Import parameters from othe

Import parameters from other Tasks

b

Is DIA file		Search Fraine Paran
Convert raw file	h raw h ed each search engines Run Prophet files to single file	New Set Modify Set © Set Uy Set name: APMS Blob © for n MATRIX Mascot Parame Create or modify M
Search Engine Parameters	Select search parameters	Create or modify G
	search engines	Create or modify C
Parameter Set (883) APMS_BioID 3 V		
Database 16HEK293Ref57cBapBevG	MS-GF+ modifications.	Enzyme
Fixed		max missed clea
Modifications selected modifications Variable Deamidated (NQ)	Available modifications Carbamidomethyl (C) Phospho (Y) Acetyl (N-term) GG (K)	Parent Mas Fragment Mas Peptide Mass Tok Binning to use on fragmen Theoretical_fragmen
Modifications Oxidation (M)	× *	Neutral Losses (H2O Isotope Peptide Max C
Set default modifications	3	MS-GF+ MSGFPL Paran
oteowizard Parameter set	Run TPP 🛛	MS-GFDB Para
arameters required for converting raw/wiff files 885) APMS_BiolD © View	TPP can be run after the search task is finished in search results page. TPP Name DDA_MEPCE TPP Parameter set (886) APMS_BiolD	Create or monity w
Add Files List of databases display or hi	Run Task de databases 🗜	Set Default Modification.
 If a database cannot be seen from a new ta hidden database cannot be used for a new ta 	ask, the database has been hidden by ProHits admin. A sk, but old tasks will still be able to used the database.	The modification list is from file "/Prohits/autoSeconverted to the Mascot style. It can be manually
 The database mormation is manually adde 	a by Promits administrator (./pop_dbs_mio.txt).	All modifications:
01BuddingYSTandREV Download		2-dimethylsuccinyl (C) 2-monomethylsuccinyl (C)
Time files compressed : Mr Number of sequences : 11 Version of Mascot : 2 Reverse sequence gi 999	on Mar 4 15:13:42 2013 1668 .3.02	2-nitroberzyl (Y) 2-succinyl (C) 2HPG (R) 3-deoxyglucosone (R) 3-phosphoglyceryl (K) 3sulfo (N-term)
02HEK293V57cRapRevTag Download [Homo sapiens] Number of residues : 33 Number of sequences : 77 Human adenovirus C(RefseqV57)txid http://ftp.thegpm.org/fasta/cRAP/cr ftp://ftp.thegpm.org/fasta/cRAP/cr	9921702 2477 1129951(146 proteins) 21p for contaminants from MPI rap.fasta for cRAP from GPM	UNE (L') -ONE (H) -ONE (H) -ONE-Delta:H(-2)O(-1) (C) -ONE-Delta:H(-2)O(-1) (H) -ONE-Delta:H(-2)O(-1) (K) -ONE-Delta:H(-2)O(-1) (K)
Acverse sequence >DECOV### Strep or some tags (GFP, BirA*, G >BirA-R118G H0QFJ5 >Streptavidin_P22629 >GST26_P08515 >mCherry_V9VHH0 >sp GFP AEOVI	ST, mCherry) List from Payman.	Acetyl (C) Acetyl (Potein N-term) Acetyl (Potein N-term) Acetyl (Potein N-term) Acetyl (T)

Set name: APMS_Biol	o o for machine	TRIPLET	OF1 ᅌ		
MATRIX Ma SCIENCE Cre	ascot Parameters eate or modify Mascot se	arch parar	meter set 🕜	×	[+]
X Cre	andem Parameters eate or modify GPM sear	ch parame	iter set	not saved	[+]
Cre	omet Parameters sate or modify COMET s	earch para	ımeter set 🕜 [online	e help]	[-]
					Save
Enzyme					
	Enzyme:	Trypsin	۵		
M	ax missed cleavages:	2 🗘	Semi-style cleave	age 🗌	
	Decoy:				
Advanced options					
	Parent Mass Type:	 Monoi 	sotopic Average	3	
	Fragment Mass Type:	 Monoi 	sotopic Average	1	
Per	tide Mass Tolerance:	35	ppm 📀		
		1.0005	amu offset po	sition to start the binning 0.4	
Theore	tical_fragment_ions:	1 0:	default peak shape	e, 1=M peak only	
Neutr	al Losses (H2O/NH3):				
	Isotope error:	0 🖸			
	Peptide Max Charge:	2+, 3+ 8	nd 4+ 🖸		
MS-GF+ Cre	SGFPL Parameters eate or modify MSGFPL	search par	rameter set 🕐	not saved	[+]
	S-GFDB Parameters	search na	arameter set 🚱	not saved	[+]

ameter Set

The modification list is from file "/Prohits/autoSearch/mod_file" which originated from Uniprot and was converted to the Mascot style. It can be manually modified.



Supplementary Fig. 1. Initiating searches for Data Dependent Acquisition.

03--HEK293RefV57cRapRev Download

a) Search Task overview. Use drop-down menus and clickable options to select parameters and options or import from a previously-defined task. Generic parameter sets for each instrument and search engine can be selected (see panel b), the search engine(s) to be used specified, and the database and modifications chosen. c) To keep the interface clean and avoid human error, an administrator can define which sequence databases will be visible to users. d) An administrator can set the list of modifications which all users are allowed to use in setting up their tasks. Additional modifications can be added at any time by an administrator and made available for all users or for selected users. ProHits tracks all of these parameters: once a task has been started, these parameters cannot be further modified, though new files can be added to the search task.



Supplementary Fig. 2. Running DIA-Umpire untargeted identification through the ProHits interface.

a) Select "DIA file" and "MS-Umpire". In this Data Management module of ProHits, DIA-Umpire SE (Signal Extraction) generates pseudo-MS/MS spectra from precursor-fragment group data. These spectra can be searched using standard tools designed for DDA analysis, including the iProphet pipeline; note that DIA-Umpire SE extracts three quality tiers of precursor-fragment group data that are each searched and modeled separately (Supplementary Fig. 3). The final iProphet ProteinProphet result merges all quality groups for all search engines used. Currently, Comet, MSGF+, X!Tandem and Mascot are supported. Select the desired database search engines, protein database and search parameters that will be used to search the pseudo MS/MS spectra. b) Select the DIA-Umpire parameters. DIA-Umpire supports a variable window design (see Tsou et al., Nature Methods, 2015). In this case, specify each individual window's boundaries. Leave blank if fixed windows are used.





a) iProphet pipeline. DIA-Umpire separates the precursor-fragment groups in three quality tiers. Q1 corresponds to MS1 clusters of 3 or more isotope peaks, Q2, to those with 2, and Q3 to those that have been detected only as clusters of at least 2 unfragmented precursor ion peaks in MS2. Because of the different ratios of correct versus incorrect identifications across these tiers, PeptideProphet is run separately for each of the tiers (see details in DIA-Umpire original paper by Tsou et al., Nature Methods 2015). To combine results from multiple search engines into one final report, searches using each search engine are first performed on individual quality tiers, and PeptideProphet is performed on those results. InterProphet parser is used to combine the results into a single pep.xml file for each of the quality tiers, and once again to combine across all search engines and all quality tiers (this is the version used for extraction of gene level information). iProphet (ProteinProphet) is then applied to these three combined pep.xml files to yield the final protein list. Note that the gene level viewer instead extracts information directly from the pep.xml to perform mapping at the gene level (grey); see Supplementary Fig. 6 for additional information. b) In addition to providing an iProphet view of the data, pep.xml and prot.xml files are generated for each search engine. In this case, PeptideProphet is run on the individual files, which are combined into a single pep.xml using the InterProphet parser (gene level information is extracted from the resulting file). ProteinProphet runs directly on the Q1, Q2 and Q3 pep.xml outputs to generate the prot.xml file.

OI

Comet

Peptide

Prophet

OI.pep.xml

ProteinProphet

prot.xml

Protein inference

О2

Comet

Peptide

Prophet

O2.pep.xm

О3

Comet

Peptide

Prophet

O3.pep.xml

InterProphet parser

pep.xml

Gene mapping

Filter

(see above)

Select Instrumen	t and search engine		
Machine TRIPLETOF1 O XTandem MSGFPL	Mascot O Comet O iProphet •		Process files
Searched tasks Tasks name [Tasks ID] UMPIRE_MEPCE_EIF_50ppm_XTandem_MSGF_n2 (237) Umpire_MEPCE (269) Select at least I task (only tasks with the same search parameters can be re-extracted jointly)	Raw files Raw file name [Raw file ID, Sample ID, Tasks ID] LongSwath. GFPJune7-Biorep1.wiff [27788] [17333] [269] LongSwath. GFPJune7-Biorep2.wiff [27791] [17334] [269] LongSwath. GFPJune7-Biorep2.wiff [2794] [17355] [269] LongSwath. ElF4aJune7-Biorep2.wiff [27854] [17578] [269] LongSwath. ElF4aJune7-Biorep3.wiff [27869] [17579] [269]	Selected raw files Raw file name [Raw file ID, Sample ID, Ta LongSwath_MEPCEJune7-Biorep1.wiff [27761] [17: LongSwath_MEPCEJune7-Biorep2.wiff [2773] [17: LongSwath_MEPCEJune7-Biorep3.wiff [27818] [17:	asks ID) 321] [269] 322] [269] 323] [269]
		>> <<	

DIA-Umpire Quant parameters 🛛

а

b

DIA-Umpire Q	Jant								
Task Name MEPCE_EIF4A_SAINT									
π	ask Description	Biolo	gical tripli	icates					
Tar	getedExtraction	true:	 false 	: ()					
	PeptideFDR	0.05							
	ProteinFDR	0.05							
	ProbThreshold	0.9							
	FilterWeight	GW:	Pep\	N: ()					
	MinWeight	0.9							
	TopNFrag	6	Suggest	ed value		IT=6: m	anDIA=20		
	TopNRop	0	Cuerce		- 041	IT-0, III			
	торигер	ь	Suggesi	eu value	es SAIN	11-0; m	арыя-20		
	Freq	0.5	Suggest	ed value	es SAIN	T=0.5;	mapDIA=0		
Run SAINT: 🧿	Run mapDIA:	0	Only Ru	in DIA-U	Impire-	Quant: (0		
Raw File ID	Sample Name					Bait Na	me/Label sample nam	ie	Is control
27761	MEPCE_SWATH_L	ongMS	1_BR1_Ju	ine		MEPCE		* *	
27773	MEPCE_SWATH_L	ongMS	1_BR2_Ju	ine		MEPCE		* *	
27818	MEPCE_SWATH_L	ongMS	1_BR3_Ju	ine		MEPCE		* *	
27788	GFP_SWATH_Long	MS1_E	3R1_June			GFP		* *	
27791	GFP_SWATH_Long	MS1_E	3R2_June			GFP		**	Image: A state of the state
27794	GFP_SWATH_Long	MS1_E	3R3_June			GFP		* *	
27851	27851 EIF4A2_SWATH_LongMS1_BR1					EIF4A2		* *	
27854 EIF4A2_SWATH_LongMS1_BR2						EIF4A2		* *	
27869 EIF4A2 SWATH LongMS1 BR3									_

Select desired parameters for targeted re-extraction

(Here, the suggested values for SAINT scoring are shown)

Options available for scoring: SAINT (shown here) mapDIA (see Sup Fig 6) none (run only DIA-Umpire Quant)

- If running SAINT, select which files are "control" runs

C SAINT parameters 🛛

	INT	5.00								
SAINT express(exp3.3) SA	MN I (2.)	5.0)								
Use SAINT with controls	You hav	ve selec	ted 3	control san	nple(s) in	previous step.				
	How ma	any con	press	sed controls	: 3					
Compress baits	3	replica	ites in	each intera	action wit	th the highest counts is inv	olved	in the		
	computa	ation of	the so	cores		v				
SAINT (2.5.0) parameters										
Burn-in period	nburn: 2	2000				Iterations	niter:	5000		
exclude extremely high counts	lowMod	e: 0				forcing separation	minF	old: 1		
	divide	spect	ral co	unts by the	e total sp	pectral counts of each IP	norm	alize: 1	L	
		Run (DIA-Un	npire Quant	& SAINT					

Specify SAINT parameters. By default, ProHits will run both MSI and MS2 data. ProHits will run both SAINT and SAINTexpress.

The options shown here (other than compression) are only used by standard SAINT.

Supplementary Figure 4. Running the semi-targeted re-extraction module of DIA-Umpire (DIA-Umpire Quant).

a) To be able to perform targeted extraction using DIA-Umpire's Quant, a DIA-Umpire SE task must first have been run in the Data Management module. All files to be re-extracted jointly must have been searched with the same database and search parameters. Select the files for re-extraction. b) Specify the parameters for DIA-Umpire extraction, and whether you want to only run DIA-Umpire or to also analyze the data with SAINT (panel c) or mapDIA (Supplementary Fig. 7). If running SAINT, specify here which samples should be used as negative controls. Note that in SAINT, all samples listed with the same name (here, the gene name by default) will be combined as one SAINT bait (with n replicates): If a separation of the samples is desired, either click the "Use Sample Name" box, or manually force separation by giving each sample a different name. c) By default, ProHits will initiate the following SAINT tasks: SAINTexpress, intensity model, with both MS1 and MS2 quantification, and standard SAINT (currently SAINT 2.5.0), also with both MS1 and MS2 quantification. In each case, the controls can be compressed (to the highest values, a more stringent parameter for scoring), and the baits can also be compressed (meaning that for each prey, the x highest values will be considered). The parameters for standard SAINT can be specified. Each of these four SAINT variations is then associated with a SAINT report that can be viewed online or downloaded for further analysis.

a



Import parameters from previous Tasks



Supplementary Fig. 5. Running MSPLIT-DIA through the ProHits interface.

a) Select "DIA file" and "MSPLIT-DIA". If using DDA files to create a library, select DDA search parameters (only MSGFDB is currently supported as a search engine). Add both DDA and DIA (SWATH) files to the list of files and select which of those are the DDA files that will be used for library generation. Existing libraries can also be searched: individual external libraries are listed (here Human_Swath_Atlas_v1 from the Aebersold group). In addition, ProHits automatically generates a spectral library from all previously-searched DDA files that it appends to externally generated libraries ("Archive"). These can be searched in isolation, or alongside the newly searched results. In all cases, ProHits will take the highest MSGFDB spectrum for spectral matching with MSPLIT-DIA. b) Select the parameters to be used for creating the spectral library from MSGFDB results, creating the decoy spectral library, performing the spectral library search and filtering the search results using retention time correlation. MSPLIT-DIA supports a variable window design (this was not specifically described in Wang et al., Nature Methods, 2015). In this case, specify each individual window's boundaries. Leave blank if fixed windows are used.



Supplementary Fig. 6. Results page for MSPLIT-DIA in the Data Management module.

a) ProHits tracks all search parameters to facilitate reporting. The identification results for each file can be downloaded as a tab delimited file (download icon by each file name). Alternatively, the results can be parsed to the "Analyst" module of ProHits. ProHits also facilitates the generation through MSPLIT-DIA of "assay libraries" for targeted extraction by the popular tools OpenSWATH, PeakView and Skyline. b) Gene level reports (e.g. here for MSPLIT-DIA) are distinguishable from protein level reports through color-coded icons. c) Gene level views. Peptides are first matched to proteins and those identifications are then used for mapping peptides to genes (using Refseq for protein and gene IDs). For peptides shared between multiple genes, spectral counts are assigned exclusively to those genes with unique peptides in proportion to the existing evidence for those genes. If a peptide matches exclusively to genes that have no unique peptides, then spectral counts are divided equally between the genes. After all spectra have been assigned, counts for each gene are rounded to the nearest integer. The final report contains all genes to which peptides have been matched. For each gene, "shared genes" match the exact same set of peptides. "Subsumed genes" are also reported that include genes that match to a subset of the current gene's peptides and nothing outside of this set. In addition to total spectral counts, unique peptide counts are also reported for each gene. In the case of genes that match to the exact same set of peptides, this "unique" number will refer to the number of unique peptides for the corresponding group.



Supplementary Fig. 7. Gene inference.

a) Peptides are first matched to proteins and those identifications are then used for mapping peptides to genes (using Refseq for protein and gene IDs). For peptides shared between multiple genes, spectral counts are assigned exclusively to those genes with unique peptides in proportion to the existing evidence for those genes. If a peptide matches exclusively to genes that have no unique peptides, then spectral counts are divided equally between the genes. b) The final report contains all genes to which peptides have been matched. For each gene, "shared genes" are reported if they exist. These encompass genes that match to the exact same set of peptides, i.e. we have no evidence to determine which of the corresponding proteins is in the sample although the peptides tell us a least one is. "Subsumed genes" are also reported that include genes that match to a subset of the current gene's peptides and nothing outside of this set. In addition to total spectral counts, unique peptide counts are also reported for each gene. In the case of genes that match to the exact same set of peptides, this "unique" number will refer to the number of unique peptides for the corresponding group.

a DIA-Umpire Quant parameters 🛛

DIA-Umpire	Quant					
	Task Name	MEPCE_EIF4A_mapDIA	l l			Select desired parameters for targeted re-extraction
Т	ask Description					
				12		(Here, the suggested values for mapDIA scoring are shown
Tar	getedExtraction	true: false: 				
	PeptideFDR	0.05				
	ProteinFDR	0.05				
	ProbThreshold	0.9				
	FilterWeight	GW: 💿 PepW: 🔘				
	MinWeight	0.9				
	TopNFrag	20 Suggested value	es SAINT=6: mapDIA=20			
	TonNPen	20 Suggested value				Options available for scoring:
		20 Suggested value	5 5AINT-0, MapDIA-20			- SAINT (see Sup. Figure 3)
	Freq	0 Suggested value	es SAINT=0.5; mapDIA=0			mapDIA (here)
Run SAINT:	Run mapD	IA: 💿 🛛 Only Run Dl	A-Umpire-Quant: 🔾 🗲			- none (run only DIA-Umpire Quant)
Raw File ID	Sample Name		Bait Name/Label Use sample name		is control	
27788	GFP_SWATH_Lo	ongMS1_BR1_June	GFP	▲ ▼		
27791	GFP_SWATH_Lo	ongMS1_BR2_June	GFP	▲ ▼		
27794	GFP_SWATH_Lo	ongMS1_BR3_June	GFP	★ ₹		Use arrows to organize order as desired
27761	MEPCE_SWATH	LongMS1_BR1_June	MEPCE	★ ▼		
27773	MEPCE_SWATH	LongMS1_BR2_June	MEPCE	* *		
27818	MEPCE_SWATH	LongMS1_BR3_June	MEPCE	* *		
27851	EIF4A2_SWATH	LongMS1_BR1	EIF4A2	★ ▼		
27854	EIF4A2_SWATH	_LongMS1_BR2	EIF4A2	* *		
27869	EIF4A2_SWATH	LongMS1_BR3	EIF4A2	★ ♥		
			Next			
						Link to the mapDIA user manual

b mapDIA parameters?

С

http://xtandemserver.mshri.	n.ca/thegpm-cgi/			
mapDIA (version:2.0.5)				
Experimental design	replicatedesign REP design: For exa biological replicates he following order (t	€ ample, if the de (A,B), then the t1-A, t1-B) (t2-/	esign is a time course experiment with 3 time points (t1,t2,t3) a conditions are time points and thus the samples should be or A, t2-B) (t3-A, t3-B)	cross 2 ganized in
Normalization	none none/T	IS/rt 30		
Filter	SDF 2 MIN_CORREL 0.2 MIN_FRAG_PER_P MAX_FRAG_PER_F MIN_PEP_PER_PR	EP 3 PEP 5 OT 1		
Sample information	Group # Raw file ID		LABEL	MIN_OBS
	1 27794 2779	91 27788	GFP	2
	2 27818 277	73 27761	MEPCE	2
	3 27869 2785	54 27851	EIF4A2	2
DEPs	MIN_DE 0.01 MA	4X_DE 0.99		
Protein_level	MAX_PEP_PER_PF	ROT 5		
Contrast matrix for group comparison				

Specify experimental design and parameters

Suggested parameters are listed by default

Run DIA-Umpire Quant & mapDIA

mapDIA "analysis output"

	_	A	B	C	D	E	F	G	H	1	J	K	L	M	N	0	P	Q	R	S
1	P	Protein	nPeptide	nFragment	Label	Label2	log2FC	score	FDR	log_oddsDE	log2FC_1	log2FC_2	log2FC_3	nUp	nDown	mcr	min.mcr	max.mcr	PreyGeneID	PreyGene
2	C	213347	8	41	2 to 1	EIF4A2/GFP	2.33989	1	. 0	164.192	2.1362	2.42028	2.46321	3	C	0.756243	0.702197	0.785021	EIF3I	8668
3	C	075821	5	28	2 to 1	EIF4A2/GFP	2.31502	1	. 0	169.756	2.5796	1.94332	2.42214	3	C	1	1	1	EIF3G	8666
4	C	200303	4	15	2 to 0	EIF4A2/MEPCE	2.23984	1	. 0	54.1806	2.03122	2.6019	2.08639	3	C	0.864434	0.594655	0.999517	EIF3F	8665
5	P	P60842	7	32	2 to 1	EIF4A2/GFP	2.0113	1	. 0	67.3016	1.53731	2.43739	2.0592	3	C	0.599081	0.514769	0.714154	EIF4A1	1973
6	P	22626	3	12	2 to 1	EIF4A2/GFP	1.99311	1	. 0	69.5806	1.92326	1.90409	2.15198	3	C	0.81223	0.775	0.884661	HNRNPA2B1	3181
7	P	P62753	4	20	2 to 1	EIF4A2/GFP	1.79617	1	. 0	134.034	1.97417	1.69194	1.72239	3	C	0.969364	0.942671	0.983032	RPS6	6194
8	C	000571	7	39	2 to 1	EIF4A2/GFP	1.7518	1	. 0	121.191	1.85427	1.9021	1.49904	3	C	0.83772	0.75	0.964453	DDX3X	1654
9	P	P60842	5	22	2 to 0	EIF4A2/MEPCE	1.67667	1	. 0	52.961	1.48226	1.39583	2.15192	3	C	0.599081	0.514769	0.714154	EIF4A1	1973

Supplementary Fig. 8. Running mapDIA from DIA-Umpire through ProHits.

a) When initiating the DIA-Umpire semi-targeted extraction, select to run mapDIA. Use arrows to organize the data as desired to accommodate experimental design. b) Select mapDIA parameters (see linked mapDIA user manual for detailed explanation of the experimental design and parameters. c) mapDIA returns a folder with several files, including this "analysis output" file which contains quantitative information and statistical parameters. Note that this can be used as an input for visualization tools (e.g. Fig. 2) that are currently run outside of ProHits.



Supplementary Fig. 9. ProHits facilitates the deposition of data in ProteomeXchange via MassIVE.

Files to be exported to MassIVE can be selected as a batch from the SAINT Comparison page (a), or manually added to the export list by clicking the download buttons located by individual files or folders in Data Management or individual Samples in the Analyst module (b). All files will be added to a "Export Raw files" list in Data Management (c). To export the data for complete ProteomeXchange submission at MassIVE, select the appropriate button, and login using your credentials. d) ProHits will retrieve both the Peaks list and the search results associated with the selected Raw files: ProHits uses by default the "Search Task" for which the results were parsed to the Analyst; this can be overridden by the user through the dropdown menu selection. For iProphet results, ProHits uses the TPP converters and renames the files for direct recognition in MassIVE. Complete submissions also require deposition of the FASTA sequence database that can be selected via the dropdown menus. e) ProHits generates at MassIVE subdirectories containing the Raw, Peak List, Results and FASTA files. They are named according to the MassIVE nomenclature and can easily be selected. f) MassIVE should recognize automatically all the files and associate them.

Saint Comparison

Color code Hit pro	operty color code 💿 Share	ed hits color code	
Sort by Spec	Sum ᅌ Sample Nar	me EIF4A2 ᅌ Descend	ing 💿 Ascending 🔵
Click to remove filters]			
Experiment Filters			
SaintScore <		SpecSum < 📀	
AvgP < 📀		maxSpec < 📀	
MaxP < 📀		NumReplicates <	0
BFDR > 0.02 ᅌ		Background Set ba	ackground list ᅌ
Frequency > 🔗 %			
Bio Filters			
Heat Shock	Ribosomal	Cytoskeleton	Bait
Keratin	Artifact Protein	Translation Elongation	Factor DEAD/H Box
Rib Nucleoprotein	 Histone 	Albumin	
BioGRID BioGRID overla	IP		
🗌 Physical HTP 🚖	Physical NON-HTP A	Genetic HTP 🛣	□ Genetic NON-HTP △
0 84 169 253 338 422 50	Select al	l desired filters, tl	nen hit "Go" 🛛 🚳
Cytoscape] [Export (table)] [Export (matrix)] [Export to	PSI-MI] [Send by email] [Export (Prohits web)] [Export (ra
F4A2	BG	Pre	ey 🚽
EI	Ge	ne Name Pro	tein ID Remove
144	EIF4A'	1 [BioGRID] 45	03529
45	EIF4G	1 [BioGRID] 302	699237
09	EIF3A	[BioGRID] 45	03509
28	PDCD	(BioGRID) 211	35596

Multiple export and visualization options are available: they will display / report the visible (post-filter) list

EIF4G2 [BioGRID]

EIF4G3 [BioGRID]

EIF3C [BioGRID]

289577080

311771714



To use the customizable export function, first select which files are to be exported, as well as the filters when applicable.

Cytoscape view of the data h



email / web link to a static html view of the data

Sent link:	gingras_94_seajudgh3dpp0m189pnnrijd14_2D_1449443980.html	
To:		
Subject:		
Contents:		
	Send Reset Close	

[Send from email account of your choice]

С

e

- Option 1: Save the FILE to your local computer by select "SAVE LINK/TARGET AS" (shown upon right click). Then attach the saved file to you email. • Option 2: Include the following URL link in your email: http://prohits-web.lunenfield.ca/prohits_report /gingras_94_seajudgh3dpp0m189pnnrijd14_2D_1449443980.html

Export Sample Report (Project: Gingras_SWATH_development)

Export rows as CSV ᅌ Preview	Generate Report	
Please select column	ns to be included in the export file	Pre-defined export format
Bait:		
🗹 Bait ID	Bait Tax ID	
🗹 Bait Gene ID	Bait Acc	[new]
Bait Gene Name	Bait Acc Type	
Bait Locus Tag	Bait MW	Selected columns
Bait Clone	Bait Vector	2.11.12
Bait Description	Is Gel Free	Bait ID
Experiment:		Bait Gene ID
∃ Sample:		Sample ID
Sample ID	Sample MW	Sample Name
Sample Intensity	Sample Name	Protein Gene Name
Instrument	Raw File Name	Protein ID
Raw File Date	Raw File Size	Protein Gene ID
Raw file path	Task ID	Protein Probability
TPP task ID	Search parameters	Total Number Peptide
TPP Protein:		Unique Number Peptide
TppID	Protein Gene Name	Coverage Percentage
Protein Acc Type	Protein Gene ID	
Protein ID	Protein Acc	
Protein Locus Tag	Protein Probability	
PCT Spectrum IDs	Indistinguishable Protein	
Protein Dec.	Total Number Peptide	
Unique Number Peptide	Coverage Percentage	
Xml File	Search Engine	
Searched Database	Xpressratio Mean	
 Xpressratio STandard Dev. 	Xpressratio Number Peptide	
Project Frequency	Filters	
 Hit Protein Length 		
TPP Protein Group Peptide:		

Supplementary Fig. 10. Selected visualization and export options in ProHits.

a) Multiple export and visualization options are available from any SAINT Comparison page. First select all desired filters and apply them by pressing "Go". Manual removal of selected hits can also be performed. The final view of the data can be downloaded as a table or matrix format, converted to the PSI-MI format, directly exported to the quantitative interaction proteomics repository prohits-web.lunenfeld.ca, viewed as a Cytoscape figure (c), or emailed in a static html format with functional links (d). Additional export options are also available, e.g. that allow for the selection of any file (d) and download of selected values (e).