

## Sequences

Type-1 singlet from *L. monocytogenes* (Lmo)

NC\_002973.5/1352518-1352823

Lmo plasmid sequence

**GGATCCTAATACGACTCACTATAG**GC GGGTGAATGTAAGCAGAGAGACTG  
CGAAAAGCGGGCGCCGACGGGGAAAGCATGTATTATGTGAAACTCTCAGGCA  
AAAGGATGTTTACGGGACGCAACTCTGGAGTCATTTTTGTGTTACGACAGG  
G**AATTC**

pink – BamHI cut site

blue – T7 promoter

red – G inserted to improve transcription initiation

orange – EcoRI cut site

Lmo transcribed sequence

**G**GC GGGUGAAUGUAAGCAGAGAGACUGCGAAAAGCGGGCGCCGACGGGGA  
AAGCAUGUAUU AUGUGAAACUCUCAGGCAAAGGAUGUUUACGGGACGCA  
ACUCUGGAGUCAUUUUUGUGUUACGACAGGG**AAUU**

Type-2 singlet from *D. hafniens* (Dha)

NC\_011830.1/1219347-1219484

Dha plasmid sequence

**GGATCCTAATACGACTCACTATAGG**CACTGGATGAGGTTTTTCAGGAGAACA  
GGGTAAGCTAACCATGATGAACTGAAAACGGACAGAACTCTGGAGAGTTCC  
GCAAGGACGCCGAAGGGGCAAGACAGCAAAGCTGTTCAATCTCTCAGGCA  
AAAGGACAGAGCG**AATTC**

Dha transcribed sequence

**GG**CACUGGAUGAGGUUUUCAGGAGAACAGGGUAAGCUAACCAUGAUGAA  
CUGAAAACGGACAGAACUCUGGAGAGUCCGCAAGGACGCCGAAGGGGC  
AAGACAGCAAAGCUGUUCAUUCUCUCAGGCAAAGGACAGAGCG**AAUU**

Lmo mutant transcribed sequences

Lmo mBS (U72A)

GGCGGGUGAAUGUAAGCAGAGAGACUGCGAAAAGCGGCGCCGACGGGGA  
AAGCAUGUAUUUGUGAAACUCACAGGCAAAGGAUGUUUACGGGACGCA  
ACUCUGGAGUCAUUUUUGUGUUACGACAGGGAAUU

Lmo Trn ( $\Delta$ 101-129)

GGCGGGUGAAUGUAAGCAGAGAGACUGCGAAAAGCGGCGCCGACGGGGA  
AAGCAUGUAUUUGUGAAACUCUCAGGCAAAGGAUGUUUACGGGACGCA  
A – GAAUU

Lmo mut $\beta$  (A66C A67C A68C)

GGCGGGUGAAUGUAAGCAGAGAGACUGCGAAAAGCGGCGCCGACGGGGA  
AAGCAUGUAUUUGUGCCCCUCUCAGGCAAAGGAUGUUUACGGGACGC  
AACUCUGGAGUCAUUUUUGUGUUACGACAGGGAAUU

Lmo TL ( $\Delta$ 106-125 *Ins* GAGA)

GGCGGGUGAAUGUAAGCAGAGAGACUGCGAAAAGCGGCGCCGACGGGGA  
AAGCAUGUAUUUGUGAAACUCUCAGGCAAAGGAUGUUUACGGGACGCA  
ACUCUG GAGA CAGGGAAUU

Lmo PL ( $\Delta$ 106-125 *Ins* GAUAA)

GGCGGGUGAAUGUAAGCAGAGAGACUGCGAAAAGCGGCGCCGACGGGGA  
AAGCAUGUAUUUGUGAAACUCUCAGGCAAAGGAUGUUUACGGGACGCA  
ACUCUG GAUAA CAGGGAAUU

Dha mutant transcribed sequences

Dha mBS (U123A)

GGCACUGGAUGAGGUUUUCAGGAGAACAGGGUAAGCUAACCAUGAUGAA  
CUGAAAACGGACAGAACUCUGGAGAGUUCGCAAGGACGCCGAAGGGGC  
AAGACAGCAAAGCUGUUCAAUUCUCACAGGCAAAGGACAGAGCGAAUU

Dha Trn ( $\Delta$ 18-52 *Ins* UUCG)

GGCACUGGAUGAGGUUU **UUCG** AAACGGACAGAACUCUGGAGAGUUC  
GCAAGGACGCCGAAGGGGCAAGACAGCAAAGCUGUUCAAUUCUCAGGC  
AAAAGGACAGAGCGAAUU

Dha mut $\alpha$  (A117C A118C)

GGCACUGGAUGAGGUUUUCAGGAGAACAGGGUAAGCUAACCAUGAUGAA  
CUGAAAACGGACAGAACUCUGGAGAGUUCGCAAGGACGCCGAAGGGGC  
AAGACAGCAAAGCUGUUCCCCUCUCUCAGGCAAAGGACAGAGCGAAUU

Dha HL ( $\Delta$ 26-46)

GGCACUGGAUGAGGUUUUCAGGAGA – GAACUGAAAACGGACAGAACUC  
UGGAGAGUUCGCAAGGACGCCGAAGGGGCAAGACAGCAAAGCUGUUCA  
AUCUCUCAGGCAAAGGACAGAGCGAAUU

## Original Singlet Alignment (J. Barrick and R. Breaker, unpublished results)

### Type-1 Singlets

|      |   |             |                 |
|------|---|-------------|-----------------|
|      | Staphylococcus aureus subsp. aureus N315  |             |                 |
| Sau  | DNA                                       | NC_002745.2 | 1577113-1576807 |
| Lmo  | Listeria monocytogenes str. 4b F2365      | NC_002973.5 | 1352518-1352823 |
| Sep  | Staphylococcus epidermidis RP62A          | NC_002976.3 | 1147162-1146812 |
| Lmo' | Listeria monocytogenes EGD-e chromosome   | NC_003210.1 | 1372690-1372995 |
| Lin  | Listeria innocua Clip11262                | NC_003212.1 | 1379476-1379781 |
| Cte  | Clostridium tetani E88                    | NC_004557.1 | 1925262-1925580 |
| Tfu  | Thermobifida fusca YX                     | NC_007333.1 | 1619770-1620134 |
| Kra  | Kineococcus radiotolerans SRS30216 strain | NZ_AAEF0200 | 6425-6800       |

### Type-2 Singlets

|     |   |                |                 |
|-----|---|----------------|-----------------|
|     | Brevibacterium linens BL2               |                |                 |
| Bli | 2662183_Cont246                         | WP_050773381.1 | 20952-20590     |
| Spn | Streptococcus pneumoniae TIGR4          | NP_344931.1    | 387343-387650   |
| Oih | Oceanobacillus iheyensis HTE831         |                | 2942953-2942663 |
| Smu | Streptococcus mutans UA159              | NP_721560.1    | 1115812-1116128 |
| Sth | Streptococcus thermophilus LMG 18311    | WP_011225955.1 | 886124-885797   |
|     | Desulfitobacterium hafniense DCB-2      |                |                 |
| Dha | ctg918                                  | WP_011461681.1 | 5607-5354       |
| Eba | Exiguobacterium sibiricum 255-15 ctg277 | ZP_00539478.1  | 30315-30643     |
| Xfa | Xylella fastidiosa 9a5c                 | WP_031337913.1 | 1336905-1337238 |
| Baq | Bartonella quintana str. Toulouse       | WP_011179698.1 | 1196121-1195790 |













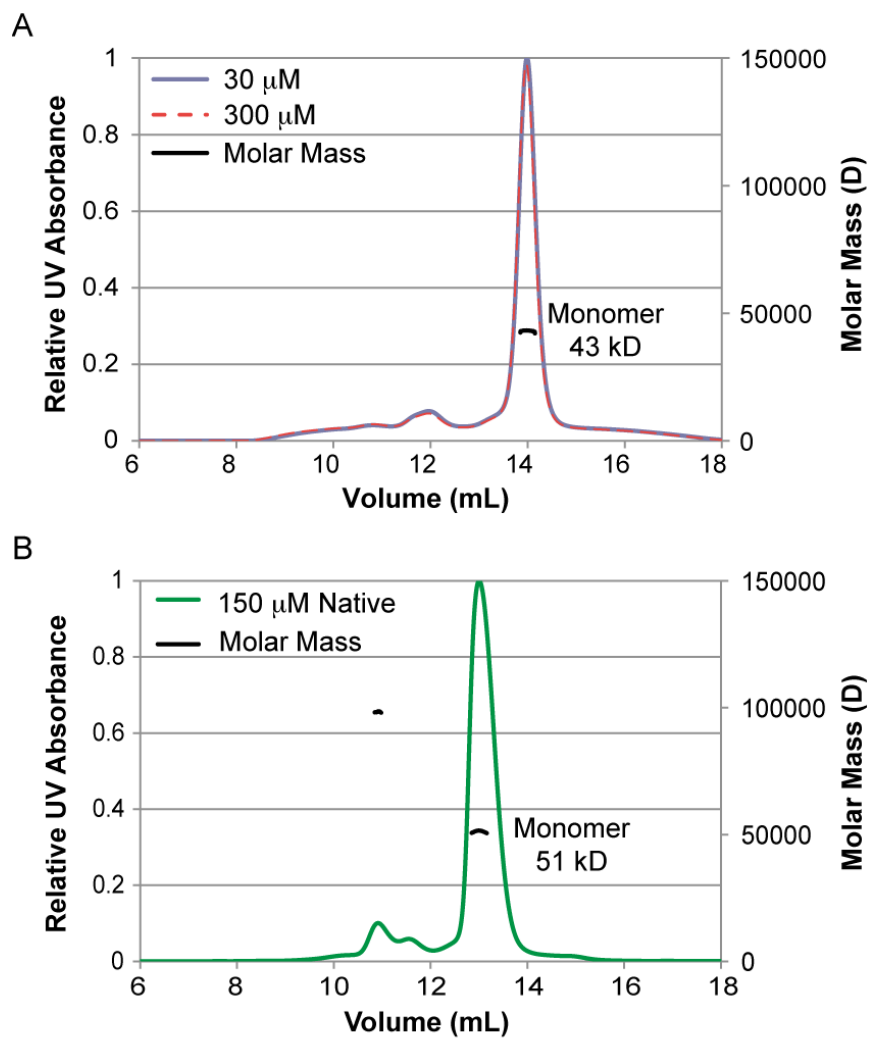


Figure S3: Singlet riboswitches are active as monomers. a) FPLC and MALS show that Lmo type-1 WT is greater than 95% monomer at both 30 and 300  $\mu\text{M}$ . b) Dha type-2 WT is stable and greater than 90% monomer when purified using a native FPLC prep.

## FPLC of Dha mutants

Dha WT that is heat-denatured and refolded forms some dimer, particularly at 300  $\mu$ M RNA, the highest concentration tested (Figure S4). Intriguingly, refolding in the presence of saturating ligand substantially decreases the fraction of the pool in this alternative dimer state ( $47 \pm 1\%$  dimer in the absence of ligand versus  $21 \pm 1\%$  in 5 mM glycine). Because binding and RNA folding are at equilibrium, this suggests that the monomeric form of Dha WT binds glycine with 4-fold stronger affinity than the dimer (corresponding to a .8 kcal/mol  $\Delta\Delta G$ ).

In contrast, the Dha truncation mutant forms much more dimer in the presence of glycine, with the contrast appearing most significantly at lower RNA concentrations (Figure S4). At 30  $\mu$ M RNA, the Dha truncation mutant forms only  $19 \pm 2\%$  dimer in the absence of ligand, compared to  $59 \pm 2\%$  in 5 mM glycine. Because binding and RNA folding are at equilibrium, this indicates that the dimeric form of Dha Trn binds glycine with 9-fold stronger affinity than the monomer (corresponding to a 1.3 kcal/mol  $\Delta\Delta G$ ).

Because a significant portion of the Dha Trn pool exists as dimer across the RNA concentrations where ligand binding occurs, and because the dimer binds ligand more tightly than monomer, we ascribe the unexpected ligand-binding activity of Dha Trn to a spurious dimer state.

The multimeric states of the other Dha mutants were also analyzed by FPLC (Figure S4). All form some dimer, particularly at the highest RNA concentrations tested, but in no other case did ligand binding promote formation of the dimer. Therefore, we attribute the observed ligand-binding affinities to monomer states of the other Dha mutants.

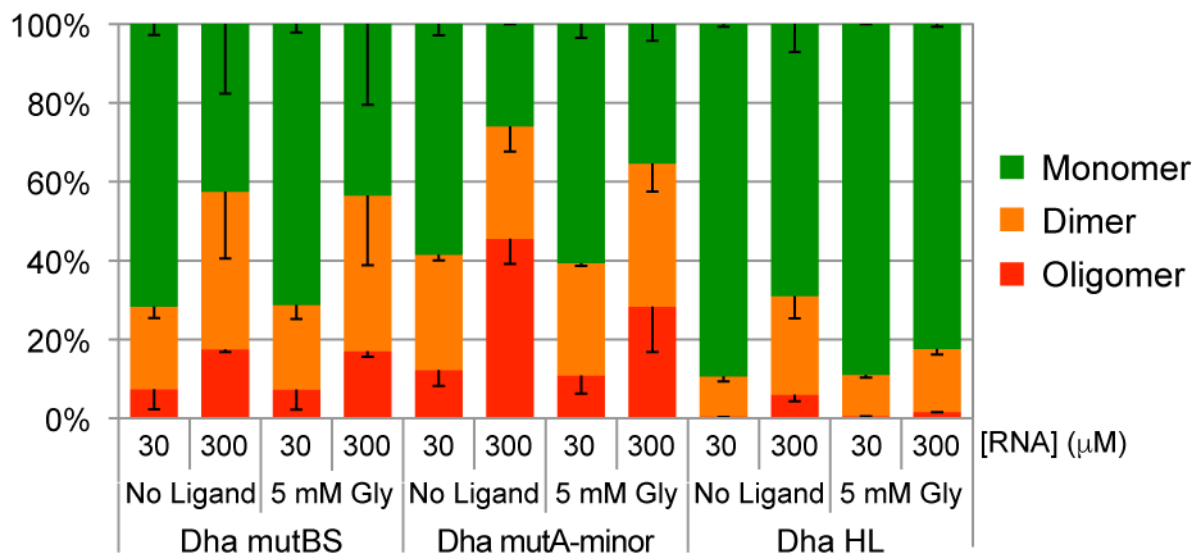
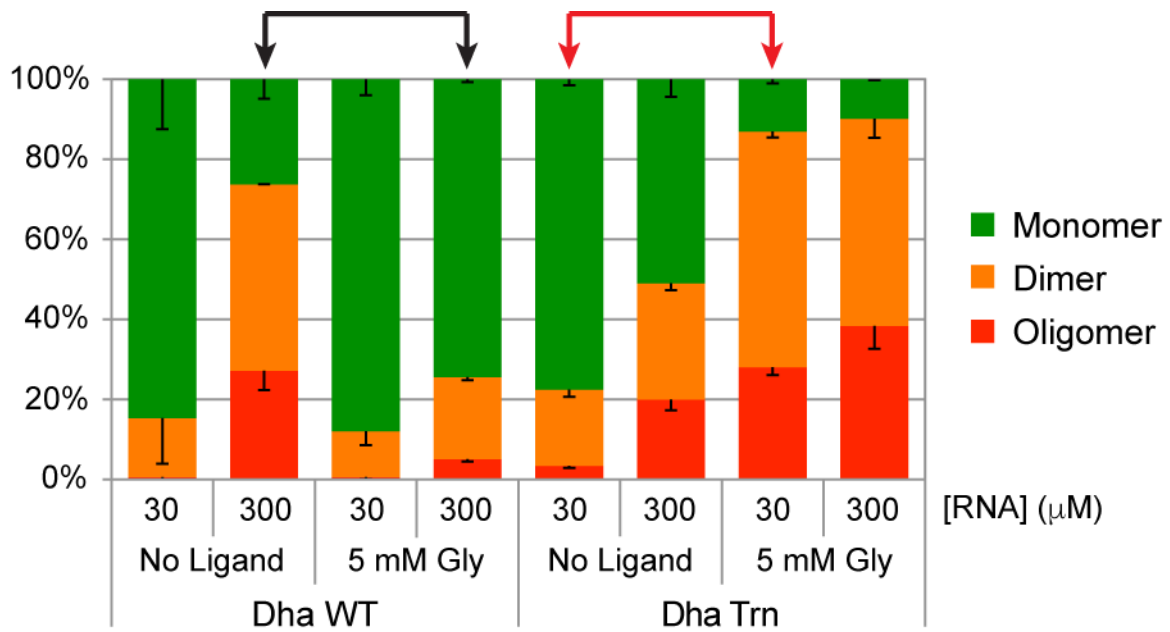


Figure S4: Analysis of multimeric state of Dha WT and mutants by size-exclusion FPLC. Moderate and high concentrations of each construct were refolded in the presence or absence of glycine. Ligand binding stabilizes the monomeric form of Dha WT (black arrows); in contrast, ligand binding stabilizes the dimeric form of Dha Trn (red arrows).

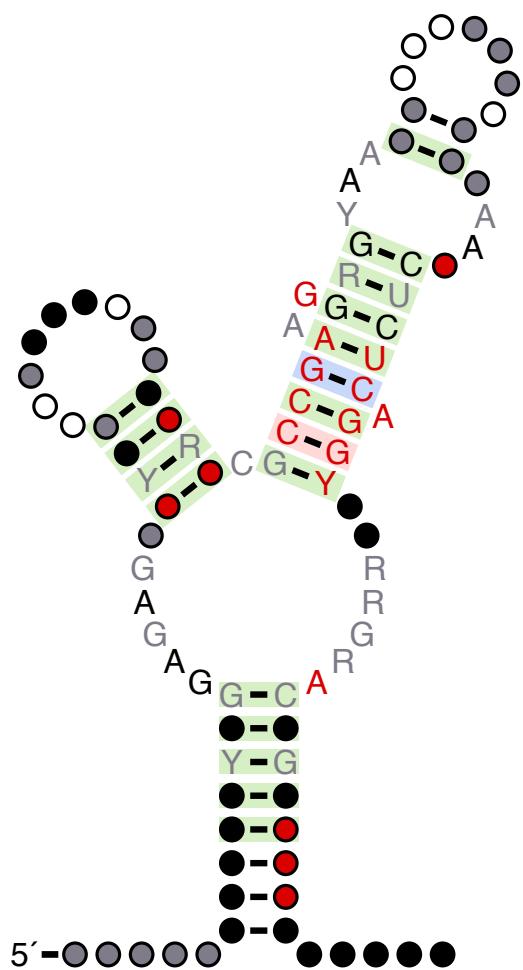


Figure S5: Minimal glycine aptamer consensus sequence used for RefSeq search.