

Figure S1 Rapid identification of heterozygous and homozygous mutant lines through pollen fluorescence. (A) *Arabidopsis thaliana* mature pollen grains of wild type. (B) The *qrt1* mature pollen grains maintain male meiotic products in tetrads. (C and D) Mature pollen grains of a heterozygous T-DNA insertion mutant. Two of four pollen grains from the same meiotic product are fluorescent; (E and F) Mature pollen grains of a homozygous T-DNA insertion mutant plant screened in T2 generation. The four pollen grains from the same meiotic product are fluorescent. Scale bar = 20 μ m.

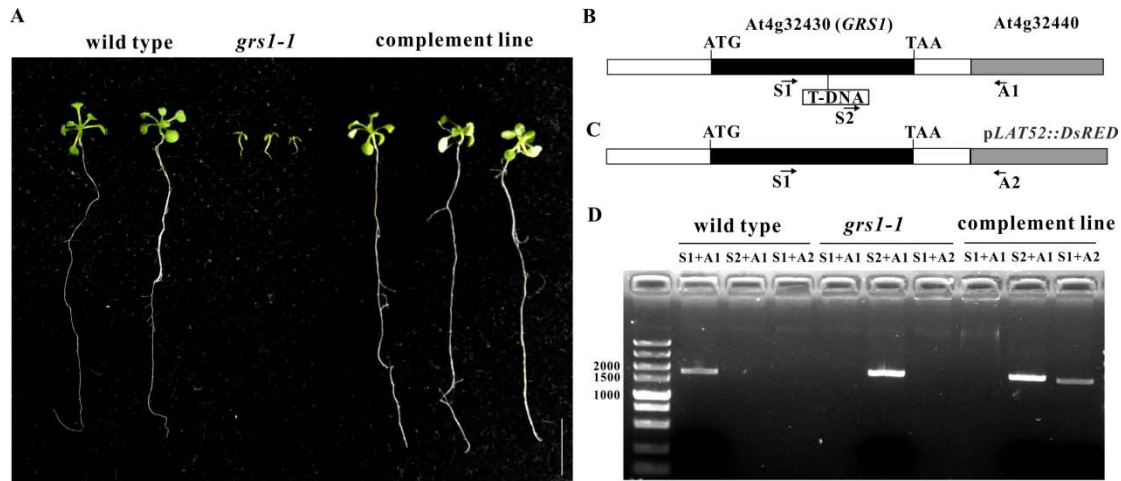


Figure S2 Genomic complementary fragment of At4g32430 rescues the phenotype of *grs1-1*.

A 11-day-old seedlings of wild type, *grs1-1* and the complemented line. Compared to wild type seedlings, *grs1-1* seedlings showed an extremely slow growth phenotype. Size of seedlings of the complemented line (homozygous for *grs1-1*, heterozygous for the genomic fragment) is similar to wild type seedlings. Scale bar = 1 cm.

B Diagram of the *GRS1* (At4g32430) genomic DNA and the insertion sites for the T-DNA in *grs1-1*. The location of primers S1, S2 and A1 used for genotyping are indicated by arrows.

C Diagram of the *GRS1* genomic complementary fragment and additional T-DNA element. The location of primers S1 and A2 used for genotyping are indicated by arrows.

D Genotypes of wild type, *grs1-1* and complemented line. Wild type shows a product of about 1700 bps (S1+A1). Homozygous *grs1-1* plants show a product of about 1800 bps (S2+A1). The complemented line shows two products of about 1800 bps (S2+A1) and 1700 bps (S1+A2).

motifs		L	S	P	L	S	P	L	S	P	L	S	P	L	S													
Residue no. within	2nd	D	C	V	F	V	I	V	E	V	V	S	I	Y	V	F	V	E										
	5th	T	N	N	S	N	N	T	N	T	V	N	N	G	S	T	L	S										
	35th	F	D	D	D	D	D	L	N	N	E	E	N	C	N	D	S	G										
			-20			-15			-10			-5					+5											
<i>nad1-265</i>	A	U	U	U	A	U	G	U	U	A	A	G	U	C	U	G	G	U	C	G	C	U	C	G	G	G	C	C
<i>nad4L-55</i>	G	A	U	U	A	U	U	U	C	U	A	U	U	U	U	A	G	G	U	A	U	U	C	G	G	G	G	A
<i>nad6-103</i>	U	U	U	G	U	U	U	C	C	C	A	U	C	C	C	A	G	U	C	U	U	U	C	G	C	G	A	C
<i>rps4-377</i>	A	C	U	A	U	U	C	C	U	C	A	A	G	C	A	A	G	G	C	A	G	C	C	G	A	U	A	A
<hr/>																												
<i>nad4-403</i>	A	U	U	U	C	U	A	A	U	G	A	U	C	G	C	C	G	U	G	U	U	C	C	G	C	A	U	G

Figure S3 Putative coordination of PPR motifs of GRS1 and RNA nucleotides around the editing sites targeted by GRS1.

Top: PPR motifs of GRS1 and their corresponding residues at the 3rd, 6th of one PPR motif and 1th positions of the next motif (1'). Bottom: alignment of the putative cis-specificity regions around the RNA editing sites targeted by GRS1. Editing at *nad1-265*, *nad4L-55*, *nad6-103* and *rps4-377* were inhibited in *grs1*. The edited C is shown in bold and the five nucleotides shared between the four sites are highlighted in grey. Another editing sites, *nad4-403*, share the same five nucleotides, but its editing is not affected in *grs1*. The last S motif of GRS1 is positioned at the -4 nucleotides of edited C.

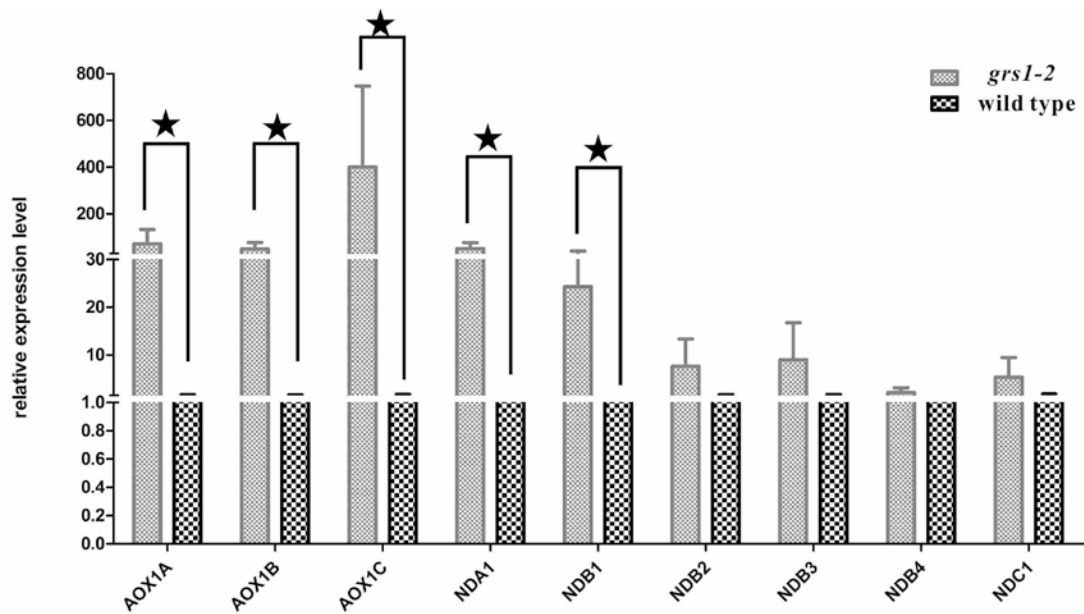


Figure S4 Relative expression of alternative pathway genes in wild type and *grsI-2*. The expression levels of alternative respiratory pathway genes in *grsI-2* increased significantly relative to WT. These genes include three alternative oxidases (AOXs) and six alternative NAD(P)H dehydrogenases (NDs). The values obtained were averaged for three independent experiments, with error bars representing SD. Statistically significant differences between *grsI-2* and wild type are indicated by asterisks (Student's t test; $P < 0.01$).

Table S1 PLS repeat structure of At4g32430

Location	Motif	Amino acid code			Classes
		2nd	5th	35th	
76-110	MDEVTLCLALKACRGDLKRGCOIHGFST	D	T	F	L1
111-141	VCVSNVAVMGMYSRKAGRFDNALCIFENL	C	N	D	S1
142-173	VVSWNTILSGFDDNOIALNFVVRMKSAG	V	N	D	P1
174-208	AFTYSTALSFCVGVSEGFLGLLOLOSTVVK	F	S	D	L1
209-239	LVVGNSFITMYSRSGSFRGARRVFDEMSF	V	N	D	S1
240-275	MISWNSLLSGLSQEGTFGFVAVVIFRDMM	I	N	D	P1
276-310	HVSFTSVITTCCHETDLKLARQIHGLCIKR	V	T	L	L1
311-341	LEVGNILMSRYSKCGVLEAVKSVFHOMS	E	N	N	S1
342-371	VVSWTTMISSNKDDAVSIFLNMRFDDGVY	V	T	N	P1
372-406	EVTFFVGLINAVKCNQIIEGLKIHGLCIKT	V	V	E	L1
407-437	PSVGNISFITLYAKFEALEDKAFEDITFR	S	N	E	S1
438-471	IISWNAMEISGFAQNGFSHEALKMFLSAAA	I	N	N	P2
472-508	EYTFGSVLNIAIFAEDISVKQGRCHAHL	Y	G	C	L2
509-539	PVVSSALLDMYAKRGNIDSEKVFNEMS	V	S	N	S2
540-573	QFVWTSIISAYSSHGDFETVMNLFHKMIK	F	T	D	P2
575-610	LVTFLSVLTACNRKGMVDKGYEIFNMMI	V	L	S	L2
611-642	HEHYSCMVDMLGRAGRLKEAEELMSEV	E	S	G	S2
643-676	ESMLQSMGLGSCRLHGNVKGAKVAELA	S	O	L	E1
677-710	SGSYVOMYNIYAEKEEWDKAAEIRKAM	G	V	K	E2
711-763	EAGFSWIDVGDTEGSLTMQGFSSGDKSHP KSDEIYRMVEIIGLEMNLEGKVAV	A	S	V	DY W

Table S2 The root length of 8-day-old seedlings

genotype	Mean root length (mm)
wild type	42.5 ± 2.6
<i>grs1-1</i>	1.9 ± 0.4
<i>grs1-1 abi5-1</i>	3.7 ± 0.4
<i>grs1-1 gin1-3</i>	1.8 ± 0.3

Table S3 Primes for analysis of RNA editing

Gene	Primers (5' to 3')
<i>atp1</i>	atp1-F1: AGAGCTGCGGAACTAACGAATCTA atp1-R1: AAAGCATCTGGTTCATTTTTCTT
<i>atp6-1</i>	atp6-1-F1: ATGCGACGAATCTTTTTGTTTGA atp6-R1: CCAAGCGAACCCACTTAAAATCTT
<i>atp9</i>	atp9-F1: ATGACAAAGCGTGAGTATAATTCTCAAC atp9-R1: TCAGAATACGAATAAGATCAAAAAGGCCATC
<i>ccb203</i>	ccb203-F1: TGGACACGGGGAGGGAGCAG ccb203-R1: CATAACATAACGGGGCGGGGTTG
<i>ccb206</i>	ccb206-F1: AACCAGCCTTGAAGTGAATG ccb206-R1: GAATCGAGGGTCCAGAAGAG
<i>ccb256</i>	ccb256-F1: CTACGCGCAAATTCTCATTGG ccb256-R1: GAGCGAGTGAAGTACTAGTTTTGGTA
<i>ccb382</i>	ccb382-F1: AATCTTGAAATGAGCACGAAATGTC ccb382-R1: CAGTACCTTTTAGTCGAGTGGCTTC
<i>ccb452</i>	ccb452-F1: TATGCAATTATGAACTCCACGGAAC ccb452-R1: ATAGAGCAGATGGTCCAACACTACATAAC
<i>cob</i>	cob-F1: TGAATAAGGAACCAACGATTCTC cob-R1: AGGTGTGATCAGTCTCATCCGTG
<i>coxII</i>	coxII-F1: GATTGTTCTAAAATGGTTATTCCTCAC coxII-R1: GGTTTGGGGGATTAATTGATTG
<i>coxIII</i>	coxIII-F1: TACCTCCCACCAATAGATAGAGAC coxIII-R1: TTATGATTGAATCTCAGAGGCATTC
<i>matR</i>	matR-F1: GAAGTTTAGACCGCTCACAGTAG matR-R1: GTAGTTGAGTGTTCTTCTTGTTG
<i>nad1</i>	nad1-F1: CCAGCTGAAATACTTGAATAAT nad1-R1: AAAGGTGACTAAAAGACCAGAAAC
<i>nad2</i>	nad2-F1: CGTTCGATCCTCCCACAC nad2-R1: TGAGTGCCATTTGATGAGTAACTG
<i>nad3</i>	nad3-F1: CCATTTTGTGCCCTATCACTTTAC nad3-R1: GCTGTAATGATGTCAGAATTTGCAC
<i>nad4</i>	nad4-F1: TTGCCATGTTGCACTAAGTTAC nad4-R1: CTTCAATGTTGATTCTACTCTATG
<i>nad4L</i>	nad4L-F1: GTTCCCGAAATGGATCTTATCAAAT nad4L-R1: GGAGAGGAATGGTTAACCTTGAATG
<i>nad5</i>	nad5-F1: GATTATTATTCTTGACTTGACTTTG nad5-R1: CGATATGATGATTGGTTTAGGTAC
<i>nad6</i>	nad6-F1: GTAGATCGTGAGTGGGTCAGTCGTC nad6-R1: CCATGATACTTTCTGTTTTGTCGAG

nad7 nad7-F1: TCCCATGACGACTAGGAAAAGG
nad7-R1: CCTATCTATCCACCTCTCCAAACAC

nad9 nad9-F1: GTTCCAAGGACTAGCAAATC
nad9-R1: TCATTTTCAAATATAGTTGGGAGAC

*orf** *orf**-F1: CATTAGCGGCTCATAAACCATG
*orf**-R1: ATTTAGATCTCGGAGATCCCTTTC

orf25 *orf25*-F1: CCACCAAGCTCTCTCGAATG
orf25-R1: TACGAATATGGATGGTAGAAAGATG

orf114 *orf114*-F1: CTAGCAGACTAATCACTAAATAGATAG
orf114-R1: AGTCCTTGGAGCGTCTTTATG

orf240A *orf240A*-F1: CCCTGAGTCGATCTCGCTG
orf240A-R1: TGCGTAGTAGCGTTCTAAGATCAC

orfB *orfB*-F1: ATCTAATCATGCCTCAACTGG
orfB-R1: AGAATCATTTTATGCTTCCTTG

orfX *orfX*-F1: ATCCTTCACTTTTAGCTTTGAATTAC
orfX-R1: ATTGATAGTTACTTTGCCAGGTTC

rpl2 *rpl2*-F1: AGCACTTCTCTATGGGCATTGTA
rpl2-R1: TGCCTGCTTCTAATTGATGACTGG

rpl5 *rpl5*-F1: ATGTTTCCACTCAATTTTCATTAC
rpl5-R1: GAGTTTCCCCCTCATCTTTT

rpl16 *rpl16*-F1: AAAATCGATTATGCTCCTG
rpl16-R1: TACGACCACTGAACAAACTT

rps3 *rps3*-F1: AATCCGATTTTCGGTAAGACTT
rps3-R1: CGTTTTCGGATATAGCACGTC

rps4 *rps4*-F1: GGCTGCTTAAAAAACTGATTC
rps4-R1: TGTCCCTATGAGTGACTACGC

rps7 *rps7*-F1: GGGGGCTTGGATGGTGA
rps7-R1: ATCTGAAATGCGCGAAACTT

rps12 *rps12*-F1: TCAATTGATTCGTCATGGTAGAGA
rps12-R1: TCATATCGATTTGGGTTTTTCTGC

*rps14** *rps14*-F1: ATGTCGTAGAAGCAAATAGTAGA
rps14-R1: TTATGCCCATCAAAGAACC

Ψ *sdh4** Ψ *sdh4*-F: ATGAAGGAACGAAAGAGTGG
 Ψ *sdh4*-R: CCAAGATCTAATTTTCGGGTC

Table S4 Primes for quantitative RT-PCR

Gene	Primers (5' to 3')
<i>ABI5</i>	ABI5-D1:ACTTGAGGATTTCTTGGTGAAGG ABI5-D2:CCTTGAAACACACCATAAAGCTG
<i>ABI3</i>	GTAAGACAACCGAGCGGACAA CATTCGCCGACGGAGAGTTAT
<i>GRS1</i>	GRS1-D1:AGATCAGGAAAGCAATGAGGAAG GRS1-D2:GAAACCTTGCATCGTTAAAGACC
<i>MT18S</i>	mt 18S rRNA-1: TCAGCATGCCGCGGTGAATATG mt 18S rRNA-2: TGCTTCGGGCGAAACCAATTCC
<i>MT26S</i>	mt 26S rRNA-1: TGCGAGGTGGGTAGTTTATC mt 26S rRNA-2: GGCAGGCTTATACCATTACG
<i>CHL 16S</i>	chl 16S rRNA-1: GCGTAGAGATCGGAAAGAAC chl 16S rRNA-2: GGGTATCTAATCCCATTTCGC
<i>CHL 23S</i>	chl 23S rRNA-1: CTATAACGGTCCTAAGGTAGCG chl 23S rRNA-2: CTATTTACCGAGCCTCTCTC
<i>AOX1A</i>	AOX1A-D1:GACGGTCCGTACGGTTTCG AOX1A-D2:CTTCTGATTCGCGTCCTCCT
<i>AOX1B</i>	AOX1B-D1:TGGAGACTTGAAGCTGATGC AOX1B-D2:TCTTTTAGCTCACGACCTTGG
<i>AOX1C</i>	AOX1C-D1:GCATCAAAGCAAGCGACATCC AOX1C-D2:TGGCTTCACGCCCAATAAC
<i>NDA1</i>	NDA1-D1:GTCCGTGAGAGCAAGGAAGG NDA1-D2:GGCGAAGTGGAGGGGATATG
<i>NDB1</i>	NDB1-D1:CTGTGACTGATAAAGATATC NDB1-D2:GCCAACTGCATATACATTC
<i>NDB2</i>	NDB2-D1:CTGACTCTCTCAAAGAGTTCC NDB2-D2:CGATTTGAACTCTTCGATC
<i>NDB3</i>	NDB3-D1:GCAAATCCATCATATAGCAAC NDB3-D2:CAAGGGAGTGAAAGCAAAG
<i>NDB4</i>	NDB4-D1:TCAAGTCTTAAGGGCACAACA NDB4-D2:CGGAAGAGAGAGGCTCTCTCG
<i>NDC1</i>	NDC1-D1:TCCATAAAGAGAGCAAGCAAC NDC1-D2:CGAATCACCTAATGCAAATATC
<i>UBQ10</i>	UBQ10-D1: GGCCTTGTATAATCCCTGATGAATAAG UBQ10-D2: AAAGAGATAACAGGAACGGAAACATAGT