

SUPPLEMENTAL MATERIAL

Control of inflorescence architecture in tomato by BTB/POZ transcriptional regulators

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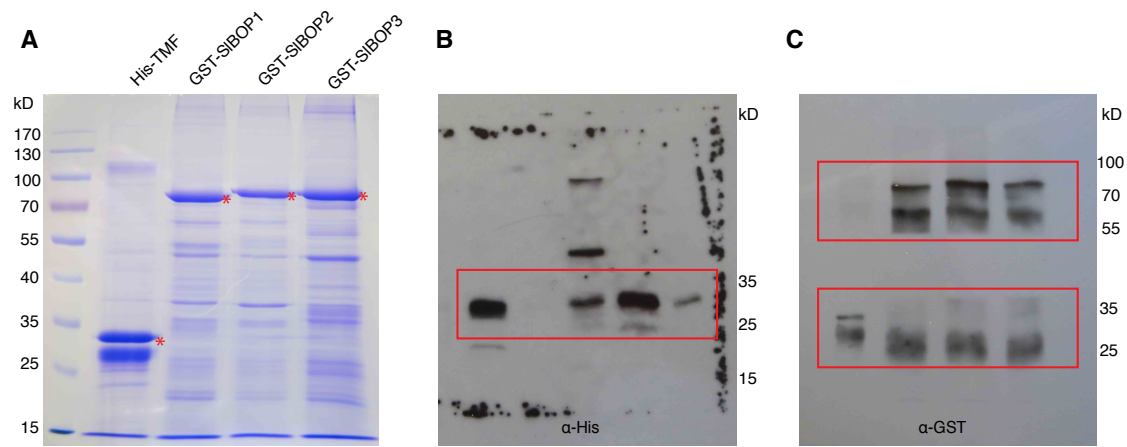
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NPR1      MDTTIDGFADSYEISSTSFVATDNTDSSIVYLAEEQVLTPGDVSAQLQLLSNSFESVFDSP
AtBOP1    MSNTF-----EESLKSMSLDVNLNLLING-----
AtBOP2    -MSNL-----EESLRSLSLDVNLNLLING-----
SlBOP1    MSNNP-----EDPLRSLSDVNLNLLING-----
SlBOP2    -MNTL-----EDSLKTLSDVNLNLLING-----
SlBOP3    -MNTL-----EDSLKTLSDVNLNLLING-----
          ..                               *: *   .: .*: * * .

          BTB/POZ

NPR1      DDFYSDAKLVLDGREVSHRCVLSARSFFKSAALAAAK-----EKDSNNTA----
AtBOP1    -QAFSDVTFV-EGRLVHAHRCILAARSLFRKFFCESDPSQ---P--GAEPANQT-GSG
AtBOP2    -QAFSDVTFV-EGRLVHAHRCILAARSLFRKFFCGTDPQ---PVTGIDPTQHG-SVP
SlBOP1    -QAFSDVTFV-EGRLVHAHRCVLAARSOFRKFFCGPSS---P--QSGP-QLG-SVN
SlBOP2    -QAFSDVTFV-EGRLVHAHRCILAARSLFRKFFCGPES---P--GGGP-DPSVGGF
SlBOP3    -QAFSDVTFV-EGRLVHAHRCILAARSLFRKFFCGPESASVSGP--RLGPFVGGAGLA
          : * * . : : * . : * * * : * * * . * . . . .

NPR1      ---AVKLELKEIAKDYEVGFDVVTVLAYVYSSRVPPPKGV--SECADENCCHVACR
AtBOP1    AR---AAVGGVIVPNSVGVYEVFLLLQLYLSGQVSIYPKHHEPRSNCGDRGCWHTHCT
AtBOP2    ASPTRGSTAPAGIIPVNSVGVYEVFLLLQLYLSGQVSIYPQKHEPRPNCGERGCWHTHCS
SlBOP1    GPRDGTSPASSVIVPNSVGVYEVFLMMQLYLSGQVSIYPQKHEPRPNCGERGCWHTHCT
SlBOP2    SPRTSTSSQVIVPNSVGVYEVFLMLQLYLSGQVSIYPQKHEPRPNCGERNCWHTHCT
SlBOP3    SSPRGTTSCSQVIVPNTVGVYEVFLMLQLYLSGQVSIYPQKHEPRPNCGERNCWHTHCT
          : * * : : : * * * : * * * . * . . . .

NPR1      PAVDFMLEVLYLAFIFKIPELITLYQRHLDDVVKVIEDTLVILKLANICGKACMKLLD
AtBOP1    AAVDLSDILDAAARYFGVEQLALLTQKHLTSMVEKASIEDVMKVLIASR--KQDMHQLWT
AtBOP2    AAVDLALDTLAARSYFGVEQLALLTQKQLASMEKASIEDVMKVLIASR--KQDMHQLWT
SlBOP1    SAVDLALDTLAARSFGVEQLALLTQKQLAMVEKASIEDVMRVLIASR--KQDMQLWT
SlBOP2    SAVDLALDTLAARSFGVEQLALLTQKQLTSMVEKTSIEDVMKVLIASR--KQDMPLWT
SlBOP3    SAVDLALDTLAARSFGVEQLALLTQKHLISMVEKASIEDVMKVLIASR--KQDMHQLWT
          . * * * : * * : * : * * * * : * * * : * * * . * . . . : * * .

NPR1      RCKEIIVKSNVDMVSLKSLPEELVKEIID-----RRKELGLEV
AtBOP1    TCSYLIKASGLPQEIILAKHLPIDVVKIIEELRLKSSMPLRSLMP-----HHDLTSTL
AtBOP2    TCSHLVAKSGLPPEILAKHLPIDVVKIIEELRLKSSIARRSLMPPH-----HHDLVSAQ
SlBOP1    TCSHLVAKSGLPPEILAKHLPIDVVKIIEELRLKSNLARRSLMPPHH-----HHDLSSSA
SlBOP2    TCSHLVAKSGLPPEILAKHLPIDVVKIIEERLKTSLARRSLISH-----HQHDLSSST
SlBOP3    TCSHLVAKSGLPPEILAKHLPIDVVKIIEELRLKSSIARRSLIPHHHHHQHQHMSNI
          * . : . * * : * * * * : * * * : * * * . . . .
          Ankyrin repeat

NPR1      PKVKHVSNVHKAALDSDIELVKLLKEDHTNLDACALHFVAVYCNVKTATDLLKDLDA
AtBOP1    DLEDQKIRRMRRALDSDVLEVKLMVMGEGLNLDSESLALHYAVENCSREVVKALLELGA
AtBOP2    DLEDQKIRRMRRALDSDVLEVKLMVMGEGLNLDSESLALHYAVENCSREVVKALLELGA
SlBOP1    ELEDQKIRRMRRALDSDVLEVKLMVMGEGLNLDSEIALHYAVENCSREVVKALLELGA
SlBOP2    ELEDQKIRRMRRALDSDVLEVKLMVMGEGLNLDSEIALHYAVENCSREVVKALLELGA
SlBOP3    ELEDQKIRRMRRALDSDVLEVKLMVMGEGLNLDSESLALHYAVENCSREVVKALLELGA
          . : : . . * * * * : * * * * : * * * * : * * * * : * * * * . * . . . * *

NPR1      DVNHRN-PRGYTVLHVAAMRKEPQILSLEKASASEATLEGRTALMIAKQATMAVECN
AtBOP1    DVNYPAGPTGKTALHIAAEMVSPDMVAVLLDHDHPNVQTVGDIPLDLRLTLT-----
AtBOP2    DVNYPAGPKGTPPLHIAAEMVSPDMVAVLLDHDHPNVRTVGGITPLDLRLTLT-----
SlBOP1    DVNFPAGPKGTPPLHIAAEMVSPDMVAVLLDHDHPNVRMLDGIPLDLRLTLT-----
SlBOP2    DVNHPAGPKGTPPLHIAAEMVSPDMVAVLLDHDHPNVRTMDGITPLDLRLTLT-----
SlBOP3    DVNYQAGPCKSPLHIAAEMVSPDMVAVLLDHDHPNQTVDGITPLDLRLTLT-----
          : * * . * * : * * * : * * * : * * * . . . . : * * * * . *

NPR1      NIPEQCKHSLKGRLCVEILEQEDKREQIPRDVPPSFAVADELKMTLLDLENRVALAQR
AtBOP1    ---SDFLFGK-----AIPGLTHIEPNKLRCL-----
AtBOP2    ---SDFLFGK-----AIPGLTHIEPNKLRCL-----
SlBOP1    ---SDFLFGK-----AIPGLTHIEPNKLRCL-----
SlBOP2    ---SDFLFGK-----AIPGLTHIEPNKLRCL-----
SlBOP3    ---SDFLFGK-----AIPGLTHIEPNKLRCL-----
          . . . : * * * * : * * * : * * * : * * * .

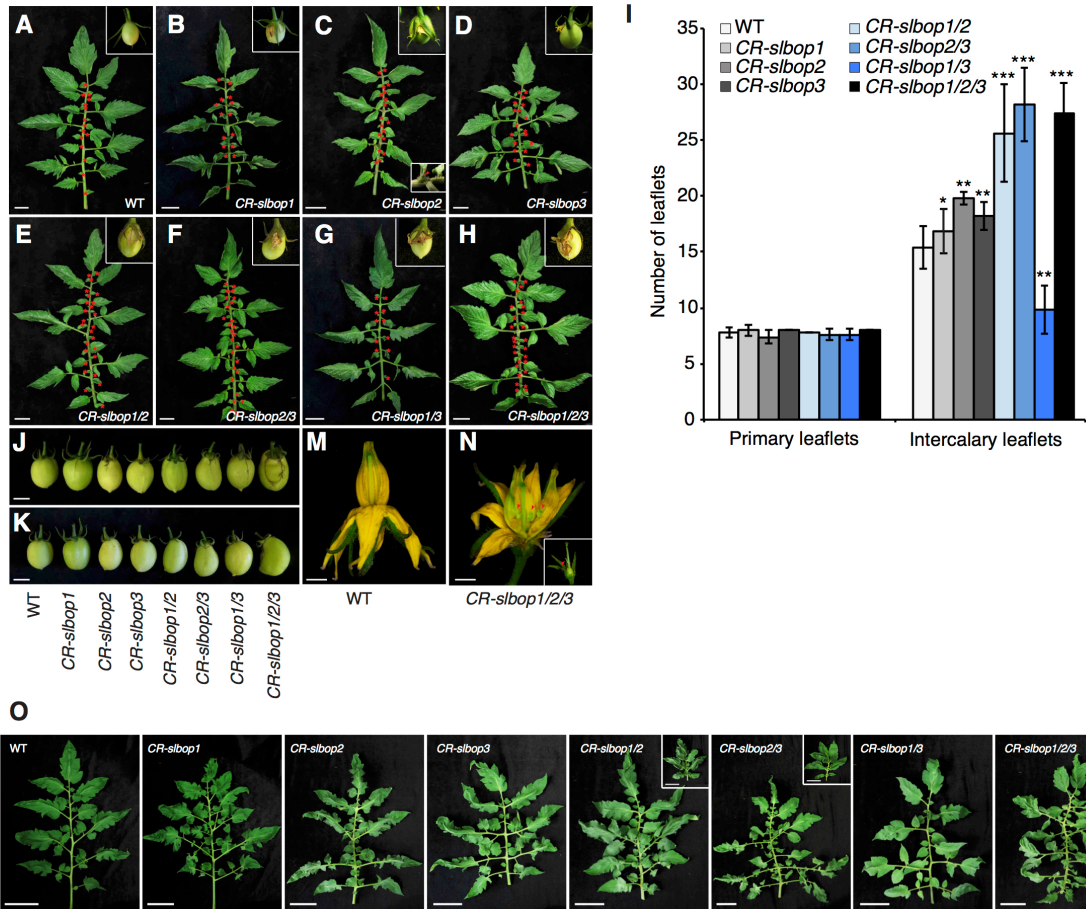
NPR1      FPTEAQAAMEIAEMKGTCEFIVTSLDPRLTGTKRTPGKVIAPPRILEEHQSRKALKS
AtBOP1    -----ELVQSAALVISREEGN---SNDNNTMIYPRM--KDEHTS-----
AtBOP2    -----ELVQSAAMVISREEGNNSNQNNDNTGIYPHM--NEEHNS-----
SlBOP1    -----ELVQSAAMVISREEGSAN---IDLSSTNIYFPNNMSDDHTS-----
SlBOP2    -----ELVQSAAMVISREETTN---NVPSSTAIYQPIN--NEDHGS-----
SlBOP3    -----ELVQSAAMVISREEGEAN---NNQSSENMYTHI--REDHSS-----
          * : . . : : * * * * : * * * : * * * .

NPR1      TVELGKRFFPRCSAVLDQIMNCEDLTQLACGEDDTAEKRLQKQRYMEIQETLKKAFSED
AtBOP1    -----GSSLDRLV-----YLNLGATNR-----
AtBOP2    -----GSS-----GSSNNLDSRLV-----YNLNGACTGMGPGR-----
SlBOP1    -----STST-----SGTN--NIDSRMV-----YNLGGGVV-----
SlBOP2    -----STNTTVGGNNLDSRMV-----YNLGAGTSIAQQIG-----
SlBOP3    -----STSS-----GNNLDSRMV-----YNLNGANVANHHHQ
          . . . : * * * * : * * * : * * * .

NPR1      NLELGNSSLDSTSTSKSTGGKRSNRKLSHRRR---
AtBOP1    DIGDDNSQREGMNLHHHH---DPSTMYHHHH--HF
AtBOP2    DQGDHNSQREGMSRHHHHQ---DPSTMYHHHHQHF
SlBOP1    ---NTSSTHD-----HPSSMYHSSH--EY
SlBOP2    CNRMTNQDDHNSHNKQRHGEFDPSSMYRPS---
SlBOP3    ACKMNNNDHDCSSSHNQ---NPSTMYHHHHSQY
          . . : . .

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Supplemental Figure S2. Alignment of SIBOP1/2/3 with NPR1 and BOP1/2 proteins from *Arabidopsis* (AtBOP1/2). Single and double lines indicate the BTB/POZ and ankyrin repeat domains, respectively. Blue color highlights the two essential domains and two oxidized cysteine residues in NPR1 that are required for its SA-induced transcriptional regulation, which are absent in SIBOP proteins. Green color and arrow noted the cysteine residue in NPR1 that is critical for its S-nitrosylation and SA-induced oligomerization. Red color highlighted the four conserved cysteine residues in the BTB/POZ domain that are shared among all six BTB/POZ proteins and are important for oligomerization of NPR1. The highly conserved domain marked by the four cysteine residues is labeled by red rectangles.



Supplemental Figure S3. Pleiotropic vegetative and reproductive phenotypes of *CR-slbop* single and higher-order mutant plants. (A-H) Representative mature leaf from WT (A), single mutants of *CR-slbop1* (B), *CR-slbop2* (C), *CR-slbop3* (D), double mutants of *CR-slbop1/2* (E), *CR-slbop2/3* (F), *CR-slbop1/3* (G), and *CR-slbop1/2/3* triple mutants (H) showing altered leaf complexity and floral organ abscission defects (insets). Inset in (C) shows ectopic shoot generation on the rachis of a *CR-slbop2* leaf. Red asterisks indicate intercalary leaflets and floral abscission defects, respectively. (I) Quantification and statistical comparison of leaflets in WT and *CR-slbop* mutants. (J, K) Representative fruits from WT and *CR-slbop* mutants showing scars on fruits (J) and fruit shape phenotypes (K) in *CR-slbop* mutants. (M, N) Representative flowers from WT (M) and

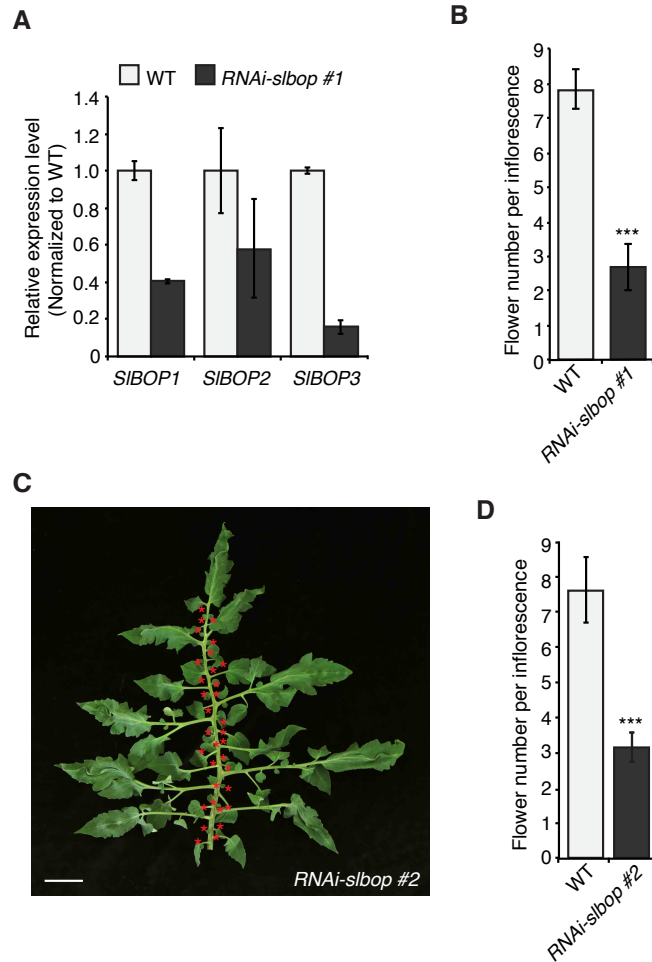
CR-slbop1/2/3 triple mutants (*N*) showing ectopic twisted stigmas fused with stamens in *CR-slbop1/2/3* triple mutants. Inset shows ectopic stigma fused to the carpels of an ovary, explaining the origin of fruit scarring. (*O*) Representative leaves from single and higher order *CR-slbop* mutants grown under greenhouse conditions, where leaf phenotypes were often enhanced. For example, *CR-slbop1* leaves appeared larger than WT, primarily due to longer petiolules, and *CR-slbop2* leaves developed more intercalary leaflets that were frequently fused along the rachis. Insets show rare examples of more extreme leaves. Data are means (\pm s.d.); $n = 5$. A two-tailed, two-sample Student's *t* test was performed, and significant differences are represented by black asterisks: * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$. Scale bars, 2 cm (*A-K*), 500 μ m (*M, N*), 5 cm (*O*).

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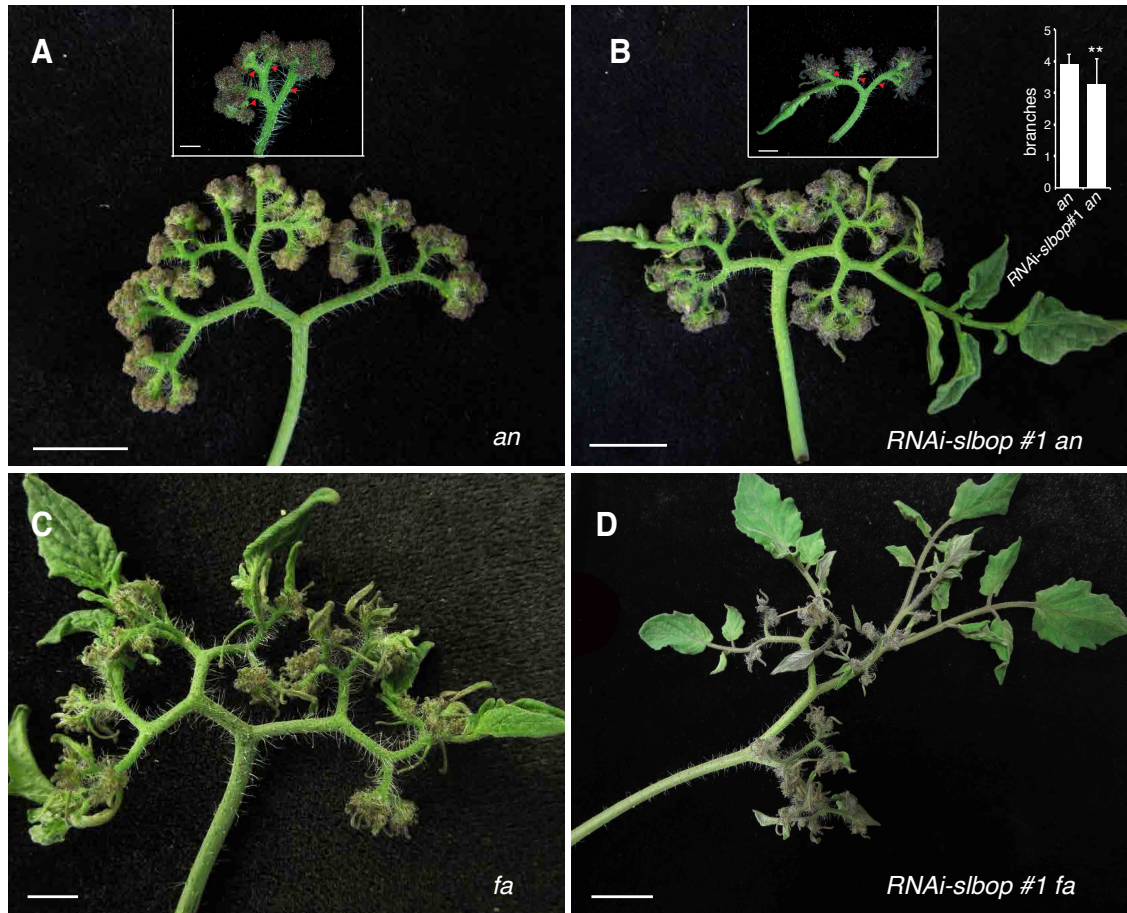
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SlBOP2      ---ATGAATACCTTTGAAGATTCCCTTAAAACTCTCTCTTTAGATTATCTAAATCTTCTC
SlBOP3      ---ATGAATACCTTTGAAGATTCCCTTAAAACTCTCTCTTTAGATTATCTAAATCTTCTC
          *   **** *   *****   ***** *   *****   *****   *****
SlBOP1      ATCAATGGTCAAGCTTTCAGTGATGTTACTTTTCATGTTGAAGGTCATTTAGTCCATGCT
SlBOP2      ATCAATGGTCAAGCTTTCAGTGATGTTACTTTTCAGTGTTGAAGGTCGTTAATACATGCT
SlBOP3      ATCAACGGTCAAGCTTTCAGTGATGTTACTTTTCAGTGTTGAAGGTCGTTAATACATGCT
          *****   *****   *****   *****   *****   *****
SlBOP1      CACCGTTGCGTCTTGGCAGCAAGGAGTCAATTCTTCAGAAAATTTTTCGCGGGCCGAGC
SlBOP2      CATAGATGCATCTTAGCTGCTAGAAGTCTTCTTCCGCAAATCTTTCGCGGGCCGAGC
SlBOP3      CATAGATGCATCTTAGCTGCTAGAAGTCTTGTGTTAGAAAATCTTTCGCGGGCCGAGC
          * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * * *
SlBOP1      TCTCCTCAGTCCGGCCCGCAACTCGGCT-----CGGTTAACGGG
SlBOP2      TCAGCATCTGTCTCCGGCCCGGGTTGGGCCCGTTCGGCGTGGCGTGGATTAGCGTGG
SlBOP3      TCGCCTGGTGGCGGTCCAGAC-----CCTTCGGTGGGCTTTGGAAGC
          * * * * * * * * * * * * * * * * * * * * * * * * * * * * *
SlBOP1      CCGAGAGATACTGGTTCACCAGCATCATCAGTAGTGATACCGGTGAATTCAGTAGGATAT
SlBOP2      TCGCCGAGGGGCACAACAAGTTGTTTCAAGTAGTAATACCTGTGAACATGTAGGGTAT
SlBOP3      CCGCGGACTAGTACTACCAGTAGTTCACAGGTAGTAATACCTGTGAACTCGGTAGGGTAT
          * * * * * * * * * * * * * * * * * * * * * * * * * * * * *
SlBOP1      GAGGTGTTTTTATGATGATGCAGTTTTTATATAGTGGACAAGTATCAATTTGACCGCAA
SlBOP2      GAGGTTTTTTTTGTTGATGTTACAAATTTTGTATAGTGGACAAGTTCATTTGACCGCAA
SlBOP3      GAGGTTTTTTTTGTTGATGTTACAAATTTTGTATAGTGGACAAGTTCATTTGACCGCAA
          *****   *****   ***** * * * * * * * * * * * * * * * *
SlBOP1      AAACATGAGCCAAGGCCAAATTTGTGGAGAGAGAGGTTGTTGGCATAACACATTGCACCTCA
SlBOP2      AAACATGAACCAAGGCCAATTTGTGGTGAAGAAACTGTTGGCATAACACATTGCACATCA
SlBOP3      AAACATGAACCAAGGCCAATTTGTGGTGAAGAAACTGTTGGCATAACACATTGCACATCA
          *****   *****   ***** * * * * * * * * * * * * * * * *
SlBOP1      GCCGTTGATCTTGCACTTGATACACTCTCAGCCGCTAGATCTTTGGTGTGAACAACCTT
SlBOP2      GCCGTTGATCTTGCACTTGATACACTCTCAGCCGCTAGATCTTTGGAGTTGAACAACCTT
SlBOP3      GCCGTTGATCTTGCACTTGATACACTCTCAGCCGCTAGATCTTTGGTGTGAACAACCTT
          *****   *****   *****   *****   *****   *****
SlBOP1      GCTTTGCTCACTCAGAAGCAATTGGCAATCATGGTAGAAAAAGCTTCAATTGAGGATGTG
SlBOP2      GCTTTGCTTACTCAGAAGCATTTGATAAGCATGGTGGAAAAAGCTTCAATTGAGGATGTG
SlBOP3      GCTTTGCTTACTCAGAAGCAATTGACAAGCATGGTAGAAAAACTTCAATTGAGGATGTG
          *****   *****   ***** * * * * * * * * * * * * * * * *
SlBOP1      ATGAGAGTTCTAATAGCATCAAGGAAGCAAGACATGAATCAACTATGGACTACGTGTTCA
SlBOP2      ATGAAAGTTTTAATAGCTTCAAGAAAACAAGACATGCATCAACTTTGGACTACTTGTTCAT
SlBOP3      ATGAAAGTTTTAGTTGCTTCAAGAAAACAAGACATGCCTCAACTTTGGACTACTTGTTCAT
          *****   *****   ***** * * * * * * * * * * * * * * * *
SlBOP1      CATTTGGTTGCAAAATCAGGTCTTCCACCCGAAATGTTGGCCAAACACCTCCCCATTGAT
SlBOP2      CATTTGGTTGCTAAATCAGGTCTCCACCTGAAATCCTAGCCAAACACCTCCCTATTGAT
SlBOP3      CATTTGGTTGCAAAATCAGGCTTCCACCTGAAATCCTAGCAAAACACCTCCCATCGAC
          *****   ***** * * * * * * * * * * * * * * * * * * * *
SlBOP1      GTTGTAGCCAAAATGAGAAGTACGCCTCAAATCCAACCTAGCACGTCGATCCTTAATG
SlBOP2      GTTGTGTCAAAATCGAAGACCTTCGTCTCAAATCCTTATATCGAGAAGATCTTAAATC
SlBOP3      GTTGTGCGAAAATCGAGGAAATTCGCCTAAAACCTTATTAGCGCGAAGATCCTTGATC
          *****   *****   ***** * * * * * * * * * * * * * * * *
SlBOP1      CCACATCATCATCACACCTCGA-----CCTCAGCTCTTCAGCTGAGCTCGAG
SlBOP2      CCTCATCATCATCACACCATCAACATCAACATCAAATGTCTCGAACATTGAGCTCGAG
SlBOP3      TCTCACATCACACGACGAC-----CTAAGTTCGACCTCGGAGCTCGAG
          * * * * * * * * * * * * * * * * * * * * * * * * * * * * *
SlBOP1      GACCAAAAATCCGTAGGATGAGACGAGCCCTTGACTCATCGGACGTTGAACTTGT
SlBOP2      GACCAGAAGATCCGACGAATGAGACGAGCGCTCGATTTCGTCGGACGTTGAACTTAGT
SlBOP3      GACCAAAAATCCGACGAATGAGACGAGCCCTAGACTCATCGGACGTTGAACTTGGT
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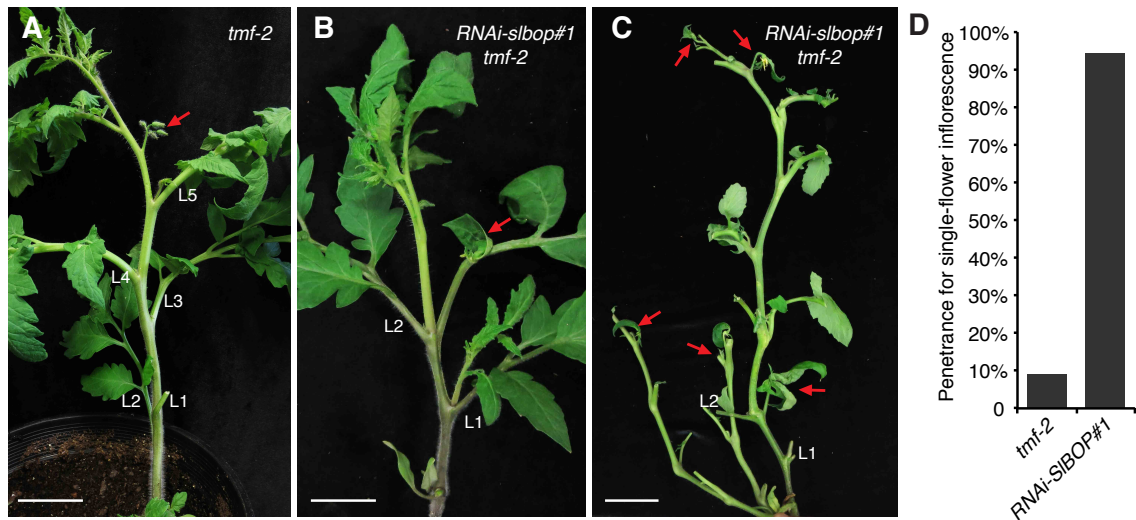
Supplemental Figure S4. Alignment of *SlBOP1*, 2, 3 coding sequences used for generating the hairpin RNAi construct to knockdown *SlBOP1*, 2 and 3 transcript levels in *RNAi-slbop* plants. Note the stretches of nucleotide consensus among all three genes, indicated by red stars.



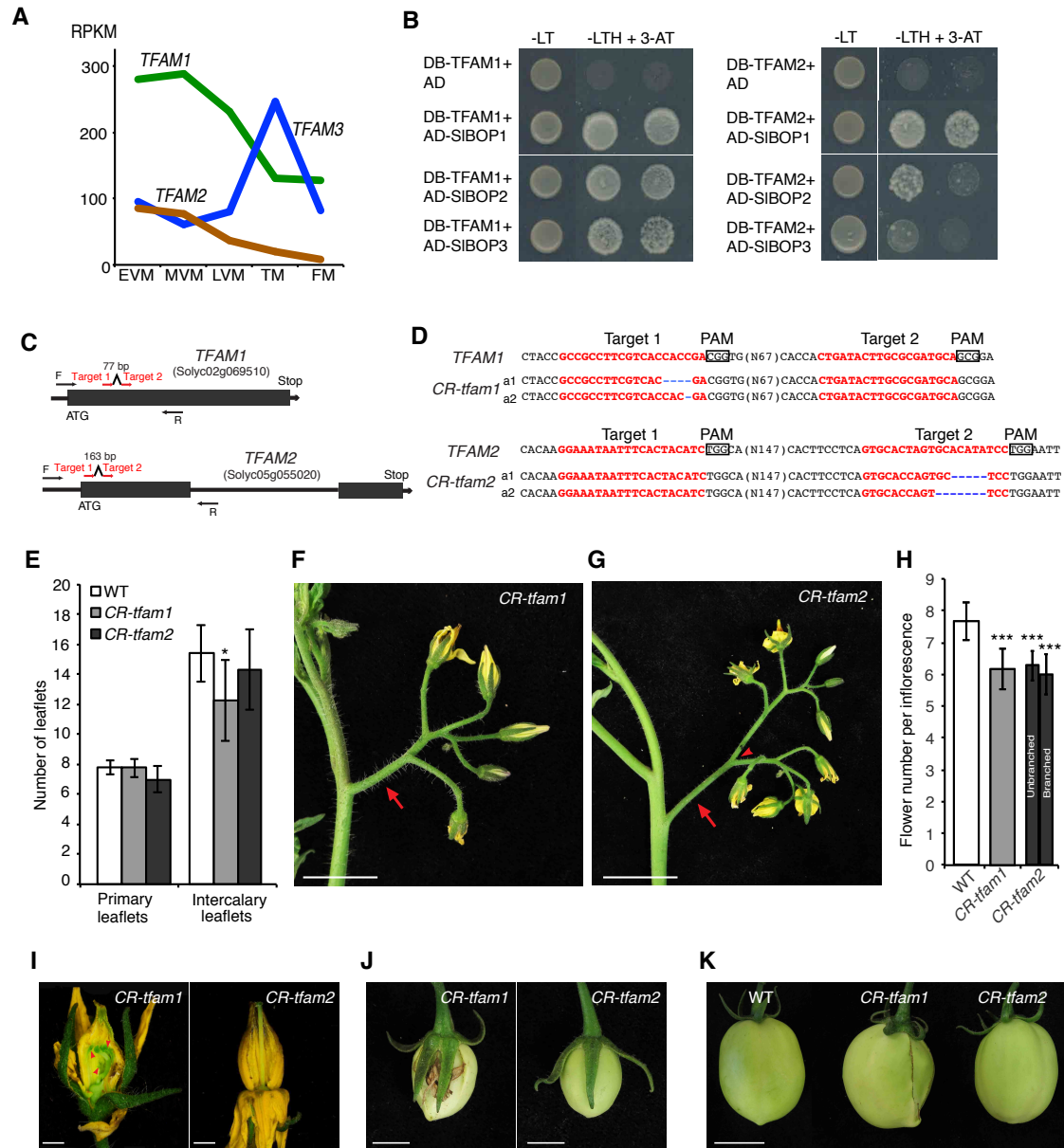
Supplemental Figure S5. Molecular and phenotypic analysis of *RNAi-slbop* plants. (A) qRT-PCR showing transcriptional knock-down of *SIBOP1*, 2 and 3 at the TM stage of *RNAi-slbop #1* used for genetic analyses shown in Figure 6. (B) Quantification and comparison of flowers per inflorescence in WT and *RNAi-slbop #1* plants. (C) Representative mature leaf from *RNAi-slbop #2* plants. (D) Quantification of flower number per inflorescence from an additional *RNAi-slbop* transgenic line. Data are means ($\pm s.d.$); $n = 3$ (A), 12 (B, D). A two-tailed, two-sample Student's *t* test was performed, and significant differences are represented by black asterisks: *** $P < 0.001$. Scale bar, 5 cm.



Supplemental Figure S6. Inflorescence phenotypes of *RNAi-slbop an* and *RNAi-slbop fa* mutant plants. (A, B) A representative inflorescence for *an* mutant (A) and *RNAi-slbop an* mutant plants (B). *RNAi-slbop an* plants branch less frequently than *an* mutants, as measured by the first branches, and thus SIMs, produced during early inflorescence development (insets and bar graph). (C, D) A highly branched leafy inflorescence from *fa* mutant (C) and *RNAi-slbop fa* mutant plants (D). Data are means (\pm s.d.); $n = 21$. A two-tailed, two-sample Student's t test was performed, and significant differences are represented by black asterisks: $**P < 0.01$. Scale bars, 2 cm.



Supplemental Figure S7. *RNAi-slbop* enhances a weak allele of *tmf*. (A) Representative primary shoot of the weak *tmf-2* mutation that flowers early, but shows normal multi-flowered primary inflorescences. (B) Introducing *RNAi-slbop* into the *tmf-2* background by crossing results in earlier flowering compared to *tmf-2* alone, and nearly full penetrance for single-flower primary inflorescences that also exhibit enhanced leaf-like sepals. (C) A whole plant view of *RNAi-slbop tmf-2* plants showing enhanced early flowering and single-flower inflorescences in both primary and side shoots showing leaf-like sepals. Red arrows indicate inflorescences. Leaves were removed to show inflorescences. (D) Quantification and comparison of the penetrance for the single-flower primary inflorescence phenotype in *tmf-2* and *RNAi-slbop tmf-2* plants. L: leaf. Scale bars: 2.5 cm (A, B), 5 cm (C).



Supplemental Figure S8. CRISPR/Cas9-generated null mutations in *TFAM1* and 2 result in phenotypes that match multiple defects of *sibop* single mutants. (A) Normalized RNA-seq read counts for *TFAM1*, 2, and 3 in five stages of primary shoot meristem maturation showing dynamic expression of *TFAM1* and *TFAM2*, similar to *TMF* and *SIBOP1* and 2 (Fig.3). (B) Yeast two-hybrid assays showing *SIBOP1*, 2, and 3 interact

with TFAM1 and TFAM2. (C) Schematics illustrating two sgRNAs (red arrows) targeting *TFAM1* and *TFAM2*. Black arrows represent PCR genotyping primers. (D) Sequences from homozygous T1 transgenic progeny plants lacking the *Cas9* transgene showing out-of-frame deletions (blue dashed lines) for *CR-tfam1* and 2, resulting in two independent null loss-of-function alleles (a1, a2). Red font highlights sgRNA targets, and black boxes indicate PAM sequences. (E) Quantification and statistical comparison of leaflets in WT and *CR-tfam1* and 2 mutants. (F, G) Representative inflorescences (red arrow) from *CR-tfam1* (F) and *CR-tfam2* mutants (G). Red arrowhead in (G) indicates a single branching event often observed in *CR-tfam2* mutants, similar to *CR-slbop2* mutants (Figure 4D). (H) Quantification and statistical comparison of flowers per inflorescence in WT and *CR-tfam1* and 2 mutants. Note that flower number for *CR-tfam2* plants was quantified as the flowers on each inflorescence branch from branched inflorescences, and from unbranched inflorescences separately. (I) Representative flowers from *CR-tfam1* (left), *CR-tfam2* (right) mutants showing ectopic twisted stigmas fused with stamens (arrowheads) in *CR-tfam1*. (J, K) Representative fruits from *CR-tfam1* and 2 showing floral organ abscission defects (J), scars on fruits, and altered fruit shape (K) in *CR-tfam1*, similar to *CR-slbop* single mutants (Supplemental Fig. S3). Scale bars, 2 cm (F, G, K), 500 μ m (I, J). Data are means (\pm s.d.); $n = 5-11$ (E); $n = 11-21$ (H). A two-tailed, two-sample Student's *t* test was performed, and significant differences represented by black asterisks: * $P < 0.05$, *** $P < 0.001$.



Supplemental Figure S9. Driving expression of the flower specification gene *AN* in the earliest stage of meristem maturation using *TMF* promoter (Transactivation, LhG4, OP system) results in single-flower primary inflorescence after producing only two leaves. Scale bars, 2 cm.

Gene	Guide RNA sequences		Genotyping primers	
	Target 1	Target 2	Forward	Reverse
<i>SIBOP1</i>	ATTGGGCCGAGCTCTCCTCAGTC	ATTGTGATACCGTGAATTCAGT	CACAAGACAAAACACCAACCA	GTCATGTTTTTGCGGTACA
<i>SIBOP2</i>	ATTGGCCCGCGGTTGGGCCCGTT	ATTGTAATACCTGTGAACACTGT	GCCACTAACCTTTCTCCTATTG	GGCTGATGTGCAATGTGTATG
<i>SIBOP3</i>	ATTGGCCCTGAGTCGCCTGGTGG	ATTGGTAGTAATACCTGTGAACT	CACAAAATCATCTTCCACTATGTGA	TGCAATGTGTATGCCAACAG
<i>TFAM1</i>	ATTGCCGCCTTCGTCACCACCGA	ATTCTGATACTTGCGCGATGCA	AAAGTAACACAGCCATGTTAGACG	GCCGAAAAACGGACAATTAC
<i>TFAM2</i>	ATTGGAAATAATTCACTACATC	ATTGTGCACTAGTGCACATATCC	TTTCCTCCCTATTTCAAGAATCA	TCCTTCAAAAATTACCACCTTATCC

Supplemental Table S1. Single guide RNA (sgRNA) sequences and genotyping primers of *CR-slbops* and *CR-tfams*.

Primer name	Primer Sequence 5' to 3'
SIBOP1-F	TGGCATCACTCCATTGGACA
SIBOP1-R	GATCAATGTTTTCGCTCCCT
SIBOP2-F	GGGTGAAGCGAATAACAATCAA
SIBOP2-R	TGCATGCCATTTGATGATGATG
SIBOP3-F	TCTTCCACCGGATATTCCA
SIBOP3-R	TTCGTCAATCTGTTCACCC
FA-F	CACTGCCTTGATGAGGATGC
FA-R	TCAGAATGGCACAGCTGTTCG
AN-F	GAGCCTTGTTGAAGGGAATG
AN-R	CCTTGTCATTCTCCACTTC
SISEP3-F	CGGCAAACAACCTCAAACCTCA
SISEP3-R	TGATAGGAAAACCATTGAGCA
FUL-F	AGAAAGCGCTCCAAGAACAA
FUL-R	CCGATCCATCTCTGTTCATTCC
S-F	TTCAACTGTTTCTCCTCTTGCT
S-R	CCTATCCAACCTCAAAGC

Supplemental Table S2. qRT-PCR primers used in this study.