

Structural and haemostatic features of pharmaceutical heparins from different animal sources: challenges to define thresholds separating distinct drugs

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Supplementary Information

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| NaCl - M | % of total heparin sample eluted | | |
|--|----------------------------------|----------------------------|-----------------|
| | HP-I (n = 6) | HB-I (n = 5) | HB-L (n = 1) |
| ≤ 1.85 | 10.18 ± 0.44 ^a | 33.39 ± 2.81 ^a | 18.90 |
| ≤ 1.90 | 27.71 ± 0.56 ^a | 56.20 ± 2.54 ^a | 35.47 |
| ≤ 1.95 | 69.89 ± 0.27 ^a | 86.66 ± 1.28 ^a | 71.60 |
| ≤ 2.00 | 94.98 ± 0.11 | 98.38 ± 0.31 | 94.25 |
| NaCl concentration (as M) at the peak elution | 1.928 ± 0.004 ^a | 1.865 ± 0.010 ^a | 1.921 |

^aSignificant different (P < 0.05) for HP-I vs HB-I.

Table S1. Capacity of retention (mean ± SD) of HP-I, HB-I and HB-L on the anion exchange chromatography.

| | | M_w (Da) | $M > 24,000$ Da (% of total) | Ratio $M 8,000 \rightarrow 16,000$ Da / $M 16,000 \rightarrow 24,000$ Da |
|--|---------|-----------------|---------------------------------|---|
| Recommendation of the USP^b | | 15,000 - 19,000 | <20 | > 1.0 |
| HP-I | Batch 1 | 17,000 | 13.96 | 1.11 |
| | Batch 2 | 17,114 | 14.29 | 1.10 |
| | Batch 3 | 17,025 | 14,00 | 1.10 |
| | Batch 4 | 17,399 | 14,36 | 1.05 |
| | Batch 5 | 17,142 | 14.36 | 1.09 |
| | Batch 6 | 17,045 | 14.04 | 1.10 |
| HB-I | Batch 1 | 18,791 | 20.07 | 0.95 |
| | Batch 2 | 17,275 | 14.88 | 1.32 |
| | Batch 3 | 17,167 | 14.56 | 1.32 |
| | Batch 4 | 17,964 | 16.78 | 1.09 |
| | Batch 5 | 16,958 | 14.28 | 1.37 |
| | Batch 6 | 18,148 | 17.54 | 1.10 |

^aColumns were calibrated using low-molecular-weight heparin and molecular-weight standard heparin calibrants, for further detail see the legend of Fig.2. The results may be slightly different using only the unfractionated heparin calibrant.

^bSee [19].

Table S2. Average molecular weight (M_w), proportions of molecular weight (M) lower than 24,000 Da and molecular weight ratios 8,000 \rightarrow 16,000 Da and 16,000 \rightarrow 24,000 Da measured for HP-I and HB-I^a.

| Signal number ^a | Structure ^c | ¹ H/ ¹³ C-chemical shifts (ppm) | | |
|-------------------------------|--|---|------------|------------|
| | | HP-I | HB-I | HB-L |
| A) α-glucosamine units | | | | |
| 1 | α-GlcN,3,6-triS -[?] | 5.52/98.1 | 5.55/98.2 | 5.51/98.3 |
| 2 ^b | α-GlcN,6-diS -[α-IdA-2S] + α-GlcNAc -[β-GlcA] | 5.40/98.6 | 5.42/98.6 | 5.40/98.6 |
| 3 | α-GlcNS -[α-IdA] | 5.36/97.4 | 5.39/97.5 | 5.34/97.5 |
| 4 | α-GlcNS -[β-GlcA] | 5.58/99.5 | 5.58/99.5 | 5.57/99.6 |
| 5 | α-GlcNS -[α-IdA-2S] | 5.31/98.9 | 5.31/99.3 | 5.30/99.2 |
| B) α-iduronic acid | | | | |
| 6 | α-IdA2S | 5.22/101.0 | 5.24/101.2 | 5.22/101.2 |
| 7 | α-IdA -[α-GlcN,6S] | 5.01/103.9 | 5.03/104.0 | 5.01/104.0 |
| 8 | α-IdA | 4.95/103.7 | 4.96/103.7 | 4.95/103.8 |
| C) β-glucuronic acid | | | | |
| 9 | β-GlcA -[α-GlcN,3,6-triS] | 4.63/102.9 | - | 4.62/103.1 |
| 10 | β-GlcA -[α-GlcNAc] | 4.52/104.2 | 4.54/104.2 | 4.52/104.2 |
| 11 | β-GlcA -[α-GlcNS] | 4.61/103.8 | 4.62/103.9 | 4.61/103.8 |
| 12 | β-GlcA2S -[?] | - | 4.72/102.7 | 4.70/102.7 |
| D) Linkage region | | | | |
| 13 | β-Gal | 5.54/103.3 | 4.55/103.4 | 4.54/103.4 |
| 14 | β-Xyl | 4.46/104.8 | 4.48/104.9 | 4.44/105.0 |
| 15 | β-GlcA -[β-Gal] | 4.68/105.7 | 4.69/105.8 | 4.66/105.7 |

^aSee panels B - D, Fig. 3.

^bThese two units have similar ¹H and ¹³C-chemical shifts of their anomeric signals.

^cThe reported structure is in bold and the subsequent unit is in bracket.

Table S3. ¹H and ¹³C chemical shifts of the anomeric protons and carbons of the constitutive units of HP-I, HB-I and HB-L.

| Heparin | APTT ^a | anti-FIIa ^a | anti-FXa ^a | anti-FIIa/ anti-FXa ^e |
|------------------------------------|---------------------------------|------------------------|-----------------------|-------------------------------------|
| HP-I (batches)^b | 203.0 ± 5.3 / n = 6 | 185.5 ± 14.1 / n = 8 | 195.2 ± 18.3 / n = 11 | 0.95 |
| HB-I (batches)^b | 88.7 ± 8.2 / n = 7 | 90.8 ± 10.2 / n = 9 | 91.7 ± 15.0 / n = 11 | 0.99 |
| HB-L (batches)^c | 97.2 ± 8.6 / n = 9 ^d | 133.8 ± 15.5 / n = 8 | 121.9 ± 16.4 / n = 10 | 1.09 |
| HP-I (parental)^c | 181.8 ± 7.8 / n = 4 | 165.8 ± 12.6 / n = 4 | 173.3 ± 10.8 / n = 6 | 0.95 |
| HB-I (parental)^c | 105.5 ± 11.5 / n = 3 | 121.7 ± 10.9 / n = 4 | 105.3 ± 10.1 / n = 6 | 1.15 |
| HB-L (parental)^c | 97.2 ± 8.6 / n = 9 ^d | 133.8 ± 15.5 / n = 8 | 121.9 ± 16.4 / n = 10 | 1.09 |
| F3 HP-I^c | 168.1 ± 6.8 / n = 3 | 167.4 ± 9.9 / n = 4 | 164.5 ± 4.3 / n = 3 | 1.01 |
| F3 HB-I^c | 89.4 ± 6.7 / n=3 | 105.2 ± 15.7 / n = 4 | 83.5 ± 10.6 / n = 4 | 1.26 |
| F3 HB-L^c | 130.7 ± 4.7 / n=4 ^d | 166.4 ± 9.4 / n = 5 | 168.7 ± 9.5 / n = 3 | 0.98 |

^a Anticoagulant assays expressed in IU mg⁻¹ (mean ± SD / number of assays).

^b n represents the number of different batches analyzed.

^c n represents the number of replicates performed with the same batch.

^d Values significantly lower (P < 0.05) than those obtained in the other assays.

^e Ratio.

Table S4. Anticoagulant activity (mean ± SD) of HP-I, HB-I and HB-L batches (batches), parental batches of F3 derivatives (parental) and F3 derivatives (F3).

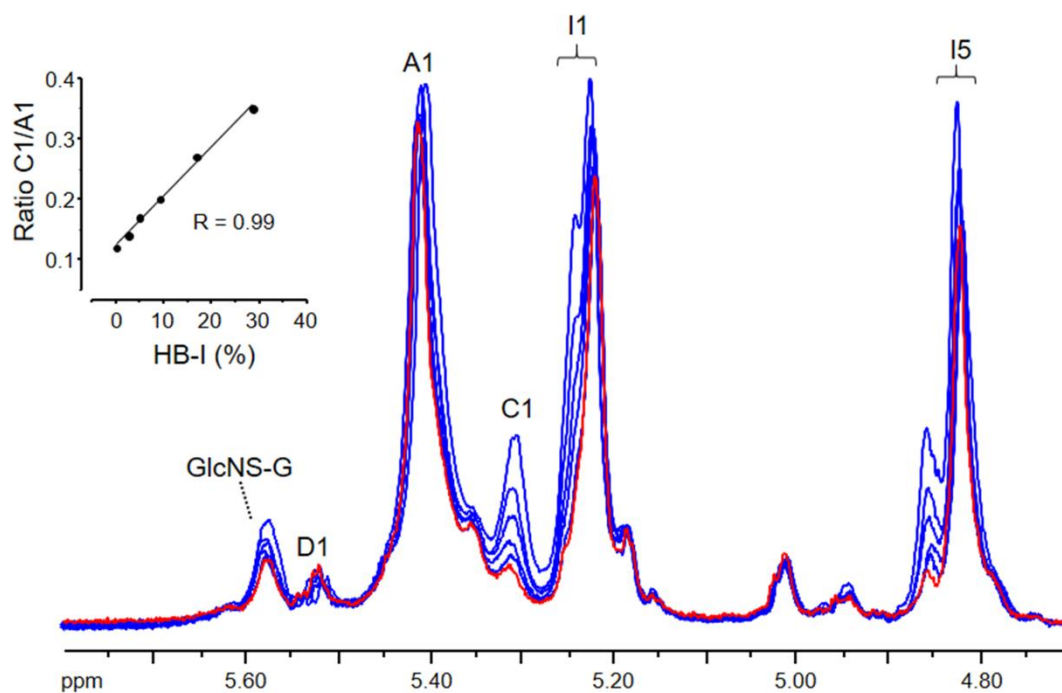


Figure S1. 1D ¹H NMR spectra (5.75 - 4.70 ppm region) of mixtures containing increasing amounts of HB-I added to a fixed amount of HP-I. Increasing quantities of HB-I (0.5 → 8.0 mg, in blue) were successively added to 20 mg of HP-I (in red) dissolved in 0.5 mL of 99.9% D₂O. See the legend of Fig. S1 for the nomenclature of the signals. The inset shows the ratio of peak integral of C1/A1 at different concentrations of HB-I (%) added to HP-I. This assay enables to detect mixtures of HB-I and HP-I up to 4%.

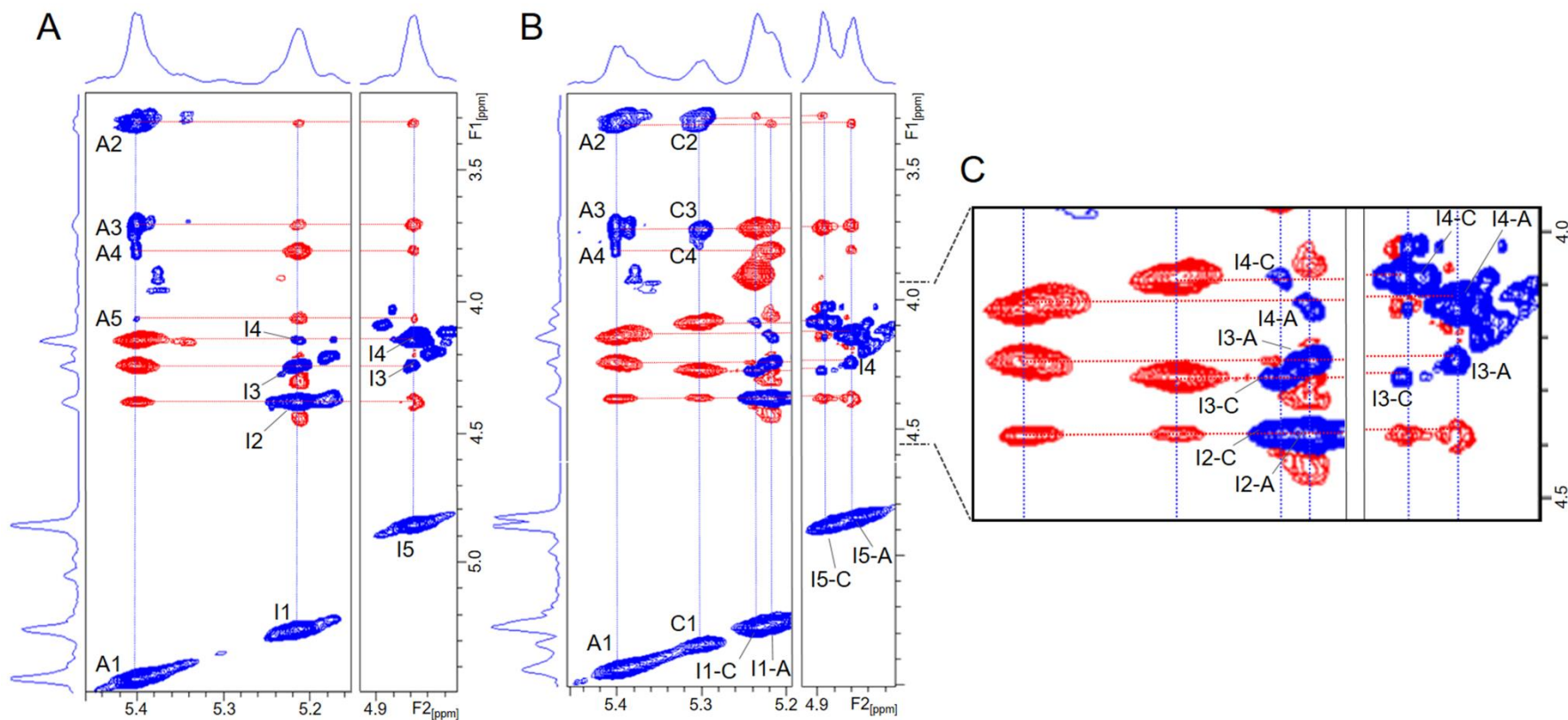


Figure S2. Phase-sensitive $^1\text{H} - ^1\text{H}$ TOCSY spectra of HB-L (A) and HB-I (B and C). These spectra discern signals of intra-residue spin systems from those of inter-residue ROEs. Saccharide connections were determined correlating the anti-phase (indicated in red and their connections by the horizontal dotted lines) and in-phase (shown in blue and their connections indicated by the vertical dotted lines) signals. The most intense anti-phase signal shows the linkage in the subsequent sugar residue. In the HB-L spectrum (A) *N*,6-disulfated α -glucosamine units (spin system A) connect with 2-sulfated α -iduronic acid residues (spin system I). A higher level of complexity can be observed in the spectrum of HB-I (B): residues of 6-desulfated α -glucosamine (spin system C) connect with the slightly downfield-shifted residues of 2-sulfated α -iduronic acid (namely I-C) while glucosamine *N*,6-disulfated (unit A) connects with the slightly upfield-shifted residues (namely I-A). These connections are amplified in the expansion shown in panel C. $^1\text{H} - ^1\text{H}$ NOESY spectra showed similar results but inter-residues signals are in-phase and thus not easily distinguishable from intra-residues signals (data not shown). Phase-sensitive $^1\text{H} - ^1\text{H}$ TOCSY spectra unambiguously demonstrate the origin of double signals in the spin system of 2-sulfated α -iduronic acid observed in the ^1H NMR spectrum of HB-I.

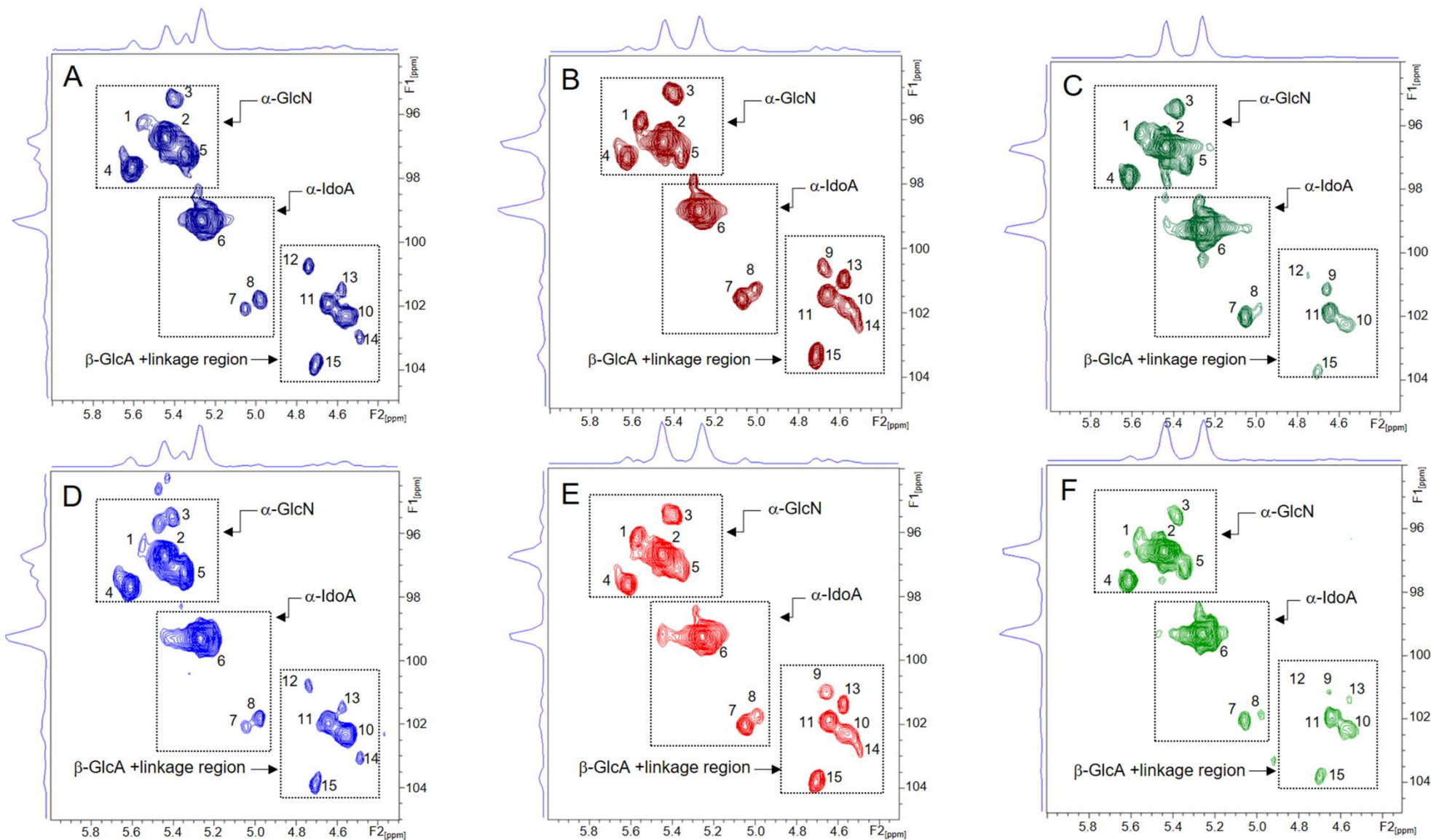


Figure S3. 2D ^1H - ^{13}C HSQC NMR spectra (strips of the anomeric region) of F3 derivatives from HB-I (A, in dark blue), HP-I (B, in dark red) and HB-L (C, in dark green) and their parental HB-I (D, in light blue), HP-I (E, in light red) and HB-L (F, in light green). Note the similarities among the spectra of F3 derivatives and their respective parental batches. See Table S3 for the chemical shifts and the legend for Fig. 3 for nomenclature of the signals and further details.