

Supporting Information Fig. 1. Recombinant canstatin reduced the proliferation of HLMECs stimulated with rhVEGF-A. HLMECs (5 × 10⁴ cells/well in EBM-2 containing 2% FBS) were added to each well of gelatinized 24-well plates and treated with different concentrations (0, 0.1, 0.5, 1, 5, 10, 20, 40, 60 µg/ml) of recombinant canstatin in the presence of 20 ng/ml of rhVEGF-A for 48 h. The cells were trypsinized and counted using a hemocytometer. Data are presented as mean \pm S.D. of three independent experiments (*p < 0.05, **p < 0.01, ***p < 0.001).

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Supporting Information Fig. 2. Recombinant canstatin reduced the tube formation and migration of HLMECs stimulated with rhVEGF-A. (a) HLMECs in EBM-2 containing 2% FBS were added to Matrigelprecoated 48-well plates and treated with different concentrations (0, 0.5, 40 µg/ml) of recombinant canstatin in the presence of 20 ng/ml of rhVEGF-A. After 8 h of incubation, cells were imaged under a phase contrast inverted microscope using a digital camera. Scale bar = $200 \text{ }\mu\text{m}$. (b) Total tube lengths of HLMECs were quantified using the Image J program. (c) The cell densities of HLMECs under the same conditions as experiment (a) were determined after trypsinization, and are represented as a bar diagram. (d) HLMECs in EBM-2 containing different concentrations (0, 0.5, 40 μ g/ml) of recombinant canstatin were added to the upper chamber of a transwell insert with 8.0 µm pore-sized polycarbonate membranes that were coated with 0.1% gelatin. EBM-2 containing rhVEGF-A (20 ng/ml) was added to the lower chamber to induce cell migration. After 24 h of incubation, cells that migrated to the undersides of the inserts were fixed with methanol, stained with Harris hematoxylin solution, and imaged under a phase contrast inverted microscope using a digital camera. Scale bar = $200 \ \mu m$. (e) Five digital images per well were obtained in experiment (d), and the numbers of migrated HLMECs were counted. Each sample was assayed in duplicate. The numbers of migrated HLMECs present in 320 mm² are presented as a bar diagram. (f) The cell densities of HLMECs under the same conditions as in experiment (d) were determined after trypsinization, and are represented as a bar diagram. Data are presented as mean \pm S.D. of three independent experiments (*p < 0.05, **p < 0.01, ***p< 0.001).



Supporting Information Fig. 3. The phosphorylation of VEGFR-1 and -2 by VEGF-A is reduced by the presence of anti- $\alpha\nu\beta3$ and recombinant canstatin. (*a*) HLMECs were treated with anti- $\alpha\nu\beta3$ (2 µg/ml) in the presence or absence of 20 ng/ml VEGF-A and recombinant canstatin (0.5 or 40 µg/ml). Cell lysates were immunoprecipitated with anti-phospho-Tyr (anti-p-Tyr) antibody. The presence of p-VEGFR-1 and p-VEGFR-2 in immunoprecipitates was detected via western blot analysis using anti-VEGFR-1 and anti-VEGFR-2 antibodies. (*b*) The amounts of p-VEGFR-1 and p-VEGFR-2 determined in three independent experiments of (*a*) were quantified and are represented as a bar diagram. The levels of the p-VEGFR-1 and p-VEGFR-2 of recombinant canstatin- and rhVEGF-A-untreated cells were established as 100%. Data are presented as mean ± S.D. of three independent experiments (p < 0.05, p < 0.01, p < 0.001).

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