## Protein Kinase D1 regulates focal adhesion dynamics and cell adhesion through Phosphatidylinositol-4phosphate 5-kinase type-l γ

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Supplemental Figure S1, relates to Figure 1: **Cellular localization of endogenous PKD1 and pY397-FAK, and co-immunoprecipitation of ectopically expressed PKD1 and FAK. A, B:** MCF-7 cells (0.10 x 10<sup>6</sup> cells /well, 24-well plate) were seeded on fibronectin-coated (2 µg/ml) glass coverslips and serum-starved for 16 hrs. Following serum-starvation, cells were stimulated with 10 µg/ml fibronectin or left untreated. After 30 minutes, cells were fixed and the localization of endogenous PKD1(Everest antibody in A; Abnova antibody in **B**) and pY397-FAK was determined by immunofluorescence analysis. Scale bars indicate 10 µm. Insets are 2.4-fold enhanced. **C, D:** Hek293T cells (0.5 x 10<sup>6</sup> cells/well, 6 well plate) in **C**, or NIH-3T3 cells (2 x 10<sup>6</sup> cells/10 cm dish) in **D**, were transfected with mCherry-FAK and vector control, FLAG-tagged wildtype PKD1 (PKD1) or FLAG-tagged constitutively-active PKD1 (PKD1.CA) as indicated. PKD1 was immunoprecipitated (anti-FLAG) and immunoprecipitates were analyzed for co-immunoprecipitated FAK. Samples were re-probed for overexpressed PKD1. Control Western blots of lysates were probed for FAK and β-actin expression.

Supplemental Figure S2, relates to Figure 2



Supplemental Figure S2, relates to Figure 2: **PIP5KIγ localizes to focal adhesions and is phosphorylated by PKD1 at S448. A:** NIH-3T3 cells were co-transfected with mCherry-Paxillin and GFP-PIP5KI**γ**. Scale bars indicate 10 µm. The inset in is 6.5-fold enhanced on the right. **B:** HeLa cells (0.65 x 10<sup>6</sup> cells/6 cm dish) were co-transfected with vector and FLAG-tagged constitutively-active PKD1 (PKD1.CA) and vector, HA-tagged PIP5KI**γ** or PIP5KI**γ**.S448A as indicated. PIP5KI**γ** was immunoprecipitated (anti-HA) and assessed for PKD1-mediated phosphorylation using the pMOTIF antibody which recognizes phosphorylated PKD substrates. Immunoprecipitates were re-probed with anti-HA for equal loading of PIP5KI**γ** and PIP5KI**γ**.S448A. Lysates were evaluated for PKD1.CA expression (anti-FLAG). **C:** HeLa cells (0.25 x 10<sup>6</sup> cells/well, 6 well plate) were co-transfected with vector control or FLAG-tagged constitutively-active PKD1 (PKD1.CA) and vector, HA-tagged PIP5KI**γ** or PIP5KI**γ**.S448A, as indicated. PIP5KI**γ** was immunoprecipitated (anti-HA) and assessed for PKD1-mediated phosphorylation using a pS448-PIP5KI**γ** antibody. Immunoprecipitates were re-probed with anti-HA for equal loading of PIP5KI**γ**.S448A, as indicated. PIP5KI**γ** was immunoprecipitated (anti-HA) and assessed for PKD1-mediated phosphorylation using a pS448-PIP5KI**γ** antibody. Immunoprecipitates were re-probed with anti-HA for equal loading of PIP5KI**γ** and PIP5KI**γ**.S448A. The lysates were evaluated for PKD1 expression (anti-PKD1).

## Supplemental Figure S3, relates to Figure 2



Supplemental Figure S3, relates to Figure 2: **PIP5KI** $\gamma$  is mainly phosphorylated by PKD1. A: Hek293T cells (0.6 x 10<sup>6</sup> cells/well, 6 well plate) were transfected with HA-tagged PIP5KI $\gamma$ , and stimulated with PMA (500 nM, 15 min), CRT0066101 (2.5  $\mu$ M, 60 min) or DMSO control as indicated. Cell lysates were probed for PIP5KI $\gamma$  phosphorylation (anti-pS448-PIP5KI $\gamma$ ) and reprobed for total PIP5KI $\gamma$  (anti-HA). Control blots were probed for  $\beta$ -actin. **B**: Hek293T cells (0.5 x 10<sup>6</sup> cells/well, 6 well plate) were co-transfected with HA-tagged PIP5KI $\gamma$  along with vector control, FLAG-tagged constitutively-active PKD1 (PKD1.CA), FLAG-tagged constitutively-active PKD2 (PKD1.CA), or GST-tagged constitutively-active PKD3 (PKD3.CA). An anti-HA antibody was used to immunoprecipitate PIP5KI $\gamma$  from cells. Immunoprecipitates were subjected to SDS PAGE, and assessed using a pS448-PIP5KI $\gamma$  antibody. Samples were counterstained for total PIP5KI $\gamma$  using an anti-HA antibody. Control blots were performed for PKD1/2 (anti-FLAG), PKD3 (anti-GST) and  $\beta$ -actin expression.

Supplemental Figure S4, relates to Figure 3



Supplemental Figure S4, relates to Figure 3: **Cellular localization of PIP5Kly and PIP5Kly.S448A. A:** Controls for Figure 3A. NIH-3T3 cells expressing HA-tagged PIP5Kly or PIP5Kly.S448A or GFP-tagged active PKD1 (GFP-PKD1.CA) were seeded ( $0.05 \times 10^6$  cells/well, 24 well plate) on fibronectin-coated (2 µg/ml) glass coverslips. Following fixation, immunofluorescence analysis was used to determine the localization of PIP5Kly and GFP-PKD1. Scale bars indicate 10 µm. **B:** NIH-3T3 cells co-expressing GFP-PIP5Kly or GFP-PIP5Kly.S448A and RFP-LifeACT (to label F-actin structures) were seeded on fibronectin-coated (2 µg/ml) glass coverslips ( $0.05 \times 10^6$  cells/well, 24 well plate). Following fixation, cells were analyzed by immunofluorescence. Scale bars indicate 10 µm.

Supplemental Figure S5, relates to Figure 3



Expression controls:



Supplemental Figure S5, relates to Figure 3: **S448 phosphorylation inhibits interaction of PIP5K1y and Talin. A-D**: NIH-3T3 cells expressing mCherry-PKD1.CA, GFP-Talin with HA-tagged PIP5Kly or PIP5Kly.S448A were seeded on fibronectin-coated (2  $\mu$ g/ml) glass coverslips. Localization of Talin and PIP5Kly was determined by immunofluorescence analysis. Scale bars indicate 10  $\mu$ m. Insets are 2.6-fold enhanced.

## Supplemental Figure S6, relates to Figure 4



Supplemental Figure S6, relates to Figure 4: **PKD1-mediated phosphorylation of S448 occurs in response to Fibronectin-RhoA signaling. A:** HeLa cells (0.25 x 10<sup>6</sup> cells/well, 6 well plate) were cotransfected with HA-tagged PIP5Kly, vector or FLAG-tagged PKD1 and vector or GST-tagged active RhoA (RhoA.CA), as indicated. PIP5Kly was immunoprecipitated (anti-HA) and analyzed using pS448-PIP5Kly antibody. Samples were re-probed with anti-HA to determine PIP5Kly loading. Control blots were probed for PKD1, RhoA.CA (anti-GST) and  $\beta$ -actin expression. **B:** HeLa cells (0.25 x 10<sup>6</sup> cells/well, 6 well plate) were co-transfected with HA-tagged PIP5Kly and vector control or GST-tagged active RhoA (RhoA.CA). Cells were stimulated with CRT0066101 (500 nM, 16 hours) or left untreated as indicated. PIP5Kly was immunoprecipitated (anti-HA) and analyzed using the pS448- PIP5Kly antibody. Immunoprecipitates were re-probed for total PIP5Kly (anti-HA). Control blots were probed for RhoA.CA (anti-GST) and  $\beta$ -actin expression.

## Supplemental Figure S7, relates to Figure 5



Supplemental Figure S7, relates to Figure 5: **Attachment of NIH-3T3 cells to fibronectin. A:** NIH-3T3 cells (0.05x 10<sup>6</sup> cells/well, 24 well plate) were plated on non-coated or fibronectin-coated (10 µg/ml) glass coverslips. Bright field images were acquired at the times indicated to determine the number of cells attached. The bar indicates 50 µm. **B:** Control blots for Figures 5C and 5D. Cell lysates were evaluated by Western blotting for the expression of PIP5Kly (anti-GFP), PKD1 and  $\beta$ -actin.