### Expression of varied GFPs in Saccharomyces cerevisiae: codon optimization yields stronger than expected expression and fluorescence intensity

# Misato Kaishima<sup>1</sup>, Jun Ishii<sup>2</sup>, Toshihide Matsuno<sup>2</sup>, Nobuo Fukuda<sup>3</sup> and Akihiko Kondo<sup>1</sup>

<sup>1</sup>Department of Chemical Science and Engineering, Graduate School of Engineering, Kobe University, 1-1 Rokkodai, Nada, Kobe 657-8501, Japan <sup>2</sup>Graduate School of Science, Technology and Innovation, Kobe University, 1-1 Rokkodai, Nada, Kobe 657-8501, Japan <sup>3</sup>Biomedical Research Institute, National Institute of Advanced Industrial Science and Technology (AIST), Higashi, Tsukuba, Japan



Figure S1. Histogram plots illustrating codon-optimized and non-codon-optimized GFPs expression in *S. cerevisiae.* The histogram plots show analytical data measured for 10,000 cells using flow cytometry.



Figure S2. Visual images of the cells under natural light. Varied codon-optimized and non-codon-optimized GFPs were used in the evaluations.

#### (A) <u>Gy recruitment system for soluble cytosolic target protein</u>



Figure S3. Schematic diagram of G $\gamma$  recruitment systems used to detect PPIs of cytosolic or membrane target proteins. (A) Schematic outline of the G $\gamma$  recruitment system for cytosolic target proteins. When target protein 'X', which is fused to G $\gamma_{cyto}$ , interacts with candidate protein 'Y<sub>1</sub>,' the G $\beta$  and G $\gamma_{cyto}$  complex (G $\beta\gamma_{cyto}$ ) migrates to the inner leaflet of the plasma membrane and restores the signaling function. If protein 'X' cannot interact with protein 'Y' G $\beta\gamma_{cyto}$  is released into the cytosol, and signaling is blocked. (B) Schematic outline of the G $\gamma$  recruitment system for membrane protein targets. When membrane target protein 'X' interacts with candidate protein 'Y', which is fused to G $\gamma_{cyto}$ , the G $\beta\gamma_{cyto}$  complex migrates to the inner leaflet of the plasma membrane and restores the signaling function. If membrane protein 'X' cannot interact with protein 'Y', G $\beta\gamma_{cyto}$  is released into the cytosol, and signaling is blocked. (C) Flow diagram of the screening procedure used in the G $\gamma$  recruitment system. Two selection methods are available to screen for new binding proteins. One method uses *GFP* reporter genes. When target candidate proteins are expressed in yeast cells and interact with each other, the proteins induce *GFP* expression, which is detected by flow cytometry.



### Figure S4. Expression of non-codon-optimized EGFP and codon-optimized ymUkG1 as fusion-tagged proteins to detect PPIs using the Gγ recruitment system.

Flow cytometry analyses using the G $\gamma$  recruitment system for membrane protein targets. The Fc protein was used as the membrane target protein 'X' and was expressed as a membrane-associated protein with an N-terminal lipid anchor (derived from Gpa1p). Four Z variants ( $Z_{WT}$ ,  $Z_{K35A,}$ ,  $Z_{I31A}$  and  $Z_{955}$ ) were used as the cytosolic candidate 'Y' proteins and were expressed as fusion proteins with G $\gamma_{cyto}$  (G $\gamma_{cyto}$ -Z variants). 'Control' indicates FN-G0 and UG2-FNG0 yeast strains without the expression of 'G $\gamma_{cyto}$ -Y'. The engineered strains were grown in media containing 5  $\mu$ M  $\alpha$ -factor and were used in the analyses of mean fluorescence intensities (MFIs). The MFIs of 10,000 cells were measured by flow cytometry.



Figure S5. Determination of gate area to dominantly including positive fluorescent cells (GFP<sup>+</sup>). Data were presented as dot plots (forward scatter, FSC-A vs green fluorescence, GFP). Y-axis is an indication of fluorescence and X-axis is an approximation of relative cell size. Positive cells express membrane-anchored  $Z_{WT}$ ,  $Z_{K35A}$  and  $Z_{I31A}$  with  $G\gamma_{cyto}$ -Fc. Negative cells express membrane-anchored  $Z_{955}$  with  $G\gamma_{cyto}$ -Fc. 'Control' indicates BFG2118 and UGFG2 yeast strains harboring the pGK413 mock plasmid (without the expression of 'Y'). (A) Flow cytometry analysis of the BFG2118 yeast transformants (EGFP reporter). (B) Flow cytometry analysis of the UGFG2 yeast transformants (ymUkG1 reporter).

### Supplementary Table S1. Nucleic acid sequences of codon-optimized GFPs for the Saccharomyces cerevisiae

#### yEGFP

#### yAcGFP1

#### yTagGFP2

#### ymUkG1

#### yZsGreen

#### ymWasabi

ATGGTCAGTAAGGGTGAAGAAACTACTATGGGTGTTATCAAGCCAGACATGAAGATCAAGTTGAAGATGGAAGGTAA CGTTAACGGTCATGCCTTTGTTATTGAAGGTGAAGGTGAAGGTAAACCATACGATGGTACTAATACCATTAACTTGGA AGTCAAAGAAGGTGCTCCATTGCCATTCTCTTACGATATTTTGACTACCGCTTTCTCATACGGTAATAGAGCTTTTACT AAGTACCCAGATGACATCCCAAACTACTTCCAAGCAATCTTTTCCAGAAGGTTACTCTTGGGAAAGAACTATGACTTTC GAAGATAAGGGTATCGTCAAGGTTAAGTCCGATATCTCTATGGAAGAAGATTCCTTCATCTACGAAATCCACTTGAAG GGTGAAAATTTCCCACCAAATGGTCCAGTCATGCAAAAAGAAAAGAAACAACTGGTTGGGATGCTTCTACCGAAAGAATGTA TGTTAGAGATGGTGTCTTGAAAGGTGACGTCAAAATGAAGTTGTTGTTGGGAAGGTGGTGGTCATCATAGAGTTGATT TCAAGACTATCTACAGAGCTAAGAAGGCTGTTAAGTTGCCAGATTACCATTTCGTTGATCACAGAATCGAAATCTTGA ACCACGATAAGGATTACAACAAGGTTACCGTTTACGAAATTGCTGTTGCTAGAAACTCTACCGATGGTATGGATGAATT ATACAAGTAA

#### ymNeonGreen

### Supplementary Table S2. List of yeast transformants used for expression of various GFPs

Transformant	Expression of protein	Figure
BY4741 (Control)	-	Fig.1
BY4741 + pGK416-EGFP	EGFP (mammalian codon-optimized)	Figs.1 and S2
BY4741 + pGK416-AcGFP1	AcGFP1 (mammalian codon-optimized)	Figs.1 and S2
BY4741 + pGK416-TagGFP2	TagGFP2 (mammalian codon-optimized)	Figs.1 and S2
BY4741 + pGK416-mUkG1	mUkG1	Figs.1 and S2
BY4741 + pGK416-ZsGreen	ZsGreen	Figs.1 and S2
BY4741 + pGK416-yEGFP	yEGFP (yeast codon-optimized)	Figs.1 , S1 and S2
BY4741 + pGK416-yAcGFP1	yAcGFP1 (yeast codon-optimized)	Figs.1 , S1 and S2
BY4741 + pGK416-yTagGFP2	yTagGFP2 (yeast codon-optimized)	Figs.1 , S1 and S2
BY4741 + pGK416-ymUkG1	ymUkG1 (yeast codon-optimized)	Figs.1 , S1 and S2
BY4741 + pGK416-yZsGreen	yZsGreen (yeast codon-optimized)	Figs.1 , S1 and S2
BY4741 + pGK416-ymWasabi	ymWasabi (yeast codon-optimized)	Fig. S1
BY4741 + pGK416-ymNeonGreen	ymNeonGeen (yeast codon-optimized)	Fig. S1
BY4741 + pGK416-EGFP-F	EGFP (mammalian codon-optimized)-FLAG	Fig.1
BY4741 + pGK416-AcGFP1-F	AcGFP1 (mammalian codon-optimized)-FLAG	Fig.1
BY4741 + pGK416-TagGFP2-F	TagGFP2 (mammalian codon-optimized)-FLAG	Fig.1
BY4741 + pGK416-mUkG1-F	mUkG1-FLAG	Fig.1
BY4741 + pGK416-ZsGreen-F	ZsGreen-FLAG	Fig.1
BY4741 + pGK416-yEGFP-F	yEGFP (yeast codon-optimized)-FLAG	Fig.1
BY4741 + pGK416-yAcGFP1-F	yAcGFP1 (yeast codon-optimized)-FLAG	Fig.1
BY4741 + pGK416-yTagGFP2-F	yTagGFP2 (yeast codon-optimized)-FLAG	Fig.1
BY4741 + pGK416-ymUkG1-F	ymUkG1 (yeast codon-optimized)-FLAG	Fig.1
BY4741 + pGK416-yZsGreen-F	yZsGreen (yeast codon-optimized)-FLAG	Fig.1
BY4741 + pGK416-ymWasabi-F	ymWasabi (yeast codon-optimized)-FLAG	Fig.1
BY4741 + pGK416-ymNeonGreen-F	ymNeonGeen (yeast codon-optimized)-FLAG	Fig.1

Supplementary Table S3. Mean fluorescence intensity (MFI) of yeast transformants used for expression of various GFPs

GFP	MFI
Control	36
EGFP	1,490
AcGFP1	542
TagGFP2	240
mUkG1	14,194
ZsGreen	15,019
yEGFP	33,551
yAcGFP1	36,177
yTagGFP2	24,250
ymUkG1	47,088
yZsGreen	44,154
ymWasabi	16,987
ymNeonGreen	5,148

# Supplementary Table S4. List of yeast strains used for expression of various Fig1-GFPs as fusion tagged reporters

Strain	Expression of protein	Figure
BY4741 (Control)	_	Fig. 2
MC-F1	Fig1-EGFP (mammalian codon-optimized)	Fig. 2
BYFAG1	Fig1-AcGFP1 (mammalian codon-optimized)	Fig. 2
BYFTG1	Fig1-TagGFP2 (mammalian codon-optimized)	Fig. 2
BYFUG1	Fig1-mUkG1	Fig. 2
BYFZG1	Fig1-ZsGreen	Fig. 2
BYFEG2	Fig1-yEGFP (yeast codon-optimized)	Fig. 2
BYFAG2	Fig1-yAcGFP1 (yeast codon-optimized)	Fig. 2
BYFTG2	Fig1-yTagGFP2 (yeast codon-optimized)	Fig. 2
BYFUG2	Fig1-ymUkG1 (yeast codon-optimized)	Fig. 2
BYFZG2	Fig1-yZsGreen (yeast codon-optimized)	Fig. 2

# Supplementary Table S5. List of yeast transformants used for the G $\gamma$ recruitment system for soluble cytosolic target protein

Transformant	Target	Candidate	Repoter	Figure
	X (cytosol)	Y (membrane)	GFP	
BFG2118 + pGk413 (Control)	Fc	_	EGFP	Figs. 3(A) and S5(A)
BFG2118 + pGK413-ZWTmem	Fc	Z <sub>WT</sub>	EGFP	Figs. 3(A), 4(A), 4(B) and S5(A)
BFG2118 + pGK413-ZK35Amem	Fc	Z <sub>K35A</sub>	EGFP	Figs. 3(A) and S5(A)
BFG2118 + pGK413-ZI31Amem	Fc	Z <sub>I31A</sub>	EGFP	Figs. 3(A), 4(E), 4(F) and S5(A)
BFG2118 + pGK413-Z955mem	Fc	Z <sub>955</sub>	EGFP	Figs. 3(A), 4(A), 4(B), 4(E), 4(F)
				and S5(A)
UGFG2 + pGk413 (Control)	Fc	-	ymUkG1	Figs. 3(A) and S5(B)
UGFG2 + pGK413-ZWTmem	Fc	Z <sub>WT</sub>	ymUkG1	Figs. 3(A), 4(C), 4(D) and S5(B)
UGFG2 + pGK413-ZK35Amem	Fc	Z <sub>K35A</sub>	ymUkG1	Figs. 3(A) and S5(B)
UGFG2 + pGK413-ZI31Amem	Fc	Z <sub>I31A</sub>	ymUkG1	Figs. 3(A), 4(G), 4(H) and S5(B)
UGFG2 + pGK413-Z955mem	Fc	Z <sub>955</sub>	ymUkG1	Figs. 3(A), 4(C), 4(D), 4(G), 4(H)
				and S5(B)

# Supplementary Table S6. List of yeast strains used for the G $\gamma$ recruitment system for membrane target protein

Strain	Target	Candidate	Repoter GFP	Figure
	X (membrane)	Y (cytosol)		
FC-G0 (Control)	C-terminally membrane-associated Fc	-	EGFP	Fig. 3(B)
FC-GW	C-terminally membrane-associated Fc	Z <sub>WT</sub>	EGFP	Fig. 3(B)
FC-GK	C-terminally membrane-associated Fc	Z <sub>K35A</sub>	EGFP	Fig. 3(B)
FC-GI	C-terminally membrane-associated Fc	Z <sub>I31A</sub>	EGFP	Fig. 3(B)
FC-G9	C-terminally membrane-associated Fc	Z <sub>955</sub>	EGFP	Fig. 3(B)
FN-G0 (Control)	N-terminally membrane-associated Fc	-	EGFP	Fig. S4
FN-GW	N-terminally membrane-associated Fc	Z <sub>WT</sub>	EGFP	Fig. S4
FN-GK	N-terminally membrane-associated Fc	Z <sub>K35A</sub>	EGFP	Fig. S4
FN-GI	N-terminally membrane-associated Fc	Z <sub>I31A</sub>	EGFP	Fig. S4
FN-G9	N-terminally membrane-associated Fc	Z <sub>955</sub>	EGFP	Fig. S4
UG2-FCG0	C-terminally membrane-associated Fc	-	ymUkG1	Fig. 3(B)
UG2-FCGW	C-terminally membrane-associated Fc	Z <sub>WT</sub>	ymUkG1	Fig. 3(B)
UG2-FCGK	C-terminally membrane-associated Fc	$Z_{K35A}$	ymUkG1	Fig. 3(B)
UG2-FCGI	C-terminally membrane-associated Fc	Z <sub>I31A</sub>	ymUkG1	Fig. 3(B)
UG2-FCG9	C-terminally membrane-associated Fc	Z <sub>955</sub>	ymUkG1	Fig. 3(B)
UG2-FNG0	N-terminally membrane-associated Fc	-	ymUkG1	Fig. S4
UG2-FNGW	N-terminally membrane-associated Fc	Z <sub>WT</sub>	ymUkG1	Fig. S4
UG2-FNGK	N-terminally membrane-associated Fc	$Z_{K35A}$	ymUkG1	Fig. S4
UG2-FNGI	N-terminally membrane-associated Fc	Z <sub>I31A</sub>	ymUkG1	Fig. S4
UG2-FNG9	N-terminally membrane-associated Fc	Z <sub>955</sub>	ymUkG1	Fig. S4

Supplementary Table S7. Summary of the FACS analysis of the G $\gamma$  recruitment system for cytosolic target proteins. The percentage of cells belonging to GFP<sup>+</sup> were measured by flow cytometry.

Reporter GFP	Type of Z variant	Percentage of $GFP^{^+}$
EGFP	Z <sub>WT</sub>	58.8
EGFP	Z <sub>K35A</sub>	13.9
EGFP	Z <sub>I31A</sub>	2.5
EGFP	Z <sub>955</sub>	0
EGFP	Control	0
ymUkG1	Z <sub>WT</sub>	98.3
ymUkG1	Z <sub>K35A</sub>	95.4
ymUkG1	Z <sub>I31A</sub>	80.3
ymUkG1	Z <sub>955</sub>	0
ymUkG1	Control	0

#### Supplementary Table S8. List of primers

No.	Name	Sequence (5' to 3)
1	Sall-start-AcGFP1-fw	aaaagtcgacatggtgagcaagggc
2	BamHI-end-AcGFP1-rv	ttttggatcctcacttgtacagctcat
3	Sall-start-TagGFP2-fw	aaaagtcgacatgagcgggggggggggggggg
4	BamHl-end-TagGFP2-rv	ttttggatccttacctgtacagctcgtc
5	Sall-start-mUkG1-fw	aaaagtcgacatggtgagtgtgattaaa
6	BamHl-end-mUkG1-rv	ttttggatccttacttcgaagcctgact
7	Sall-start-EGFP(Yeast)-fw	aaaagtcgacatggtcagtaagggtgaa
8	BamHI-end-EGFP(Yeast)-rv	ttttggatccttacttgtataattcgtc
9	Sall-start-AcGFP1(Yeast)-fw	aaaagtcgacatggtttctaagggtgct
10	BamHI-end-AcGFP1(Yeast)-rv	ttttggatccttacttgtataattcgtc
11	Sall-start-TagGFP2(Yeast)-fw	aaaagtcgacatgtctggtggtgaagaa
12	BamHI-end-TagGFP2(Yeast)-rv	ttttggatcctcatctgtataattcgtc
13	Sall-start-mUkG1(Yeast)-fw	aaaagtcgacatggtcagtgtcatcaaaga
14	BamHI-end-mUkG1(Yeast)-rv	ttttggatccttacttagaagcttgagatg
15	Sall-start-ZsGreen(Yeast)-fw	aaaagtcgacatggctcaatccaaacat
16	BamHI-end-ZsGreen (Yeast)-rv	ttttggatccttatggcaaagcagaacc
17	Sall-start-mWasabi (Yeast)-fw	tagcgtcgacatggtgagcaagggcgagg
18	BamHI-end-mWasabi (Yeast)-rv	tagacccgggttacttgtataattcatccataccatcgg
19	Sall-start-mNeonGreen(Yeast)-fw	agcgtcgacatggtcagtaagggtgaaga
20	BamHI-end-mNeonGreen (Yeast)-rv	cgggggatccttacttgtataattcatc
21	Sall-atg-EGFP-fw	tagcgtcgacatggtgagcaagggcgagga
22	BamHI-end-frag-EGFP-rv	cgggggatccttacttgtcatcgtcatccttgtagtccttgtacagctcgtccatgccga
23	BamHI-end-frag-AcGFP1-rv	cgggggatcctcacttgtcatcgtcatccttgtagtccttgtacagctcatccatgccgt
24	BamHI-end-frag-TagGFP2-rv	cgggggatccttacttgtcatcgtcatccttgtagtccctgtacagctcgtccatgccgt
25	BamHI-end-frag-mUkG1-rv	cgggggatccttacttgtcatcgtcatccttgtagtccttcgaagcctgacttggcagca
26	Sall-atg-ZsGreen-fw	tagcgtcgacatggctcagtcaaagcac
27	BamHI-end-frag-ZsGreen-rv	cgggggatcctcacttgtcatcgtcatccttgtagtcgggcaatgcagatccggatgcaa
28	BamHI-end-frag-yEGFP-rv	cgggggatccttacttgtcatcgtcatccttgtagtccttgtataattcgtccataccca
29	BamHI-end-frag-yAcGFP1-rv	cgggggatccttacttgtcatcgtcatccttgtagtccttgtataattcgtccataccat

30	BamHI-end-frag-yTagGFP2-rv	cgggggatcctcacttgtcatcgtcatccttgtagtctctgtataattcgtccataccgt
31	BamHI-end-frag-ymUkG1-rv	cgggggatccttacttgtcatcgtcatccttgtagtccttagaagcttgagatggcaaca
32	BamHI-end-frag-yZsGreen-rv	cgggggatccttacttgtcatcgtcatccttgtagtctggcaaagcagaacctgaagcaa
33	BamHI-end-frag-ymWasabi-rv	cgggggatccttacttgtcatcgtcatccttgtagtccttgtataattcatccataccat
34	BamHI-end-frag-ymNeonGreen-rv	cgggggatccttacttgtcatcgtcatccttgtagtccttgtataattcatccataccca
35	Xhol-FIG1down-fw	ggggctcgagttttatcctcaaataaacat
36	Kpnl-FIG1down-rv	ccccggtaccaacagacggtaatgattaga
37	TFIG1hr40-URA3-fw	ttttatcctcaaataaacatataagttttgagcggatatttttttt
38	Xhol-URA3-rv	gaggataaaactcgaggggtaataactgatataatt
39	Sacll-FIG1end50-AcGFP1-fw	aaaaccgcggataggtacaataactactcttcggattcatctacattgcattccaaagttgtgagca
		agggcgccgagctgttcaccggc
40	AcGFP1-TFIG1hr40-rv	aatatccgctcaaaacttatatgtttatttgaggataaaatcacttgtacagctcatccatgccgtgg
		gt
41	Sacll-FIG1end50-TagGFP2-fw	aaaaccgcggataggtacaataactactcttcggattcatctacattgcattccaaagttagcggg
		ggcgaggagctgttcgccggc
42	TagGFP2-TFIG1hr40-rv	aatatccgctcaaaacttatatgtttatttgaggataaaattacctgtacagctcgtccatgccg
43	Sacll-FIG1end50-mUkG1-fw	aaaaccgcggataggtacaataactactcttcggattcatctacattgcattccaaagttgtgagtg
		tgattaaagagga
44	mUkG1-TFIG1hr40-rv	aatatccgctcaaaacttatatgtttatttgaggataaaattacttcgaagcctgacttggcagc
45	Sacll-FIG1end50-ZsGreen-fw	aaaaccgcggataggtacaataactactcttcggattcatctacattgcattccaaagttgctcagt
		caaagcacggtct
46	ZsGreen-TFIG1hr40-rv	aatatccgctcaaaacttatatgtttatttgaggataaaatcagggcaatgcagatccggatgca
47	Sacll-FIG1end50-EGFP(Y)-fw	aaaaccgcggataggtacaataactactcttcggattcatctacattgcattccaaagttgtcagta
		agggtgaagaattattcactggt
48	EGFP(Y)-TFIG1hr40-rv	aatatccgctcaaaacttatatgtttatttgaggataaaattacttgtataattcgtccataccc
49	Sacll-FIG1end50-AcGFP1(Y)-fw	aaaaccgcggataggtacaataactactcttcggattcatctacattgcattccaaagttgtttctaa
		gggtgctgaatt
50	AcGFP1(Y)-TFIG1hr40-rv	aatatccgctcaaaacttatatgtttatttgaggataaaattacttgtataattcgtccatacca
51	Sacll-FIG1end50-TagGFP2(Y)-fw	aaaaccgcggataggtacaataactactcttcggattcatctacattgcattccaaagtttctggtgg
		tgaagaattatt
52	TagGFP2(Y)-TFIG1hr40-rv	aatatccgctcaaaacttatatgtttatttgaggataaaatcatctgtataattcgtccataccg
53	SacII-FIG1end50-mUkG1(Y)-fw	aaaaccgcggataggtacaataactactcttcggattcatctacattgcattccaaagttgtcagtgt

		catcaaagaaga
54	mUkG1(Y)-TFIG1hr40-rv	aatatccgctcaaaacttatatgtttatttgaggataaaattacttagaagcttgagatggcaac
55	SacII-FIG1end50-ZsGreen(Y)-fw	aaaaccgcggataggtacaataactactcttcggattcatctacattgcattccaaagttgctcaat
		ccaaacatggttt
56	ZsGreen(Y)-TFIG1hr40-rv	aatatccgctcaaaacttatatgtttatttgaggataaaattatggcaaagcagaacctgaagca
57	FIG1end50-fw	ataggtacaataactactcttcggattcat
58	TFIG200-rv	aacagacggtaatgattagagtttaggtaa
59	Ste18pro-fw	atattatatatatagggtcgt
60	Ste18t-rv	aaattatagaaagcagtagataaaa
61	HIS3pro80-URA3-fw	tatataaagtaatgtgatttcttcgaagaatatactaaaaaatgagcaggcaagataaacgaagg
		caaagttcaattcatcattttttttttattctttt
62	HIS3t end40-rv	ggagccataatgacagcagttgggtaggcctttctttggt