SUPPLEMENTARY METHODS

Microtiter dish biofilm formation assay

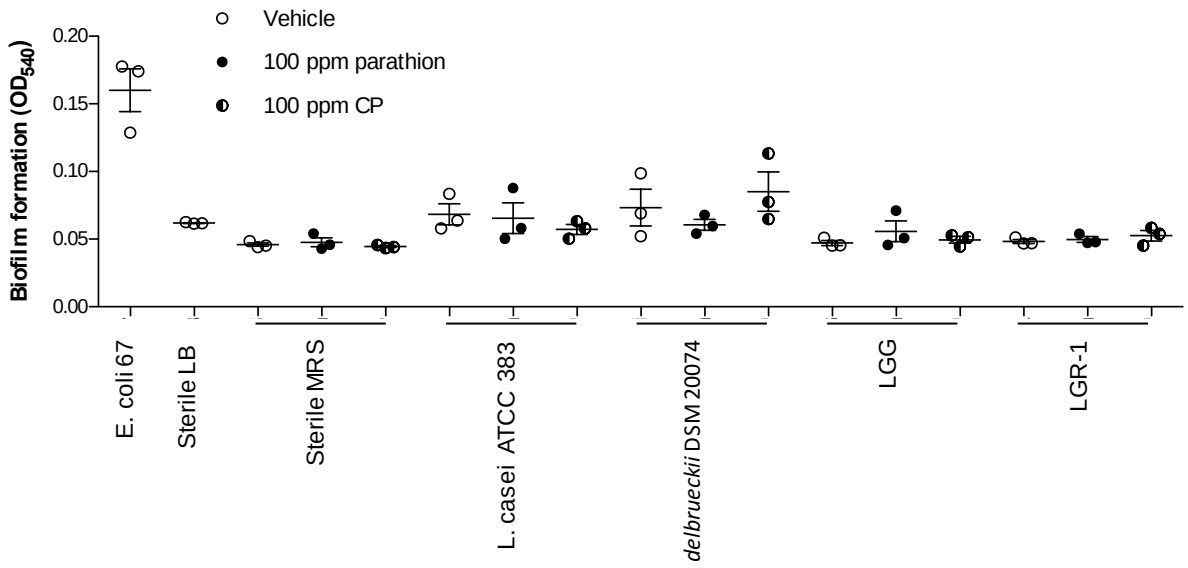
Bacterial cultures and organophosphate pesticide exposure for this assay were prepared as described in the pesticide tolerance assay for LGG, LGR-1, *L. casei* ATCC 393, and *L. delbrueckii* DSM 20074. Subcultures (1:100 dilution) of overnight (18 h) *Escherichia coli* 67—isolated from a woman with a urinary tract infection (1)—Luria-Bertani broth cultures were used as a positive control. Sterile broth was included as negative controls. 96-well plates were sealed with optically clear multi well plate sealing films or incubated aerobically at 37°C for lactobacilli and *E. coli*, respectively. The crystal violet microtiter dish biofilm formation assay was performed as described after 24 h incubation (2) and quantified using a wavelength of 540 nm with a Labsystems Multiskan Ascent microplate reader.

REFERENCES

1. **Reid G, Brooks JL**. 1984. *In vitro* attachment of *E. coli* to human epithelial cells. NZ Med J **97**:439–42.

2. **O'Toole GA**. 2011. Microtiter dish biofilm assay. JoVE **47**. http://www.jove.com/details.php?id=2437, doi: 10.3791/2437.

SUPPLEMENTARY FIGURE



Supplementary Figure 1. Lactobacilli do not demonstrate increased biofilm formation following organophosphate pesticide exposure.

Comparison of crystal violet-stained bacterial biofilms formed on 96-well polystyrene microtiter plates following 24 h incubation in MRS or LB broth for *E. coli* 67 and lactobacilli, respectively. The absorbance values at optical density 540 nm (OD_{540}) represent the biofilm formation capacity. Data are depicted as means ± SEM of 3 independent experiments consisting of 4-8 technical replicates. CP = chlorpyrifos, LGG = *Lactobacillus rhamnosus* GG, LGR-1 = *Lactobacillus rhamnosus* GR-1.