doses of MMR			serum but details not given.		epidemic.
		previously					

Table J. Risk of bias assessment of included tetanus toxoid vaccine studies

Author		Random sequence generation (selection bias)	Allocation concealment (selection bias)	Blinding (performance bias and detection bias) All outcomes	Incomplete outcome data (attrition bias) All outcomes	Selective reporting (reporting bias)	Other bias
Armitage et al 1993	Authors judgement	High risk	High risk	High risk	Unclear risk	Unclear risk	
	Support for judgement	Non-random	Non-random, different number of vaccination doses received depending on prior history	Open trial, staff participating in trial	No information	No information	
Borut et al 1980	Authors judgement	High risk	High risk	High risk	Unclear risk	Unclear risk	
	Support for judgement	Non-random	Non-random. Differing age ranges. Vaccination status determined on history	Open trial, staff participating in trial	No information	No information	
Chollet et al 1979	Authors judgement	High risk	High risk	Unclear risk	Unclear risk	Unclear risk	High risk
	Support for judgement	Non-random	Non-random	Open trial	No information	No information	General lack of information on subject demographics and study design
	Authors judgement	High risk	High risk	Unclear risk	Unclear risk	Unclear risk	High risk
Chui et al 2004	Support for judgement	Non-random	Non-random	Open trial	Missing information for 2 participants	No information	Small cohort sizes
Cooper et al 1998	Authors judgement	High risk	High risk	Unclear risk	Unclear risk	Unclear risk	
	Support for judgement	Non-random	Non-random. Groups defined on presence of <i>Onchocera volvulus</i> infection	Open trial	No information	No information	
Di Genova et al 2006	Authors judgement	High risk	High risk	Unclear risk	Unclear risk	High risk	
(Also BCG)	Support for judgement	Non-random	Non-random, cohort	Open trial	No information	Results selectively reported for maximal response time- point	
Fernandez et al 1994	Authors judgement	High risk	High risk	Unclear risk	Unclear risk	Unclear risk	High risk
	Support for judgement	Non-random	Non-random	Open trial	No information, although missing data unlikely given only 3 subjects in study	No information	Only 3 subjects
Fevrier et al 1977	Authors judgement	High risk	High risk	Unclear risk	Unclear risk	High risk	High risk
	Support for judgement	Non-random	Non-random, cohort with very little demographic details reported	Open trial	No information	Selective reporting/conduction of experiments in specific subjects	Small cohort with experiments performed on selected individuals
Gentile et al 2006	Authors judgement	High risk	High risk	Unclear risk	Low risk	Unclear risk	Unclear risk
	Support for judgement	Non-random	Non-random	Open trial	All subjects completed the trial	Allergy skin testing not reported. Primary outcome defined	Exclusion of those who were on treatment for allergic rhinitis
Livingston et al 2013	Authors judgement	High risk	High risk	Unclear risk	Low risk	Unclear risk	
	Support for judgement	Non-random	Non-random	Open trial	All participants completed all visits	No information	
Mahalingham et al 2010	Authors judgement	Unclear risk	Unclear risk	Unclear risk	Unclear risk	Unclear risk	
	Support for judgement	Only females selected	No information on randomisation procedure	Double blinded to receipt of palm oil	No information	No information	

Author		Random sequence generation (selection bias)	Allocation concealment (selection bias)	Blinding (performance bias and detection bias) All outcomes	Incomplete outcome data (attrition bias) All outcomes	Selective reporting (reporting bias)	Other bias
Dirix et al 2009	Authors judgement	High risk	Unclear risk	Unclear risk	Unclear risk	Unclear risk	Unclear risk
	Support for judgement	Subjects were selected by having HIV positive mothers but being HIV negative. All had received Zidovudine for 6 weeks until HIV status confirmed. All had been included in	Not randomised	Not blinded	Not all infants were bled at all time points and not all assays could be performed due to limited blood volumes.	Some assays only report results from 11 subjects.	All children had been involved in previous studies on cellular immune responses to pertussis vaccine. All had received 6
		previous studies.					weeks of Zidovudine prophylaxis until HIV status was confirmed.
							Part funded by Sanofi Pasteur.
Fernandes et al 2010	Authors judgement	Unclear risk	Unclear risk	Unclear risk	Low risk	Unclear risk	Unclear risk
	Support for judgement	Male only subjects.	Not randomised, no unvaccinated controls.	Not blinded	All completed protocol	Frequency of B lymphocyte subjects is from one representative subject.	Funding source not reported.
Fryauff et al 1998	Authors judgement	High risk	High risk	Unclear risk	Unclear risk	Unclear risk	High risk
	Support for judgement	Subjects were recruited from a pre-existing malaria prophylaxis trial. All male.	Not randomised, controls differed as not receiving anti-malarials.	Not blinded	Attrition rate not reported. Some assays not done (table 1)	PHA and PPD data not shown.	Subset of other (anti- malarial) trial. Unclear if control group was part of this trial or not.
Halasa et al 2008	Authors judgement	Low risk	Unclear risk	Unclear risk	Low risk	Low risk	Unclear risk
	Support for judgement	270 parents approached to get sample size of 50.	Randomised but no details given.	Parents blinded, but unclear if study staff were.	42/50 completed study, reasons for withdrawal given.	All results appear present	Part funded by Sanofi- Pasteur
He et al 1998	Authors judgement	Unclear risk	Unclear risk	Unclear risk	Unclear risk	High risk	High risk
	Support for judgement	Children randomly selected. Adult vaccinees and controls were convenience sample from department.	Not randomised. Natural infection cases had been symptomatic for a wide range of time.	Not blinded	PHA and pokeweed responses not shown.	Results reported for certain patients only which varies by assay	Controls not well matched for group as a whole.
Heine et al 2011	Authors judgement	High risk	Unclear risk	Low risk	Unclear risk	High risk	High risk
	Support for judgement	Recruitment from department of Dermatology and Allergy but no details of any medical conditions reported.	Random assignment to Vit D treatment group but no details of process given	Double-blinded (NB blinded intervention is Vitamin D supplementation)	Attrition rates not reported	Subjects with evidence of spontaneous cytokine release were excluded due to assumed subclinical concomitant infection.	Randomized intervention is Vitamin D supplementation therefore no unimmunised controls.
Jorgensen et al 2013	Authors judgement	Unclear risk	Low risk	Low risk	Unclear risk	Unclear risk	Unclear risk
	Support for judgement	Subjects recruited across six districts. No differences in demographics. Rules for inclusion modified mid-study	Randomised using a code held by the pharmacist.	Double blinded	No information on how many/or if all of those recruited into the main study were approached for this sub- study	Recruitment protocol modified significantly with respect to DTP outcome reported	DTP vaccinated cohort much smaller than unvaccinated cohort
Lin et al 1997	Authors judgement	Unclear risk	Unclear risk	Unclear risk	High risk	Unclear risk	
	Support for judgement	All medical personnel from one Children's hospital in Taiwan.	No details of randomisation process given	Unclear if blinded	Only 15/80 were bled before and after vaccination. However all 80 provide adverse reaction data.	Unclear how many subjects are included in each table.	
Rowe et al 2000	Authors judgement	Unclear risk	Unclear risk	Unclear risk	Low risk	High risk	Unclear risk
		1					

Supplementary Table 20. Risk of bias assessment of included DTP and DT vaccine studies

	Support for judgement	Recruitment details not given	Not randomized	Not blinded	Samples were obtained from $\geq 78\%$ of the group at eah	Results are reported for positive responders only	Funding source not reported
					time point	positive responders only	reported
Yousfi et al 2005	Authors judgement	Unclear risk	Unclear risk	Unclear risk	Unclear risk	Low risk	Unclear risk
	Support for judgement	Recruitment details not given.	Not randomised, no unvaccinated controls	Not blinded	Attrition rates not reported however some subjects appear missing.	**	Funding source not reported
Zorzeto et al 2009	Authors judgement	Low risk	Unclear risk	Low risk	Unclear risk	High risk	
	Support for judgement		Randomized to receive one of two vaccines but no details of randomization process.	Double-blinded	234/237 completed the study. Results reported for varying number of subjects depending on assay (reported in figures/tables)		

Table K. Risk of bias assessment of the included pertussis vaccine study.

Author		Random sequence generation (selection bias)	Allocation concealment (selection bias)	Blinding (performance bias and detection bias) All outcomes	Incomplete outcome data (attrition bias) All outcomes	Selective reporting (reporting bias)	Other bias
Di Tommaso et al 1997	Authors judgement	Unclear risk	Unclear risk	Unclear risk	Low risk	Low risk	Unclear risk
	Support for judgement	Subjects recruited from those taking part in previous trials, unclear how many subjects there were in total in these previous trials.	Not randomised	Not blinded	All data appears to have been reported for each subject	All data appears to have been reported for each subject	All authors from company producing vaccine

Reference list of all included studies

- 1. Akkoc T, Aydogan M, Yildiz A, et al. Neonatal BCG vaccination induces IL-10 production by CD4+ CD25+ T cells. Pediatric allergy and immunology : official publication of the European Society of Pediatric Allergy and Immunology 2010;**21**(7):1059-63 doi: 10.1111/j.1399-3038.2010.01051.x[published Online First: Epub Date]l.
- Andersen A, Roth A, Jensen KJ, et al. The immunological effect of revaccination with Bacille Calmette-Guerin vaccine at 19 months of age. Vaccine 2013;31(17):2137-44 doi: 10.1016/j.vaccine.2013.02.050[published Online First: Epub Date]|.
- 3. Black GF, Dockrell HM, Crampin AC, et al. Patterns and implications of naturally acquired immune responses to environmental and tuberculous mycobacterial antigens in northern Malawi. The Journal of infectious diseases 2001;**184**(3):322-9 doi: 10.1086/322042[published Online First: Epub Date].
- 4. Black GF, Weir RE, Floyd S, et al. BCG-induced increase in interferon-gamma response to mycobacterial antigens and efficacy of BCG vaccination in Malawi and the UK: two randomised controlled studies. Lancet 2002;**359**(9315):1393-401 doi: 10.1016/S0140-6736(02)08353-8[published Online First: Epub Date]].
- Burl S, Adetifa UJ, Cox M, et al. Delaying bacillus Calmette-Guerin vaccination from birth to 4 1/2 months of age reduces postvaccination Th1 and IL-17 responses but leads to comparable mycobacterial responses at 9 months of age. Journal of immunology 2010;185(4):2620-8 doi: 10.4049/jimmunol.1000552[published Online First: Epub Date]l.
- 6. Burl S, Adetifa UJ, Cox M, et al. The tuberculin skin test (TST) is affected by recent BCG vaccination but not by exposure to non-tuberculosis mycobacteria (NTM) during early life. PloS one 2010;5(8):e12287 doi: 10.1371/journal.pone.0012287[published Online First: Epub Date]l.
- 7. Burl S. The Role of Regulatory T cells in Early Life Immunity to BCG: Influence of Exposure to Environmental Mycobacteria. Open University, 2009.
- Djuardi Y, Sartono E, Wibowo H, et al. A longitudinal study of BCG vaccination in early childhood: the development of innate and adaptive immune responses. PloS one 2010;5(11):e14066 doi: 10.1371/journal.pone.0014066[published Online First: Epub Date]l.
- Elliott AM, Mawa PA, Webb EL, et al. Effects of maternal and infant co-infections, and of maternal immunisation, on the infant response to BCG and tetanus immunisation. Vaccine 2010;29(2):247-55 doi: 10.1016/j.vaccine.2010.10.047[published Online First: Epub Date]l.
- Faustman DL, Wang L, Okubo Y, et al. Proof-of-concept, randomized, controlled clinical trial of Bacillus-Calmette-Guerin for treatment of long-term type 1 diabetes. PloS one 2012;7(8):e41756 doi: 10.1371/journal.pone.0041756[published Online First: Epub Date]l.
- Fjallbrant H, Ridell M, Larsson LO. Primary vaccination and revaccination of young adults with BCG: a study using immunological markers. Scandinavian journal of infectious diseases 2007;**39**(9):792-8 doi: 10.1080/00365540701367777[published Online First: Epub Date]l.
- 12. Gruber C, Kulig M, Bergmann R, et al. Delayed hypersensitivity to tuberculin, total immunoglobulin E, specific sensitization, and atopic manifestation in longitudinally followed early Bacille Calmette-Guerin-vaccinated and nonvaccinated children. Pediatrics 2001;**107**(3):E36
- Hoft DF, Brown RM, Roodman ST. Bacille Calmette-Guerin vaccination enhances human gamma delta T cell responsiveness to mycobacteria suggestive of a memory-like phenotype. Journal of immunology 1998;161(2):1045-54
- 14. Hoft DF, Kemp EB, Marinaro M, et al. A double-blind, placebo-controlled study of Mycobacterium-specific human immune responses induced by intradermal bacille Calmette-Guerin vaccination. The Journal of laboratory and clinical medicine 1999;**134**(3):244-52
- 15. Hussey GD, Watkins ML, Goddard EA, et al. Neonatal mycobacterial specific cytotoxic T-lymphocyte and cytokine profiles in response to distinct BCG vaccination strategies. Immunology 2002;**105**(3):314-24
- 16. Kagina BM, Abel B, Bowmaker M, et al. Delaying BCG vaccination from birth to 10 weeks of age may result in an enhanced memory CD4 T cell response. Vaccine 2009;27(40):5488-95 doi: 10.1016/j.vaccine.2009.06.103[published Online First: Epub Date]l.
- Kleinnijenhuis J, Quintin J, Preijers F, et al. Bacille Calmette-Guerin induces NOD2-dependent nonspecific protection from reinfection via epigenetic reprogramming of monocytes. Proceedings of the National Academy of Sciences of the United States of America 2012;109(43):17537-42 doi: 10.1073/pnas.1202870109[published Online First: Epub Date]l.
- 18. Lalor MK, Ben-Smith A, Gorak-Stolinska P, et al. Population differences in immune responses to Bacille Calmette-Guerin vaccination in infancy. The Journal of infectious diseases 2009;**199**(6):795-800
- 19. Lalor MK, Smith SG, Floyd S, et al. Complex cytokine profiles induced by BCG vaccination in UK infants. Vaccine 2010;**28**(6):1635-41 doi: 10.1016/j.vaccine.2009.11.004[published Online First: Epub Date]l.
- Lalor MK, Floyd S, Gorak-Stolinska P, et al. BCG vaccination induces different cytokine profiles following infant BCG vaccination in the UK and Malawi. The Journal of infectious diseases 2011;204(7):1075-85 doi: 10.1093/infdis/jir515[published Online First: Epub Date]l.

- 21. Libraty DH, Zhang L, Woda M, et al. Neonatal BCG vaccination is associated with enhanced T-helper 1 immune responses to heterologous infant vaccines Trials in Vaccinology 2014;**3**:1-5
- 22. Lowry PW, Ludwig TS, Adams JA, et al. Cellular immune responses to four doses of percutaneous bacille Calmette-Guerin in healthy adults. The Journal of infectious diseases 1998;**178**(1):138-46
- 23. Marchant A, Goetghebuer T, Ota MO, et al. Newborns develop a Th1-type immune response to Mycobacterium bovis bacillus Calmette-Guerin vaccination. Journal of immunology 1999;**163**(4):2249-55
- 24. Marks GB, Ng K, Zhou J, et al. The effect of neonatal BCG vaccination on atopy and asthma at age 7 to 14 years: an historical cohort study in a community with a very low prevalence of tuberculosis infection and a high prevalence of atopic disease. The Journal of allergy and clinical immunology 2003;**111**(3):541-9
- 25. Miles DJ, van der Sande M, Crozier S, et al. Effects of antenatal and postnatal environments on CD4 T-cell responses to Mycobacterium bovis BCG in healthy infants in the Gambia. Clinical and vaccine immunology : CVI 2008;**15**(6):995-1002 doi: 10.1128/CVI.00037-08[published Online First: Epub Date]l.
- 26. Miles DJ, Gadama L, Gumbi A, et al. Human immunodeficiency virus (HIV) infection during pregnancy induces CD4 T-cell differentiation and modulates responses to Bacille Calmette-Guerin (BCG) vaccine in HIV-uninfected infants. Immunology 2010;**129**(3):446-54 doi: 10.1111/j.1365-2567.2009.03186.x[published Online First: Epub Date]l.
- Ota MO, Vekemans J, Schlegel-Haueter SE, et al. Influence of Mycobacterium bovis bacillus Calmette-Guerin on antibody and cytokine responses to human neonatal vaccination. Journal of immunology 2002;168(2):919-25
- Smith SG, Lecher S, Blitz R, et al. Broad heparin-binding haemagglutinin-specific cytokine and chemokine response in infants following Mycobacterium bovis BCG vaccination. European journal of immunology 2012;42(9):2511-22 doi: 10.1002/eji.201142297[published Online First: Epub Date].
- Soares AP, Kwong Chung CK, Choice T, et al. Longitudinal changes in CD4(+) T-cell memory responses induced by BCG vaccination of newborns. The Journal of infectious diseases 2013;207(7):1084-94 doi: 10.1093/infdis/jis941[published Online First: Epub Date]l.
- 30. Steenhuis TJ, van Aalderen WM, Bloksma N, et al. Bacille-Calmette-Guerin vaccination and the development of allergic disease in children: a randomized, prospective, single-blind study. Clinical and experimental allergy : journal of the British Society for Allergy and Clinical Immunology 2008;38(1):79-85 doi: 10.1111/j.1365-2222.2007.02859.x[published Online First: Epub Date]l.
- 31. Tastan Y, Arvas A, Demir G, et al. Influence of Bacillus Calmette-Guerin vaccination at birth and 2 months old age on the peripheral blood T-cell subpopulations [gamma/delta and alpha-beta T cell]. Pediatric allergy and immunology : official publication of the European Society of Pediatric Allergy and Immunology 2005;**16**(8):624-9 doi: 10.1111/j.1399-3038.2005.00329.x[published Online First: Epub Date]l.
- 32. van den Biggelaar AH, Prescott SL, Roponen M, et al. Neonatal innate cytokine responses to BCG controlling T-cell development vary between populations. The Journal of allergy and clinical immunology 2009;**124**(3):544-50, 50 e1-2 doi: 10.1016/j.jaci.2009.03.040[published Online First: Epub Date]l.
- 33. Vargas MH, Bernal-Alcantara DA, Vaca MA, et al. Effect of BCG vaccination in asthmatic schoolchildren. Pediatric allergy and immunology : official publication of the European Society of Pediatric Allergy and Immunology 2004;**15**(5):415-20 doi: 10.1111/j.1399-3038.2004.00198.x[published Online First: Epub Date]l.
- 34. Vekemans J, Ota MO, Sillah J, et al. Immune responses to mycobacterial antigens in the Gambian population: implications for vaccines and immunodiagnostic test design. Infection and immunity 2004;72(1):381-8
- 35. Vijaya Lakshmi V, Kumar S, Surekha Rani H, et al. Tuberculin specific T cell responses in BCG vaccinated children. Indian pediatrics 2005;**42**(1):36-40
- 36. Weir RE, Black GF, Dockrell HM, et al. Mycobacterial purified protein derivatives stimulate innate immunity: Malawians show enhanced tumor necrosis factor alpha, interleukin-1beta (IL-1beta), and IL-10 responses compared to those of adolescents in the United Kingdom. Infection and immunity 2004;72(3):1807-11
- 37. Weir RE, Fine PE, Floyd S, et al. Comparison of IFN-gamma responses to mycobacterial antigens as markers of response to BCG vaccination. Tuberculosis 2008;**88**(1):31-8
- Weir RE, Gorak-Stolinska P, Floyd S, et al. Persistence of the immune response induced by BCG vaccination. BMC infectious diseases 2008;8:9 doi: 10.1186/1471-2334-8-9[published Online First: Epub Date]l.
- Bertley FM, Ibrahim SA, Libman M, et al. Measles vaccination in the presence of maternal antibodies primes for a balanced humoral and cellular response to revaccination. Vaccine 2004;23(4):444-9 doi: 10.1016/j.vaccine.2004.06.021[published Online First: Epub Date]l.
- 40. Gans HA, Maldonado Y, Yasukawa LL, et al. IL-12, IFN-gamma, and T cell proliferation to measles in immunized infants. Journal of immunology 1999;162(9):5569-75

- 41. Gans HA, Yasukawa LL, Alderson A, et al. Humoral and cell-mediated immune responses to an early 2dose measles vaccination regimen in the United States. The Journal of infectious diseases 2004;**190**(1):83-90 doi: 10.1086/421032[published Online First: Epub Date]l.
- 42. Hennino A, Cornu C, Rozieres A, et al. Influence of measles vaccination on the progression of atopic dermatitis in infants. Pediatric allergy and immunology : official publication of the European Society of Pediatric Allergy and Immunology 2007;**18**(5):385-90 doi: 10.1111/j.1399-3038.2007.00537.x[published Online First: Epub Date]l.
- 43. Hussey GD, Goddard EA, Hughes J, et al. The effect of Edmonston-Zagreb and Schwarz measles vaccines on immune response in infants. The Journal of infectious diseases 1996;**173**(6):1320-6
- 44. Jaye A, Magnusen AF, Sadiq AD, et al. Ex vivo analysis of cytotoxic T lymphocytes to measles antigens during infection and after vaccination in Gambian children. The Journal of clinical investigation 1998;**102**(11):1969-77 doi: 10.1172/JCI3290[published Online First: Epub Date]l.
- 45. Liguori G, Tolone C, D'Avanzo M, et al. Modifications of IFN-alpha, TNF-alpha and IFN-gamma serum levels in children vaccinated against measles. Annali di igiene : medicina preventiva e di comunita 1998;**10**(5-6):303-6
- 46. Lisse IM, Aaby P, Knudsen K, et al. Long term impact of high titer Edmonston-Zagreb measles vaccine on T lymphocyte subsets. The Pediatric infectious disease journal 1994;**13**(2):109-12
- 47. Njie-Jobe J, Nyamweya S, Miles DJ, et al. Immunological impact of an additional early measles vaccine in Gambian children: responses to a boost at 3 years. Vaccine 2012;30(15):2543-50 doi: 10.1016/j.vaccine.2012.01.083[published Online First: Epub Date]l.
- 48. Okada H, Sato TA, Katayama A, et al. Comparative analysis of host responses related to immunosuppression between measles patients and vaccine recipients with live attenuated measles vaccines. Archives of virology 2001;146(5):859-74
- 49. Ovsyannikova IG, Reid KC, Jacobson RM, et al. Cytokine production patterns and antibody response to measles vaccine. Vaccine 2003;**21**(25-26):3946-53
- 50. Pabst HF, Spady DW, Carson MM, et al. Cell-mediated and antibody immune responses to AIK-C and Connaught monovalent measles vaccine given to 6 month old infants. Vaccine 1999;**17**(15-16):1910-8
- 51. Samb B, Whittle H, Aaby P, et al. No evidence of long-term immunosuppression after high-titer Edmonstron-Zagreb measles vaccination in Senegal. The Journal of infectious diseases 1995;**171**(2):506-8
- 52. Schnorr JJ, Cutts FT, Wheeler JG, et al. Immune modulation after measles vaccination of 6-9 months old Bangladeshi infants. Vaccine 2001;**19**(11-12):1503-10
- 53. Nakayama T, Urano T, Osano M, et al. Evaluation of live trivalent vaccine of measles AIK-C strain, mumps Hoshino strain and rubella Takahashi strain, by virus-specific interferon-gamma production and antibody response. Microbiology and immunology 1990;**34**(6):497-508
- 54. Pabst HF, Spady DW, Carson MM, et al. Kinetics of immunologic responses after primary MMR vaccination. Vaccine 1997;**15**(1):10-4
- 55. Rager-Zisman B, Bazarsky E, Skibin A, et al. The effect of measles-mumps-rubella (MMR) immunization on the immune responses of previously immunized primary school children. Vaccine 2003;**21**(19-20):2580-8
- 56. Armitage KB, Duffy EG, Mincek MA, et al. Transient normalization of lymphocyte blastogenic and specific antibody responses following boosting of healthy elderly subjects with tetanus toxoid. Journal of gerontology 1993;48(1):M19-25
- 57. Borut TC, Ank BJ, Gard SE, et al. Tetanus toxoid skin test in children: correlation with in vitro lymphocyte stimulation and monocyte chemotaxis. The Journal of pediatrics 1980;**97**(4):567-73
- 58. Chollet P, Chassagne J, Philippe P, et al. Peripheral lymphocytes changes in the anamnestic response to tetanus toxoid challenge. Clinical and experimental immunology 1979;**37**(1):152-61
- 59. Stephen Chui MTMC, PhD† Paul J. Mosca, MD, PhD† Amy C. Hobeika† Takuya Osada, MD† Laurent Galibert, PhD‡ Dania Caron, MD‡, H. Kim Lyerly MMAM, MD*. Flt3-ligand as a Vaccine Adjuvant: Results in a Study of Flt3-ligand Plus Tetanus Toxoid Immunization. The Journal of Applied Research 2004;4(4):536-49
- 60. Cooper PJ, Espinel I, Paredes W, et al. Impaired tetanus-specific cellular and humoral responses following tetanus vaccination in human onchocerciasis: a possible role for interleukin-10. The Journal of infectious diseases 1998;**178**(4):1133-8
- 61. Di Genova G, Roddick J, McNicholl F, et al. Vaccination of human subjects expands both specific and bystander memory T cells but antibody production remains vaccine specific. Blood 2006;**107**(7):2806-13 doi: 10.1182/blood-2005-08-3255[published Online First: Epub Date]l.
- Fernandez V, Andersson J, Andersson U, et al. Cytokine synthesis analyzed at the single-cell level before and after revaccination with tetanus toxoid. European journal of immunology 1994;24(8):1808-15 doi: 10.1002/eji.1830240813[published Online First: Epub Date]l.
- 63. Fevrier M, Bona C, Eyquem A, et al. Inhibition of mixed lymphocyte reactions in humans after immunization with tetanus toxoid. Transplantation 1977;**23**(3):199-209

- 64. Gentile D, Trecki J, Patel A, et al. Effect of tetanus immunization on t-helper cytokine production in adults with and without allergic rhinitis. Allergy and asthma proceedings : the official journal of regional and state allergy societies 2006;27(3):197-201
- 65. Livingston KA, Jiang X, Stephensen CB. CD4 T-helper cell cytokine phenotypes and antibody response following tetanus toxoid booster immunization. Journal of immunological methods 2013;**390**(1-2):18-29 doi: 10.1016/j.jim.2013.01.001[published Online First: Epub Date]l.
- 66. Mahalingam D, Radhakrishnan AK, Amom Z, et al. Effects of supplementation with tocotrienol-rich fraction on immune response to tetanus toxoid immunization in normal healthy volunteers. European journal of clinical nutrition 2011;65(1):63-9 doi: 10.1038/ejcn.2010.184[published Online First: Epub Date]I.67. Dirix V, Verscheure V, Goetghebuer T, et al. Monocyte-derived interleukin-10 depresses the Bordetella pertussis- specific gamma interferon response in vaccinated infants. Clinical and vaccine immunology : CVI 2009;16(12):1816-21 doi: 10.1128/CVI.00314-09[published Online First: Epub Date]I.
- 68. Fernandes JR, Wasserman S, Snider DP. Stimulation of anti-polio and anti-HSV IgA pre-plasma cell response in blood following parenteral immunization with tetanus-diphtheria vaccine. Vaccine 2010;28(6):1493-8 doi: 10.1016/j.vaccine.2009.11.057[published Online First: Epub Date]].
- 69. Fryauff DJ, Mouzin E, Church LW, et al. Lymphocyte response to tetanus toxin T-cell epitopes: effects of tetanus vaccination and concurrent malaria prophylaxis. Vaccine 1999;**17**(1):59-63
- 70. Halasa NB, O'Shea A, Shi JR, et al. Poor immune responses to a birth dose of diphtheria, tetanus, and acellular pertussis vaccine. The Journal of pediatrics 2008;153(3):327-32 doi: 10.1016/j.jpeds.2008.03.011[published Online First: Epub Date]|.
- 71. He Q, Tran Minh NN, Edelman K, et al. Cytokine mRNA expression and proliferative responses induced by pertussis toxin, filamentous hemagglutinin, and pertactin of Bordetella pertussis in the peripheral blood mononuclear cells of infected and immunized schoolchildren and adults. Infection and immunity 1998;66(8):3796-801
- Heine G, Drozdenko G, Lahl A, et al. Efficient tetanus toxoid immunization on vitamin D supplementation. European journal of clinical nutrition 2011;65(3):329-34 doi: 10.1038/ejcn.2010.276[published Online First: Epub Date]l.
- 73. Jorgensen MJ, Fisker AB, Sartono E, et al. The effect of at-birth vitamin A supplementation on differential leucocyte counts and in vitro cytokine production: an immunological study nested within a randomised trial in Guinea-Bissau. The British journal of nutrition 2012:1-11 doi: 10.1017/S0007114512001304[published Online First: Epub Date]l.
- 74. Lin TY, Chiang BL. Specific immune response in adult medical personnel immunized with acellular pertussis vaccine with special emphasis on T helper cell response. Vaccine 1997;**15**(17-18):1917-21
- 75. Rowe J, Macaubas C, Monger TM, et al. Antigen-specific responses to diphtheria-tetanus-acellular pertussis vaccine in human infants are initially Th2 polarized. Infection and immunity 2000;**68**(7):3873-7
- 76. El Yousfi M, Mercier S, Breuille D, et al. The inflammatory response to vaccination is altered in the elderly. Mechanisms of ageing and development 2005;126(8):874-81 doi: 10.1016/j.mad.2005.03.008[published Online First: Epub Date]l.
- 77. Zorzeto TQ, Higashi HG, da Silva MT, et al. Immunogenicity of a whole-cell pertussis vaccine with low lipopolysaccharide content in infants. Clinical and vaccine immunology : CVI 2009;16(4):544-50 doi: 10.1128/CVI.00339-08[published Online First: Epub Date]l.
- 78. Di Tommaso A, Bartalini M, Peppoloni S, et al. Acellular pertussis vaccines containing genetically detoxified pertussis toxin induce long-lasting humoral and cellular responses in adults. Vaccine 1997;15(11):1218-24