

# A Parameterization of Cholesterol for Mixed Lipid Bilayer Simulation within the Amber Lipid14 Force Field

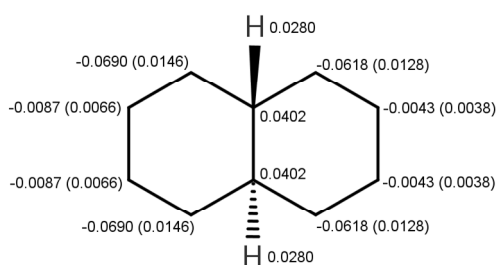
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## PARAMETERIZATION

**Sterol Ring Parameters.** Because the majority of the cholesterol is based on the basic sterol unit and is largely composed of *sp*<sup>3</sup> carbon atoms, parameters of the sterol carbons and hydrogens are important to overall lipid bilayer structure. Within the time scales of an all-atom lipid bilayer and cholesterol MD simulation it is likely that the sterol portion of the molecule will mostly be inflexible due to the ring arrangement of the sterol.<sup>1,2</sup> However, the van der Waals of this region can influence important bilayer structural properties such as head group packing as well as volume of the bilayer.<sup>1,2</sup> Therefore, the van der Waals parameters of the carbons and hydrogens in this region were re-examined.

The atom types of most sterol carbons in cholesterol are basic *sp*<sup>3</sup> atom types adapted from GAFF.<sup>3</sup> The hydrogen atom types from GAFF are hydrogens bonded to an aliphatic carbon without an electron withdrawing group.<sup>3</sup> The initial parameters for carbon include a van der Waals radius of 1.908 Å and a well-depth of 0.1094 kcal/mol. Hydrogens have a radius 1.4870 Å and a well-depth of 0.0157 kcal/mol. In order to examine a similar system and isolate these parameters, a small ringed molecule was chosen: decalin also known as decahydronaphtalene. The relevant *trans* conformation of decalin was modeled and simulated with MD.



**Figure S1.** Decahydronaphtalene (*trans*-decalin) partial charges. Decalin was used as a sterol model for study of van der Waals parameters. All carbons are adapted from the GAFF *sp*<sup>3</sup> carbons (Lipid14 *cA*) and all hydrogens are adapted from the GAFF hydrogens bonded to an aliphatic carbon without an electron withdrawing group (Lipid14 *hA*).<sup>3,4</sup>

Figure S1 shows the structure and partial atomic charges of *trans*-decalin used for the gas and liquid phase simulations. GAFF parameters were applied to decalin, with the intent of evaluating the carbon and hydrogen parameters. Partial charges were derived for decalin using the RESP method described previously.<sup>4</sup> One structure was optimized and ESP was calculated at the HF/6-31G\* level. Atomic partial charges were fit in two stages in the normal RESP method.<sup>4</sup> Topology and parameters were assembled with the Amber program *Leap*.<sup>5</sup>

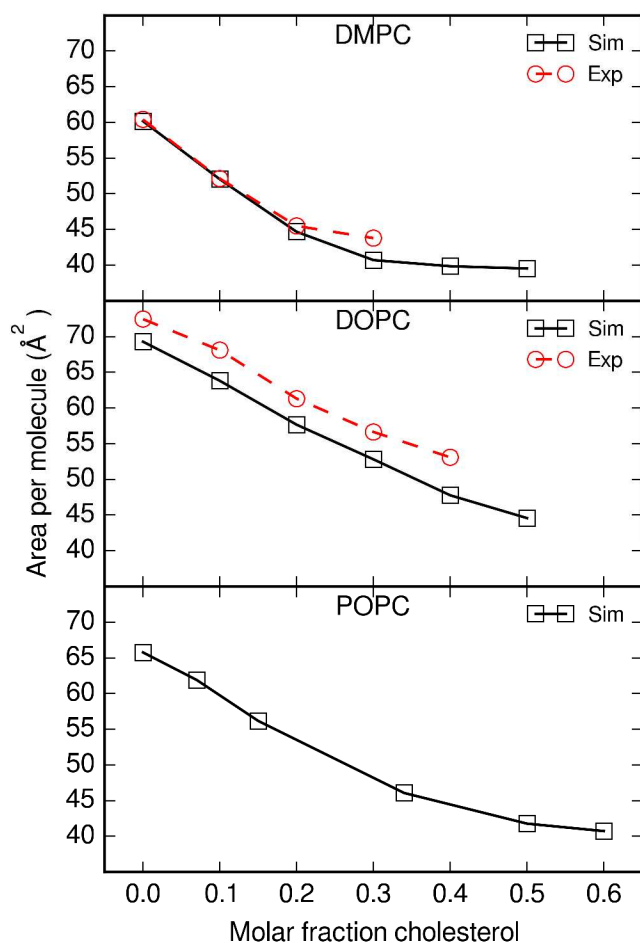
**Table S1.** Partial atomic charges for hexadecane

Atom name	Charge	Atom name	Charge
C	-0.0991	H, H1, H2	0.0214
C1	0.0220	H3, H4	0.0059
C2	-0.0248	H5, H6	0.0147
C3	-0.0336	H7, H8	0.0130
C4	-0.0214	H9, H10	0.0124
C5	-0.0192	H11, H12	0.0093
C6	-0.0215	H13, H14	0.0124
C7	-0.0244	H15, H16	0.0136
C8	-0.0310	H17, H18	0.0133
C9	-0.0315	H19, H20	0.0148
C10	-0.0280	H21, H22	0.0171
C11	-0.0395	H23, H24	0.0176
C12	-0.0384	H25, H26	0.0175
C13	-0.0341	H27, H28	0.0195
C14	0.0193	H29, H30	0.0095
C15	-0.1210	H31, H32, H33	0.0269

Hexadecane was also simulated in mixtures of *trans*-decalin. Parameters were taken directly from Lipid14 including the updated van der Waals and dihedrals terms. Partial charges were derived using the Lipid14 RESP methodology with multiple conformations.

## VALIDATION

**Phospholipid and Cholesterol Area per Molecule.** Another experimental comparison is the lateral area per molecule for cholesterol and lipid bilayers. Usually the experimental area per molecule is derived from volume and thickness bilayer measurements.<sup>6</sup> While it is possible to calculate individual area per molecules for a binary bilayer,<sup>7</sup> the simple average area per molecule  $A_M$  for both cholesterol and lipid is reported here, where  $A_M = A_{XY}/(n_L + n_{CHL})$ . The lateral area is denoted by  $A_{XY}$  and the number of lipids and cholesterol is  $n_L$  and  $n_{CHL}$ , respectively.



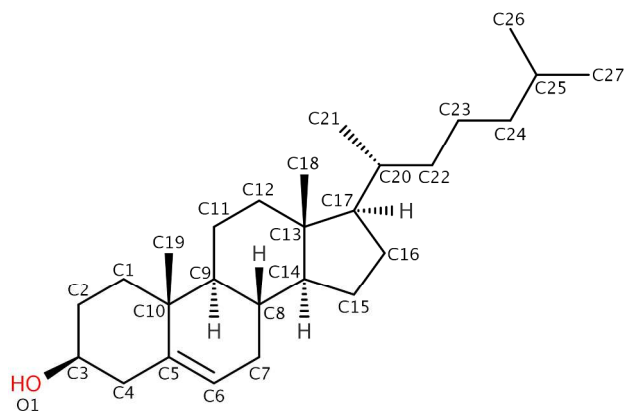
**Figure S2.** Average area per molecule for lipid and cholesterol bilayers. Boxes are the average simulation area per molecule. Circles are the experimental area per average molecule at 30 C from Pan et al.<sup>6</sup>

Figure S2 shows the average area per molecule for cholesterol and lipid bilayers. Increasing cholesterol content correlates to decreasing area per molecule in both simulation and experiment. This condensing effect is well known in bilayers with cholesterol.<sup>8,9</sup> Simulations with other cholesterol force fields result in bilayers with low area per molecule: Jambeck et al reports an area of  $57 \pm 0.5 \text{ \AA}^2$  and  $43 \pm 0.4 \text{ \AA}^2$  for 20% and 50% cholesterol in a DOPC bilayer, respectively.

Cholesterol also has an experimentally observed saturation point when mixed with lipid bilayers.<sup>10,11</sup> Beyond saturation, cholesterol precipitates from the bilayer as cholesterol monohydrate. Huang et al report a cholesterol saturation point in phosphatidylcholine bilayers to be 66%.<sup>11</sup> Hung et al report a lower saturation point: ~44% in DMPC bilayers and ~40% in DOPC bilayers.<sup>10</sup> More recently, Mainali et al show that cholesterol bilayer domains form in DMPC bilayers at 50% molar fraction cholesterol and form cholesterol crystals at 66% molar fraction cholesterol.<sup>12</sup> It is possible that the simulations presented here approach the saturation points of these bilayers.

## PARAMETERS

**Lipid14 Cholesterol Parameters.** Please see <http://ambermd.org> for specific implementation details for AMBER. Please see <http://ambermd.org/formats> for the specification for the AMBER force field parameter and topology files.



**Figure S3.** Atom names for cholesterol. The Lipid14 residue name is CHL. Hydrogens follow the Lipid11 naming convention. Namely, hydrogens follow the format “HXY” where X is the carbon number and Y is 1, 2, or 3 for each attached hydrogen. All atom names are also listed in table S2.

**Table S2.** Lipid14 cholesterol atom names, types, and charges. Bonded hydrogens are displayed in the same row as their corresponding heavy atom.

<b>Name</b>	<b>Type</b>	<b>Charge</b>	<b>Name</b>	<b>Type</b>	<b>Charge</b>
C1	cA	-0.0317	H11, H12	hA	0.0094
C2	hA	-0.0881	H21, H22	hA	0.0446
C3	hE	0.2936	H31	hA	0.0296
C4	cA	-0.1622	H41, H42	hA	0.0900
C5	cB	-0.1395			
C6	cB	-0.2082	H61	hB	0.1228
C7	cA	-0.0699	H71	hA	0.0479
C8	cA	-0.0114	H81	hA	0.0729
C9	cA	0.0196	H91	hA	0.0293
C10	cA	0.0791			
C11	cA	-0.0664	H111, H112	hA	0.0301
C12	cA	-0.0650	H121, H122	hA	0.0104
C13	cA	0.0574			
C14	cA	0.0058	H141	hA	0.0316
C15	cA	-0.1020	H151, H152	hA	0.0265
C16	cA	-0.0921	H161, H162	hA	0.0354
C17	cA	0.0324	H171	hA	0.0166
C18	cA	-0.1150	H181, H182, H183	hA	0.0245
C19	cA	-0.1081	H191, H192, H193	hA	0.0340
C20	cD	0.0443	H201	hL	0.0210
C21	cD	-0.1546	H211, H212, H213	hL	0.0363
C22	cD	-0.0390	H221, H222	hL	0.0084
C23	cD	-0.0285	H231, H232	hL	0.0157
C24	cD	-0.1256	H241, H242	hL	0.0401
C25	cD	0.2124	H251	hL	-0.0025
C26	cD	-0.2578	H261, H262, H263	hL	0.0580
C27	cD	-0.2578	H271, H272, H273	hL	0.0580
O1	oH	-0.7030	HO1	hO	0.4148

**Lipid14 Cholesterol Parameter File.** This is the cholesterol parameter file (*CHL.frcmod*) containing parameters compatible with

Lipid14 parameters included in AmberTools 14 (*leaprc.lipid14*, *lipid14.lib*, *lipid14.dat*).

CHL Lipid14 frcmod

MASS

hA 1.008		H bonded to aliphatic carbon without electrwd. group (GAFF hc-)
hO 1.008		H in Hydroxyl group (GAFF ho-)
oH 16.00		sp3 oxygen in hydroxyl group (GAFF oh-)

BOND

CA-CB	328.3	1.5080	Lipid11 v1.0 (GAFF c3-c2)
CA-cD	303.1	1.5350	Lipid14 CHL (GAFF c3-c3)
CA-hA	337.3	1.0920	Lipid11 v1.0 (GAFF c3-hc)
CA-oH	314.1	1.4260	Lipid11 v1.0 (GAFF c3-oh)
hO-oH	369.6	0.9740	Lipid11 v1.0 (GAFF ho-oh)

ANGLE

CA-CA-CB	63.530	111.440	Lipid11 v1.0 (GAFF c3-c3-c2)
CA-CA-cD	63.210	110.630	Lipid14 CHL (GAFF c3-c3-c3)
CA-CA-hA	46.370	110.050	Lipid11 v1.0 (GAFF c3-c3-hc)
CA-CA-oH	67.720	109.430	Lipid11 v1.0 (GAFF c3-c3-oh)
CA-CB-CA	62.700	116.520	Lipid11 v1.0 (GAFF c3-c2-c3)
CA-CB-cB	64.330	123.420	Lipid11 v1.0 (GAFF c3-c2-c2)
CA-CB-hB	45.660	117.300	Lipid11 v1.0 (GAFF c3-c2-ha)
CA-cD-hL	46.370	110.050	Lipid14 CHL (GAFF c3-c3-hc)
CA-cD-cD	63.210	110.630	Lipid14 CHL (GAFF c3-c3-c3)
CA-oH-hO	47.090	108.160	Lipid11 v1.0 (GAFF c3-oh-ho)
CB-CA-hA	47.030	110.490	Lipid11 v1.0 (GAFF c2-c3-hc)
cD-CA-hA	46.370	110.050	Lipid14 CHL (GAFF c3-c3-hc)
hA-CA-hA	39.430	108.350	Lipid11 v1.0 (GAFF hc-c3-hc)
hE-CA-oH	50.970	109.880	Lipid11 v1.0 (GAFF ha-c3-oh)

DIHE

CA-CA-CA-CA	1	0.18	0.0	-3.000	SCEE=1.2	SCNB=2.0	Lipid11 v1.0 (GAFF c3-c3-c3-c3)
CA-CA-CA-CA	1	0.25	180.0	-2.000	SCEE=1.2	SCNB=2.0	Lipid11 v1.0 (GAFF c3-c3-c3-c3)
CA-CA-CA-CA	1	0.20	180.0	1.000	SCEE=1.2	SCNB=2.0	Lipid11 v1.0 (GAFF c3-c3-c3-c3)
CA-CA-CA-cB	1	0.1556	0.0	3.000	SCEE=1.2	SCNB=2.0	Lipid11 v1.0 (GAFF X -c3-c3-X )
CA-CA-CA-cD	1	0.3112	180.000	1.000	SCEE=1.2	SCNB=2.0	Lipid14 CHL (Lipid14 cD-cD-cD-cD)
CA-CA-CA-cD	1	-0.1233	180.000	2.000	SCEE=1.2	SCNB=2.0	Lipid14 CHL (Lipid14 cD-cD-cD-cD)
CA-CA-CA-cD	1	0.1149	0.000	3.000	SCEE=1.2	SCNB=2.0	Lipid14 CHL (Lipid14 cD-cD-cD-cD)
CA-CA-CA-cD	1	-0.2199	0.000	4.000	SCEE=1.2	SCNB=2.0	Lipid14 CHL (Lipid14 cD-cD-cD-cD)
CA-CA-CA-cD	1	0.2170	0.000	5.000	SCEE=1.2	SCNB=2.0	Lipid14 CHL (Lipid14 cD-cD-cD-cD)
CA-CA-CA-hA	1	0.16	0.0	3.000	SCEE=1.2	SCNB=2.0	Lipid11 v1.0 (GAFF c3-c3-c3-hc)

cA-cA-cA-oH	1	0.1556	0.0	3.000	SCEE=1.2	SCNB=2.0	Lipid11	v1.0	(GAFF X -c3-c3-X )
cA-cA-cB-cA	1	0.000	0.0	2.000	SCEE=1.2	SCNB=2.0	Lipid11	v1.0	(GAFF X -c3-c2-X )
cA-cA-cB-cB	1	0.000	0.0	2.000	SCEE=1.2	SCNB=2.0	Lipid11	v1.0	(GAFF X -c3-c2-X )
cA-cA-cB-hB	1	0.000	0.0	2.000	SCEE=1.2	SCNB=2.0	Lipid11	v1.0	(GAFF X -c3-c2-X )
cA-cA-cD-cD	1	0.3112	180.000	1.000	SCEE=1.2	SCNB=2.0	Lipid14	CHL	(Lipid14 cD-cD-cD-cD)
cA-cA-cD-cD	1	-0.1233	180.000	2.000	SCEE=1.2	SCNB=2.0	Lipid14	CHL	(Lipid14 cD-cD-cD-cD)
cA-cA-cD-cD	1	0.1149	0.000	3.000	SCEE=1.2	SCNB=2.0	Lipid14	CHL	(Lipid14 cD-cD-cD-cD)
cA-cA-cD-cD	1	-0.2199	0.000	4.000	SCEE=1.2	SCNB=2.0	Lipid14	CHL	(Lipid14 cD-cD-cD-cD)
cA-cA-cD-cD	1	0.2170	0.000	5.000	SCEE=1.2	SCNB=2.0	Lipid14	CHL	(Lipid14 cD-cD-cD-cD)
cA-cA-cD-hL	1	0.16	0.0	3.000	SCEE=1.2	SCNB=2.0	Lipid14	CHL	(GAFF c3-c3-c3-hc)
cA-cA-oH-hO	1	0.16	0.0	-3.000	SCEE=1.2	SCNB=2.0	Lipid11	v1.0	(GAFF c3-c3-oh-ho)
cA-cA-oH-hO	1	0.25	0.0	1.000	SCEE=1.2	SCNB=2.0	Lipid11	v1.0	(GAFF c3-c3-oh-ho)
cA-cB-cA-hA	1	0.000	0.0	2.000	SCEE=1.2	SCNB=2.0	Lipid11	v1.0	(GAFF X -c3-c2-X )
cA-cB-cB-cA	1	6.65	180.0	-2.000	SCEE=1.2	SCNB=2.0	Lipid11	v1.0	(GAFF c3-c2-c2-c3)
cA-cB-cB-cA	1	1.90	180.0	1.000	SCEE=1.2	SCNB=2.0	Lipid11	v1.0	(GAFF c3-c2-c2-c3)
cA-cB-cB-hB	1	6.650	180.000	2.000	SCEE=1.2	SCNB=2.0	Lipid11	v1.0	(GAFF X -c2-c2-X )
cB-cA-cA-hA	1	0.1556	0.0	3.000	SCEE=1.2	SCNB=2.0	Lipid11	v1.0	(GAFF X -c3-c3-X )
cB-cA-cA-oH	1	0.1556	0.0	3.000	SCEE=1.2	SCNB=2.0	Lipid11	v1.0	(GAFF X -c3-c3-X )
cB-cB-cA-hA	1	0.38	180.0	-3.000	SCEE=1.2	SCNB=2.0	Lipid11	v1.0	(GAFF hc-c3-c2-c2)
cB-cB-cA-hA	1	1.15	0.0	1.000	SCEE=1.2	SCNB=2.0	Lipid11	v1.0	(GAFF hc-c3-c2-c2)
cB-cA-cA-hE	1	0.1556	0.0	3.000	SCEE=1.2	SCNB=2.0	Lipid11	v1.0	(GAFF X -c3-c3-X )
cD-cD-cD-cA	1	0.3112	180.000	1.000	SCEE=1.2	SCNB=2.0	Lipid14	CHL	(Lipid14 cD-cD-cD-cD)
cD-cD-cD-cA	1	-0.1233	180.000	2.000	SCEE=1.2	SCNB=2.0	Lipid14	CHL	(Lipid14 cD-cD-cD-cD)
cD-cD-cD-cA	1	0.1149	0.000	3.000	SCEE=1.2	SCNB=2.0	Lipid14	CHL	(Lipid14 cD-cD-cD-cD)
cD-cD-cD-cA	1	-0.2199	0.000	4.000	SCEE=1.2	SCNB=2.0	Lipid14	CHL	(Lipid14 cD-cD-cD-cD)
cD-cD-cD-cA	1	0.2170	0.000	5.000	SCEE=1.2	SCNB=2.0	Lipid14	CHL	(Lipid14 cD-cD-cD-cD)
cD-cD-cA-hA	1	0.16	0.0	3.000	SCEE=1.2	SCNB=2.0	Lipid14	CHL	(GAFF c3-c3-c3-hc)
cD-cA-cA-hA	1	0.16	0.0	3.000	SCEE=1.2	SCNB=2.0	Lipid14	CHL	(GAFF c3-c3-c3-hc)
cA-cD-cD-hL	1	0.16	0.0	3.000	SCEE=1.2	SCNB=2.0	Lipid14	CHL	(GAFF c3-c3-c3-hc)
hA-cA-cA-hA	1	0.15	0.0	3.000	SCEE=1.2	SCNB=2.0	Lipid11	v1.0	(GAFF hc-c3-c3-hc)
hA-cA-cA-hE	1	0.1556	0.0	3.000	SCEE=1.2	SCNB=2.0	Lipid11	v1.0	(GAFF X -c3-c3-X )
hA-cA-cA-oH	1	0.00	0.0	-3.000	SCEE=1.2	SCNB=2.0	Lipid11	v1.0	(GAFF hc-c3-c3-oh)
hA-cA-cA-oH	1	0.25	0.0	1.000	SCEE=1.2	SCNB=2.0	Lipid11	v1.0	(GAFF hc-c3-c3-oh)
hA-cA-cB-hB	1	0.000	0.0	2.000	SCEE=1.2	SCNB=2.0	Lipid11	v1.0	(GAFF X -c3-c2-X )
hA-cA-cD-hL	1	0.15	0.0	3.000	SCEE=1.2	SCNB=2.0	Lipid14	CHL	(GAFF hc-c3-c3-hc)
hE-cA-oH-hO	1	0.500	0.0	3.000	SCEE=1.2	SCNB=2.0	Lipid11	v1.0	(GAFF X -c3-oh-X )

IMPROPER

cA-cB-cB-cA	1	6.65	180.0	-2.000	SCEE=1.2	SCNB=2.0	Lipid11	v1.0	(GAFF c3-c2-c2-c3)
cA-cB-cB-cA	1	1.90	180.0	1.000	SCEE=1.2	SCNB=2.0	Lipid11	v1.0	(GAFF c3-c2-c2-c3)

NONBON

hA	1.4870	0.0157	OPLS
hO	0.0000	0.0000	OPLS
oH	1.7210	0.2104	OPLS



## REFERENCES

- (1) Lim, J. B.; Rogaski, B.; Klauda, J. B. Update of the Cholesterol Force Field Parameters in CHARMM. *J. Phys. Chem. B* **2011**, *116* (1), 203–210.
- (2) Jämbeck, J. P. M.; Lyubartsev, A. P. Another Piece of the Membrane Puzzle: Extending Slipids Further. *J. Chem. Theory Comput.* **2012**, *9* (1), 774–784.
- (3) Wang, J.; Wolf, R. M.; Caldwell, J. W.; Kollman, P. A.; Case, D. A. Development and Testing of a General Amber Force Field. *J. Comput. Chem.* **2004**, *25* (9), 1157–1174.
- (4) Dickson, C. J.; Madej, B. D.; Skjevik, Å. A.; Betz, R. M.; Teigen, K.; Gould, I. R.; Walker, R. C. Lipid14: The Amber Lipid Force Field. *J. Chem. Theory Comput.* **2014**, *10* (2), 865–879.
- (5) Case, D.; Babin, V.; Berryman, J.; Betz, R.; Cai, Q.; Cerutti, D.; Cheatham III, T.; Darden, T.; Duke, R.; Gohlke, H.; et al. Amber 14. *Amber 14* **2014**.
- (6) Pan, J.; Tristram-Nagle, S.; Nagle, J. F. Effect of Cholesterol on Structural and Mechanical Properties of Membranes Depends on Lipid Chain Saturation. *Phys. Rev. E Stat. Nonlin. Soft Matter Phys.* **2009**, *80* (2 Pt 1), 021931.
- (7) Edholm, O.; Nagle, J. F. Areas of Molecules in Membranes Consisting of Mixtures. *Biophys. J.* **2005**, *89* (3), 1827–1832.
- (8) Edidin, M. The State of Lipid Rafts: From Model Membranes to Cells. *Annu. Rev. Biophys. Biomol. Struct.* **2003**, *32* (1), 257–283.

- (9) Simons, K.; Vaz, W. L. C. Model Systems, Lipid Rafts, and Cell Membranes. *Annu. Rev. Biophys. Biomol. Struct.* **2004**, *33* (1), 269–295.
- (10) Hung, W.-C.; Lee, M.-T.; Chen, F.-Y.; Huang, H. W. The Condensing Effect of Cholesterol in Lipid Bilayers. *Biophys. J.* **2007**, *92* (11), 3960–3967.
- (11) Huang, J.; Buboltz, J. T.; Feigenson, G. W. Maximum Solubility of Cholesterol in Phosphatidylcholine and Phosphatidylethanolamine Bilayers. *Biochim. Biophys. Acta BBA - Biomembr.* **1999**, *1417* (1), 89–100.
- (12) Mainali, L.; Raguz, M.; Subczynski, W. K. Formation of Cholesterol Bilayer Domains Precedes Formation of Cholesterol Crystals in Cholesterol/Dimyristoylphosphatidylcholine Membranes: EPR and DSC Studies. *J. Phys. Chem. B* **2013**, *117* (30), 8994–9003.