

Supplemental table S1 – Laxa et al.

Supplemental table I. Diurnal expression of the individual transformation events used to generate the data in Figure 2D, 3D and 5. The table summarizes the relative mRNA *gusA* transcript levels 1h and 7h after illumination. Additionally, the ratio between 1h and 7h indicates the decrease of *gusA* transcript throughout the day reflecting the endogenous diurnal behavior of *GGT1*.

Figure 2D			
Line	1h	7h	Ratio 1h:7h
<i>GGT1</i> -1	7381.42	1432.13	5.15
<i>GGT1</i> -2	1885.53	308.21	6.12
<i>GGT1</i> -3	4322.97	2828.71	1.53
<i>GGT1</i> -4	13327.16	3198.18	4.17
<i>GGT1</i> -5	2891.19	1608.25	1.80
<i>GGT1</i> -6	4661.64	3003.14	1.55
<i>GGT1</i> -7	5270.45	1015.82	5.19
<i>GGT1</i> -8	8419.68	5360.64	1.57
-1122-1	4904.38	866.04	5.66
-1122-2	10805.33	3390.72	3.19
-1122-3	7990.88	2537.05	3.15
-1122-4	16325.01	2997.57	5.45
-1122-5	9813.33	5983.80	1.64
-1122-6	3892.76	406.68	9.57
-1122-7	1521.92	283.72	5.36
-1122-8	12755.77	3330.24	3.83
-684-1	6079.50	1654.54	3.67
-684-2	1454.54	469.40	3.10
-684-3	2020.74	248.72	8.12
-684-4	8405.23	1531.47	5.49
-684-5	10981.16	4191.67	2.62
-684-6	8802.29	740.97	11.88
-684-7	4055.72	1539.70	2.63
-684-8	5777.80	2243.63	2.58
-684-9	1526.53	959.30	1.59
-684-10	11437.66	4054.88	2.82
-157-1	1232.47	689.70	1.79
-157-2	4613.22	1299.17	3.55
-157-3	559.30	77.97	7.17
-157-4	13605.56	1218.32	11.17
-157-5	2348.68	2191.23	1.07
-157-6	9564.45	4138.74	2.31
-157-7	11642.15	1477.81	7.88
-157-8	7452.24	1660.24	4.49
-157-9	9548.04	1259.44	7.58
-157-10	3094.02	1434.99	2.16

Figure 3D

Line	1h	7h	Ratio 1h:7h
GGT1-1	11250.99	2089.41	5.38
GGT1-2	6750.08	12545.02	0.54
GGT1-3	3071.11	799.25	3.84
GGT1-4	2019.54	181.70	11.11
GGT1-5	11887.34	2605.71	4.56
GGT1-6	1180.61	225.61	5.23
GGT1-7	15726.83	4111.26	3.83
GGT1-8	16228.31	2812.77	5.77
GGT1-9	4793.84	2962.11	1.62
GGT1-Δ5I-1	9808.88	2069.69	4.74
GGT1-Δ5I-2	2.70	16.07	0.17
GGT1-Δ5I-3	0.00	0.00	
GGT1-Δ5I-4	1.56	0.22	7.18
GGT1-Δ5I-5	2491.90	664.83	3.75
GGT1-Δ5I-6	2.85	0.00	
GGT1-Δ5I-7	3.48	1.18	2.95
GGT1-Δ5I-8	0.44	0.75	0.58
GGT1-Δ5I-9	1849.38	659.05	2.81

Figure 5A and B

Line	1h	7h	Ratio 1h:7h
GGT1-1	21211.40	9939.05	2.13
GGT1-2	2281.58	3004.49	0.76
GGT1-3	10838.56	8599.98	1.26
GGT1-4	6176.47	1244.05	4.96
GGT1-5	11070.28	3023.53	3.66
GGT1-6	8140.45	998.59	8.15
GGT1-7	2507.03	1390.08	1.80
GGT1-8	5985.21	1182.72	5.06
GGT1-9	3437.55	479.01	7.18
GGT1-10	1001.86	1425.96	0.70
GGT1-Δ5I-1	6084.68	2363.96	2.57
GGT1-Δ5I-2	7983.21	4678.77	1.71
GGT1-Δ5I-3	7475.36	3760.27	1.99
GGT1-Δ5I-4	297.21	619.51	0.48
GGT1-Δ5I-5	951.95	512.43	1.86
GGT1-Δ5I-6	334.09	174.80	1.91
GGT1-Δ5I-7	3786.31	2521.27	1.50
GGT1-Δ5I-8	5001.94	307.91	16.24
GGT1-Δ5I-9	1061.36	169.21	6.27
GGT1-Δ5I-10	2302.04	459.51	5.01
GGT2-1	63.35	125.83	0.50
GGT2-2	171.72	170.85	1.01
GGT2-3	5.87	11.44	0.51
GGT2-4	84.71	160.76	0.53
GGT2-5	17.31	17.58	0.98

<i>GGT2-7</i>	7.69	7.44	1.03
<i>GGT2-8</i>	7.73	9.67	0.80
<i>GGT2-9</i>	18.63	15.79	1.18
<i>GGT2-10</i>	22.83	52.40	0.44
<i>GGT2-Δ5I-1</i>	3.16	1.35	2.35
<i>GGT2-Δ5I-3</i>	4.05	8.24	0.49
<i>GGT2-Δ5I-4</i>	13.27	7.80	1.70
<i>GGT2-Δ5I-5</i>	1.26	3.04	0.41
<i>GGT2-Δ5I-6</i>	23.13	14.40	1.61
<i>GGT2-Δ5I-7</i>	6.35	18.51	0.34
<i>GGT2-Δ5I-8</i>	1.62	8.44	0.19
<i>GGT2-Δ5I-9</i>	1.30	1.48	0.88
<i>GGT1 25I-1</i>	340.60	677.23	0.50
<i>GGT1 25I-2</i>	1571.02	1984.99	0.79
<i>GGT1 25I-3</i>	4955.78	1031.92	4.80
<i>GGT1 25I-4</i>	6462.45	9037.18	0.72
<i>GGT1 25I-5</i>	3549.35	329.69	10.77
<i>GGT1 25I-6</i>	6312.42	4856.22	1.30
<i>GGT1 25I-7</i>	21740.90	4332.06	5.02
<i>GGT1 25I-8</i>	10417.43	5810.84	1.79
<i>GGT1 25I-9</i>	15544.19	9515.55	1.63
<i>GGT1 25I-10</i>	14192.08	669.05	21.21
<i>GGT2 15I-1</i>	876.64	254.51	3.44
<i>GGT2 15I-2</i>	349.72	299.93	1.17
<i>GGT2 15I-3</i>	221.89	165.72	1.34
<i>GGT2 15I-4</i>	189.68	1004.09	0.19
<i>GGT2 15I-5</i>	245.85	312.92	0.79
<i>GGT2 15I-6</i>	635.40	42.85	14.83
<i>GGT2 15I-7</i>	482.73	1609.62	0.30
<i>GGT2 15I-8</i>	571.94	451.72	1.27

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Supplemental table II. List of primers used for quantitative real-time PCR analysis. For clarity the AGI code of *GAPDH* is given. P1 designates the position 146 bp downstream of ATG on the *gusA* coding sequence used for qPCR analysis after chromatin immunoprecipitation (ChIP). Fw: forward primer, rv: reverse primer, SF: splice form.

Primer	Sequence (5' to 3')
<i>GAPDH</i> _mRNA_fw (At1g13440)	TTGGTGACAACAGGTCAAGCA
<i>GAPDH</i> _mRNA_rv	AAACTTGTCGCTCAATGCAATC
<i>GGT1</i> _unspliced RNA_fw	CGGTAATGGAGATGGGGTAA
<i>GGT1</i> _unspliced _rv	AGAATCTGCCAAACGGAAA
<i>GGT2</i> _SF1/2_fw	CTGCTCTGGAACGTGACTTTG
<i>GGT2</i> _SF2_fw	CGCTATTGCAATAAGCGCCC
<i>GGT2</i> _SF3_fw	CAATCTTTATGATCGACTCAATAAG
<i>GGT2</i> _SF4_fw	TATCGTTTGAGACTGTTCTGTAG
<i>GGT2</i> _5'UTR_rv	CTAAAAGCAAATGATTCCTCAC
<i>GusA</i> _ChIP_fw (P1)	CTGTGGAATTGGTCAGCGTTG
<i>GusA</i> _ChIP_rv (P1)	CAGACGTTGCCCGCATAATTAC
<i>GusA</i> _unspliced RNA_fw	GTTGATGTGCAGGTATCACCG
<i>GusA</i> _unspliced RNA_rv	TTGCCGTTTTTCGTCCGGTAATC
<i>GusA</i> _mRNA_fw	GAAGCCGATGTCACGCCG
<i>GusA</i> _mRNA_rv	TTGCCGTTTTTCGTCCGGTAATC
<i>NptII</i> _mRNA_fw	CTGTCCGGTGCCCTGAATG
<i>NptII</i> _mRNA_rv	CAACGTCCGAGCACAGCTGC

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Supplemental table III. Primer used for the amplification of the *promoter::gusA* constructs used in this study. Besides the primer name and nucleotide sequences the table includes information on the relative position of the primer to the transcription initiation start (TIS), specific numbering for the overlap extension (OE) PCR (P1-P6; scheme available in Supplemental figure S10). Underlined nucleotide sequences (solid line) mark overlapping regions that were added for the OE PCR to generate of both *GGT1* 25I and *GGT2* 15I, respectively. Underlined nucleotide sequences (dashed line) mark the multiple cloning site introduced downstream of the 5'UTRs of *CAT3* (At1g26020), *GLDP1-P2* (At4g33010, Adwy et al., 2015) and a root-specific peroxidase (*PRX*, At3g01190). Underlined nucleotide sequences (dotted line) mark the restriction sites that have been introduced 5' and 3' of the *GGT1* 5'UTR intron, respectively. Fw: forward primer, rv: reverse primer.

Position relative to TIS	Primer	Sequence (5' to 3')
	35S_fw	CACCGGTCCAAGACCAGAGGGC
	35S_rv	TGTCCTCTCAAATGAAATGAAC
-1458	<i>GGT1</i> _-1458_fw	CACCTAAAGCATTATTCAGTAGTATGTG
-1122	<i>GGT1</i> _-1122_fw	CACCCGGCTGAGAACTTGCATGAA
-684	<i>GGT1</i> _-684_fw	CACCCGGACAATAAATCGCTACAGAC
-157	<i>GGT1</i> _-157_fw	CACCCCTATTGGTCATGTTCCAAACG
	<i>GGT1</i> _5'UTR_rv	TTATGTTCACTCTGAACCAAT
	<i>GGT1</i> _Δ5I_rv	TTATGTTCACTCTGAACCAATCCCTAGCTCACTTCTCCCTC CTCTTCAAACCTCAATGTTGTGACTC
-347	<i>GGT2</i> _-347_fw	CACCATTTTGCACATTCATGTTACTTAAT
	<i>GGT2</i> _5'UTR_rv	TTCCAGGATTTACTTTTCAAAGC
	<i>GGT2</i> _Δ5I_rv	TTCCAGGATTTACTTTTCAAAGCTAAAAGCAAAATGATTCCTCA CTTCCCCCTTAGATCAAACCTCACGACAC
Overlap extension primer <i>Ggt1</i> 25I	Primer	Sequence (5' to 3')
P1	<i>GGT1</i> _-1458_fw	CACCTAAAGCATTATTCAGTAGTATGTG
P6	<i>GGT1</i> _5'UTR_rv	TTATGTTCACTCTGAACCAAT
P2	<i>GGT1</i> _P2_rv	<u>AGAAATTATACCAACCTTCAAACCTCAATGTTGTGACTC</u>
P3	<i>GGT2</i> _5I_fw	<u>ACATTGAGTTTGAAGGTTGGTATAATTTCTGTAGTTTTG</u>
P4	<i>GGT2</i> _5I_rv	<u>CACTTCTTCCCTCCTTCTACAACCAAAATAAGCAGATG</u>
P5	<i>GGT1</i> _P5_fw	<u>TATTTTGGTTGTAGAGGAGGGAAGAAGTGAGCTAGGGATTGGT</u> TCAGAGTGAACATAACCTAAGGGTGGGCGC

P5	<i>GGT1_P5_rv</i>	GCGCCCACCCTTAGGTTATGTTCACTCTGAACCAATCCCTAGC TCACTTCTTCCCTCCTT <u>CTACAACCAAAATA</u>
Overlap extension primer <i>Ggt2</i> 15l	Primer	Sequence (5' to 3')
P1	<i>GGT2_-347_fw</i>	CACCATTTTGCACATTCATGTTACTTAAT
P6	<i>GGT2_5'UTR_rv</i>	TTCCAGGATTTACTTTTCAAAGC
P2	<i>GGT2_P2_rv</i>	<u>AACCAAATCACAAAC</u> CTTAGATCAACTCACGACACA
P3	<i>GGT1_5l_fw</i>	<u>GTGAGTTGATCTAAG</u> GTTTGTGATTTGGTTTTCTTTCTC
P4	<i>GGT1_5l_rv</i>	<u>ATTCCTCACTTCCCC</u> CTACAATTGGAAGAAAACAATTCATC
P5	<i>GGT2_P5_fw</i>	<u>TTCTCCAATTGTAGAGGGGAAGTGAGGAATC</u> ATTTTGCTTTTA GCTTTGAAAAGTAAATCCTGGAACCTAAGGGTGGGCGC
P5	<i>GGT2_P5_rv</i>	GCGCCCACCCTTAGGTTCCAGGATTTACTTTTCAAAGCTAAAA GCAAAATGATTCCTCACTT <u>CCCCCTACAATTGGAAGAA</u>
Promoter 15l constructs	Primer	Sequence (5' to 3')
	<i>GGT1</i> 5l <i>Acc65l_fw</i>	<u>AAAAAGGTACC</u> GTTTGTGATTTGGTTTTCTTTC
	<i>GGT1</i> 5l <i>BglII_rv</i>	<u>AAAAAAGATCT</u> CTACAATTGGAAGAAAACAATTC
	<i>GLDP1-P2_fw</i>	CACCTATGTCCCATTAGAGGGGAA
	<i>GLDP1_MCS_rv</i>	<u>AGGCCTGTCAGTAGATCTGTCAGTGGTACC</u> TGGGAAAAAAGGT TGCAGTC
	<i>CAT3_fw</i>	CACCCGAGAATATTATCACACAAC
	<i>CAT3_rv</i>	GGTGATGATAGAAGGTTGATG
	<i>CAT3_MCS_rv</i>	<u>GTCGACGTCAGTGGTACC</u> GGTGATGATAGAAGGTTGATG
	<i>PRX_fw</i>	CACCTCTATTTTACTCTTTCTATTCCG
	<i>PRX_rv</i>	TTTCTTAAAAAATCTTTAGTTTGTTC
	<i>PRX_MCS_rv</i>	<u>GTCGACGTCAGTGGTACC</u> TTTCTTAAAAAATCTTTAGTTTGTTC C

Supplemental table S4 – Laxa et al.

Supplemental table IV. Segregation analysis of *GGT1* $\Delta 5l::gusA$ transgenic lines (F2-generation) used to generate the data in Figure 4 and 5. The table summarizes the number of individuals analyzed per transgenic event, the number of kanamycin-sensitive and insensitive plants as well as the expected numbers according to mendelian segregation. Additionally, transcript levels of *gusA* mRNA (standardized to *GAPDH*) were added to correlate *gusA* expression to the probability of a single insertion indicated by the χ^2 -test.

Event	Individuals	insensitive	sensitive	expected insensitive	expected sensitive	<i>gusA</i> mRNA/ <i>GAPDH</i>	χ^2 -test
<i>GGT1</i> $\Delta 5l$ -1	49	39	10	36.75	12.25	0.299	0.46
<i>GGT1</i> $\Delta 5l$ -2	50	46	4	37.5	12.5	0.814	0.01
<i>GGT1</i> $\Delta 5l$ -3	49	40	9	36.75	12.25	0.764	0.28
<i>GGT1</i> $\Delta 5l$ -4	47	39	8	35.25	11.75	0.026	0.21
<i>GGT1</i> $\Delta 5l$ -5	51	35	16	38.25	12.75	0.067	0.29
<i>GGT1</i> $\Delta 5l$ -6	50	33	17	37.5	12.5	0.046	0.14
<i>GGT1</i> $\Delta 5l$ -7	51	40	11	38.25	12.75	0.493	0.57
<i>GGT1</i> $\Delta 5l$ -8	53	38	15	39.75	13.25	0.264	0.58
<i>GGT1</i> $\Delta 5l$ -9	50	36	14	37.5	12.5	0.044	0.62