

**New Phytologist Supporting Information Figs S1–S6 and Tables S1 & S2**

Article title: Multifunctional oxidosqualene cyclases and cytochrome P450 involved in the biosynthesis of apple fruit triterpenic acids

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**Fig. S1** Chromatograms of typical LC-APCI-MS analysis of the products of MdOSC3.

**Fig. S2** Expression analysis of triterpene biosynthetic genes (MdOSC1, MdOSC3, MdOSC4, MdOSC5) by real-time qPCR on RNA extracted from apple skin tissues.

**Fig. S3** Mass spectra of lupeol, germanicol,  $\beta$ -amyrin, and  $\alpha$ -amyrin.

**Fig. S4** Chromatographic trace of extract of *Nicotiana benthamiana* leaf transiently transformed with MdOSC4 and the same sample spiked with taraxerol.

**Fig. S5** Alignment of P450 predicted amino acid sequences from apple (CYP716A175) and other species.

**Fig. S6** Mass spectra comparison (MS1, MS2, and MS3) between betulinic acid, ursolic acid, oleanolic acid, and a putative morolic acid.

**Table S1** Primer sequences and properties of triterpene-related and housekeeping genes in this study.

**Table S2** Summary of the skin triterpene composition of 20 apple cultivars

**Table S1** Primer sequences and properties of triterpene-related and housekeeping genes used to (a) isolate oxidosqualene cyclases and (b) used on the panel of 20 apple cultivars

**(a) Primers used to isolate Oxidosqualene Cyclases**

Primer name	Sequence
attB1 F_OSC4_MR	GGGGACAAGTTGTACAAAAAAAGCAGGCTCCATGTGGAAGCTTAAGGTGCG
attB2 R_OSC4_MR	GGGGACCACTTGTACAAGAAAGCTGGGTCTAACGCTTGGAAGG
attB1 F_OSC5_RG	GGGGACAAGTTGTACAAAAAAAGCAGGCTCCATGTGGAAGCTTAAGGTGCG
attB2 R_OSC5_RG	GGGGACCACTTGTACAAGAAAGCTGGTTAGACATCAACAAATGGAACCC

**(b) Primers used to measure gene expression by PCR**

Genes	Accession Number	Primer Sequences (5'-3')	Size (bp)	T <sub>m</sub> ( °C)	E	References
eF-1 $\alpha$	AJ223969.1	F: ACTGTCCTGTTGGACGTGTTG  R: GAGTTGGAAGCAACGTACCC	208	82.5	1.861	Giorno <i>et al.</i> (2012)

IMPA-9	CN909679	F : TCGTGAACTCAGGCGCTTACTG  R : AAGCAACGGTAAAGCGGGCAAC	205	83.5	1.953	Giorno <i>et al.</i> (2012)
MdGAPDH	EB146750	F : TGAGGGCAAGCTGAAGGGTATCTT  R: TCAAGTCAACCACACGGGTACTGT	185	82.5	1.874	Malladi & Hirst (2010)
MdACTIN	EB127077	F: ACCATCTGCAACTCATCCGAACCT  R: ACAATGCTAGGGAACACGGCTCTT	185	82	1.919	Malladi & Hirst (2010)
MdOSC1	FJ032006.1	F : 238 TTGTACTACTAATCCAGTGATCAAGATGTGG  R : CTCTCTTAGTATCTGAAAACGCCATAGGAG	238	84	1.857	Brendolise <i>et al.</i> (2011)
MdOSC3	FJ032008.1	F : GCAATCGTGATCAAAGAAGATGTGGAGG  R : TTCTCTTAAAATCTGAAAACGCCATAGG	232	84	1.882	Brendolise <i>et al.</i> (2011)
MdOSC4	KT383435	F : CTCTCGAAGTAACAATGAAGCACA	124	79	1.864	

R : TAATCACCATTGGGTCTTCAG

MdOSC5 KT383436 F : TATGCATCCAGCAAAATGTTT 152 77.5 1.851

R : TCCAATTAATTGCCATAAGGT

CYP716A17 EB148173 F : AGGCACGTTCCCTCGCTTC 70 77.5 1.974

5

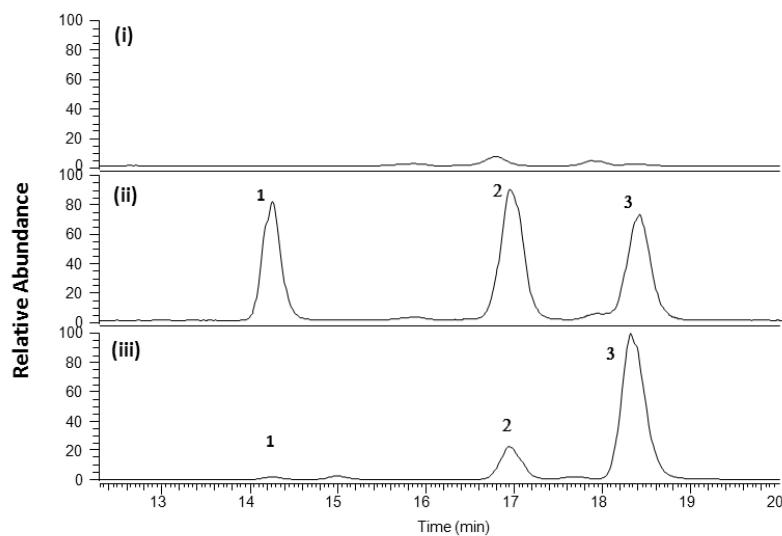
R : CAAACCCTAACAGAGGAGGGTCA

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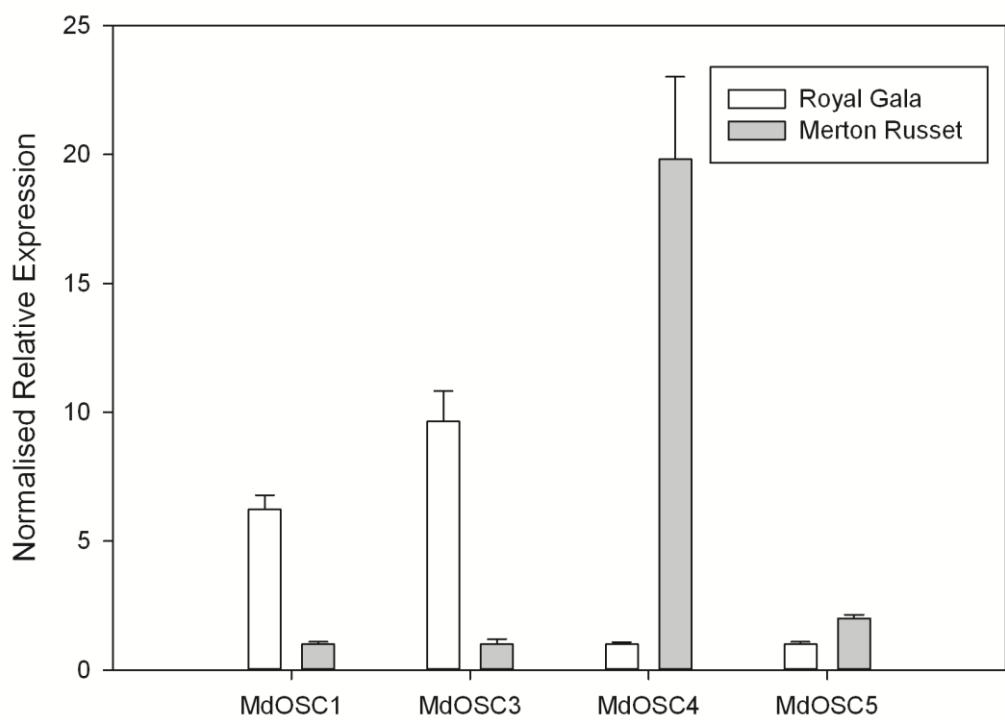
**Table S2** Summary of the skin triterpene composition of 20 apple cultivars separated into three russetting groups: russeted (four cultivars), semi-russeted (nine cultivars), and waxy (seven cultivars)

Total Triterpenes	Triterpene Acids (nmol g <sup>-1</sup> DW)			Triterpene-caffeates (nmol g <sup>-1</sup> DW)		
	Ursolic acid	Oleanolic acid	Betulinic acid	Oleanolic acid -3-trans-caffeate	Betulinic acid -3-trans-caffeate	Betulinic acid -3-cis-caffeate
<b>Russeted</b>						
Average	6899 <sup>a</sup>	1569 <sup>a</sup>	800 <sup>a</sup>	1628 <sup>b</sup>	416 <sup>b</sup>	2115 <sup>c</sup>
Min–Max	4971–8075	670–2802	500–1270	1094–2095	248–511	1287–3722
<b>Semi-russeted</b>						
Average	5567 <sup>a</sup>	1981 <sup>ab</sup>	1515 <sup>b</sup>	1028 <sup>ab</sup>	205 <sup>b</sup>	693 <sup>b</sup>
Min–Max	4228–7636	1464–2504	991–1974	527–1712	72–406	291–1312
<b>Waxy</b>						
Average	5864 <sup>a</sup>	2839 <sup>b</sup>	2045 <sup>b</sup>	687 <sup>a</sup>	90 <sup>a</sup>	159 <sup>a</sup>
Min	3562–7379	1680–3816	1203–2540	324–834	23–175	82–252
25–73						

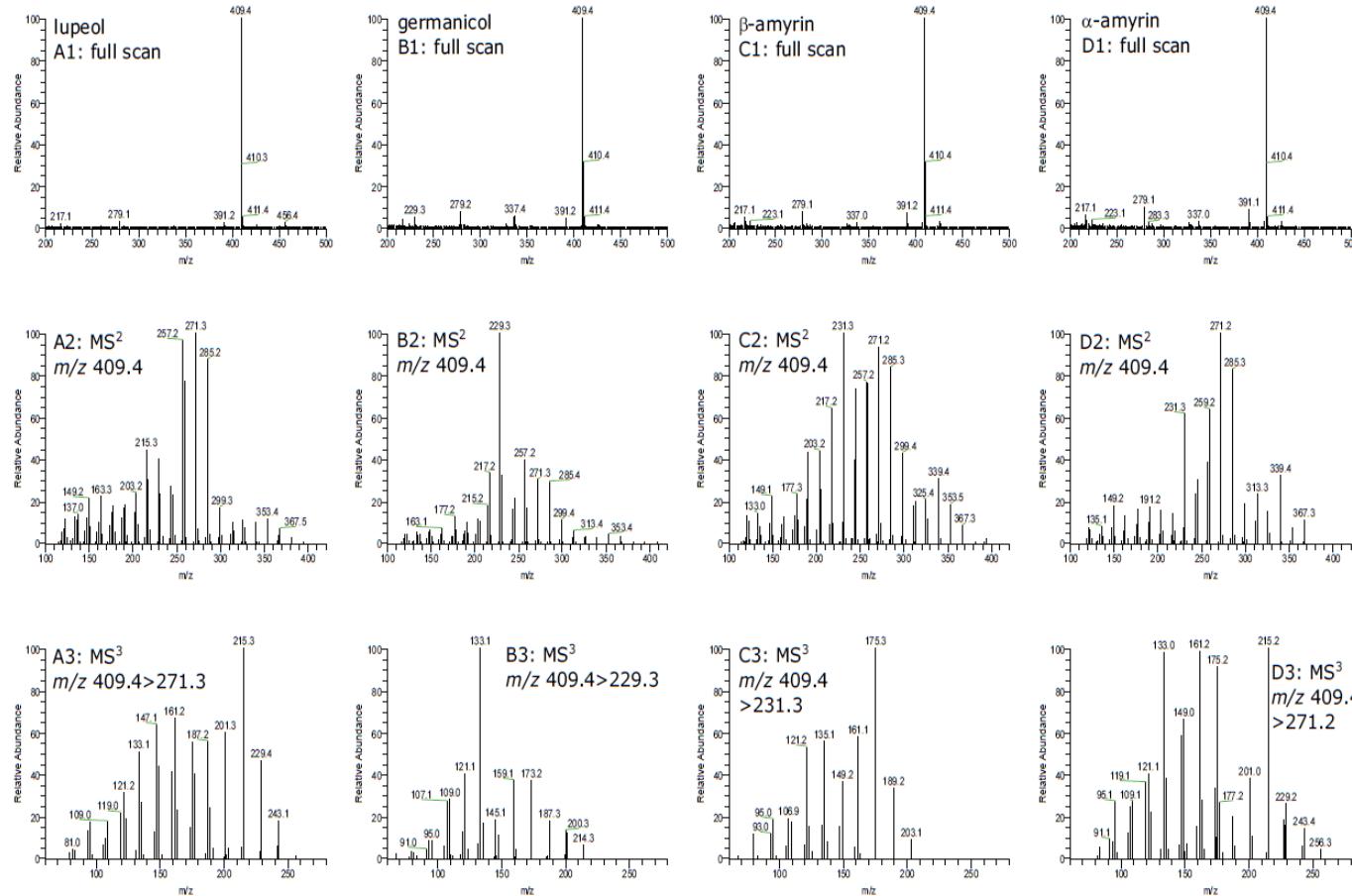
Data are expressed in nmol g<sup>-1</sup> DW. Data were obtained after HPLC-DAD analysis as described in the Materials and Methods section. Averages with no letter in common within a column are significantly different ( $P < 0.05$ ; One-way ANOVA, Tukey's pairwise results).



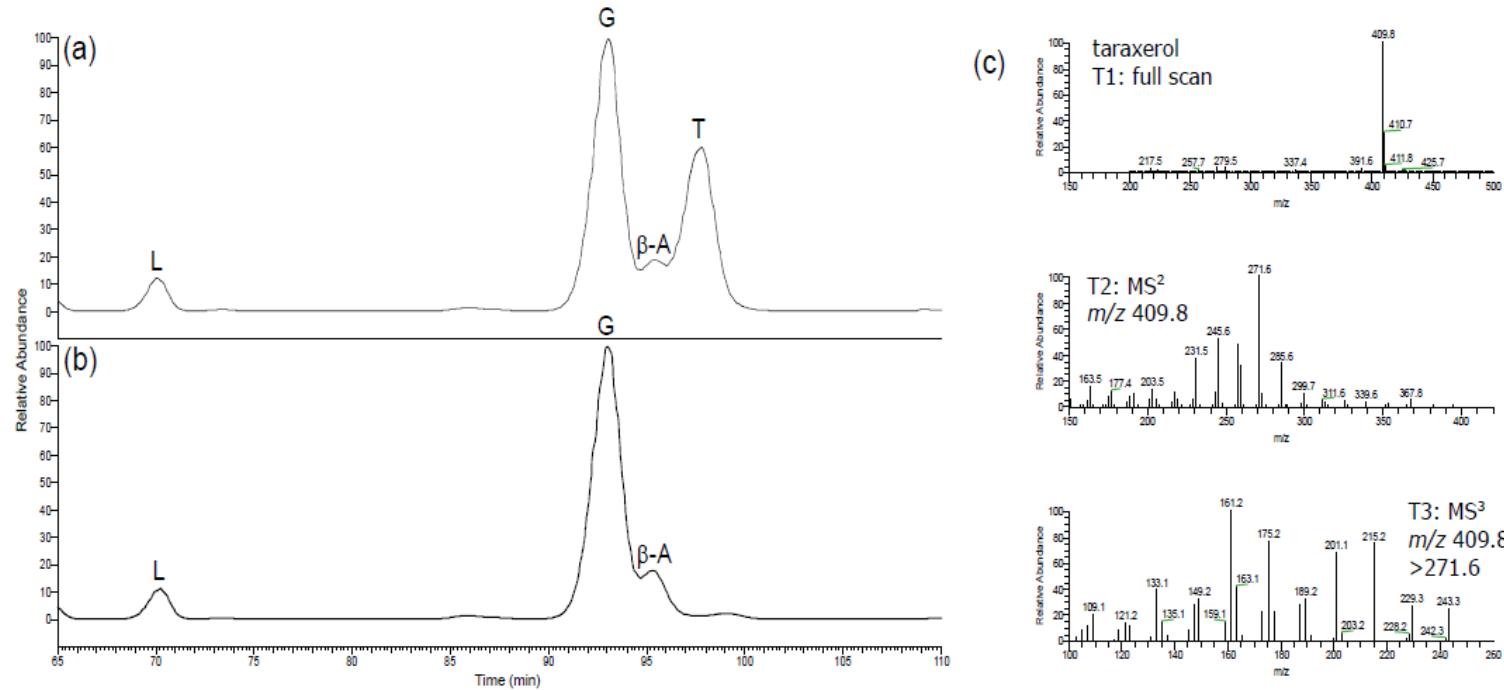
**Fig. S1** Chromatograms of typical LC-APCI-MS analysis of the products of MdOSC3 (iii) after transient expression in *Nicotiana benthamiana*, following the methodology described in the Materials and Methods section. p19 was used as a negative control (i) and mixed with authentic standards at 20 µg ml<sup>-1</sup> (1, lupeol; 2, β-amyrin; 3, α-amyrin) (ii). Compounds were identified and quantified on the basis of their mass spectral data (Fig. S3). Chromatograms are presented as selected ion plots of the m/z 409.8 [MH-H<sub>2</sub>O]<sup>+</sup> ion.



**Fig. S2** Expression analysis of triterpene biosynthetic genes (MdOSC1, MdOSC3, MdOSC4, MdOSC5) by real-time qPCR on RNA extracted from apple skin tissues. Error bars show SE ( $n = 3$ ).

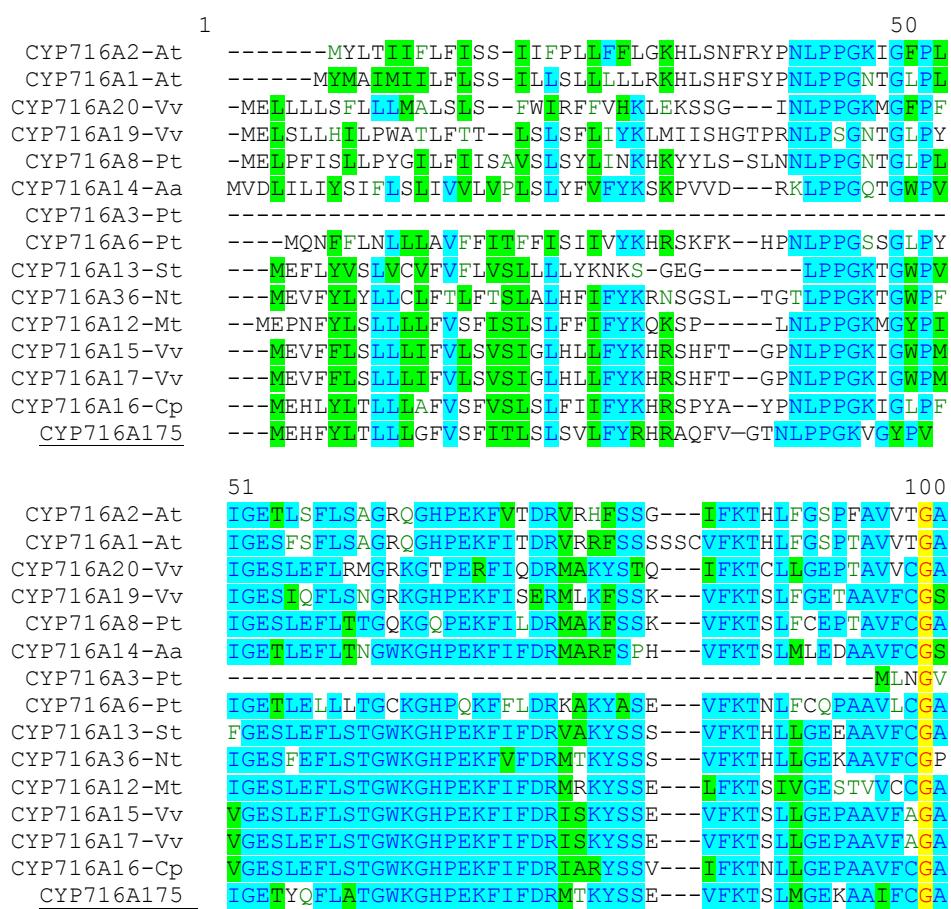


**Fig. S3** Mass spectra comparison (MS1, MS2, and MS3) between lupeol, germanicol,  $\beta$ -amyrin, and  $\alpha$ -amyrin.



**Fig. S4** Chromatographic trace of extract of *Nicotiana benthamiana* leaf transiently transformed with MdOSC4 (b) and the same sample spiked with taraxerol (T) (a) showing no taraxerol in the leaf sample. Mass spectra of the taraxerol peak (MS1, MS2, and MS3) (c). Transient expression of MdOSC4 (b) led to the formation of germanicol (G), β-amyrin (β-A), and lupeol (L). The presence of taraxerol (T) could therefore be excluded.

**Fig. S5** Alignment of P450 predicted amino acid sequences from apple (CYP716A175) and other species. The prefix is the cytochrome P450 designation, and the suffix identifies the species as follows: At, *Arabidopsis thaliana*; Aa, *Artemisia annua*; Mt, *Medicago truncatula*; Vv, *Vitis vinifera*; Cp, *Carica papaya*; Pt, *Populus trichocarpa*; St, *Solanum tuberosum*; Nt, *Nicotiana tabacum*. GenBank (GB) numbers (or Arabidopsis identifiers) in the order listed in the alignment are: AT5G36140 (CYP716A2-At), AT5G36110 (CYP716A1-At), XP\_002264643 (CYP716A20-Vv), XP\_002280969.1 (CYP716A19-Vv), XP\_002309057.1 (CYP716A8-Pt), ABC94483.1 (CYP716A14-Aa), XP\_002324668.2 (CYP716A3-Pt), XP\_002325241 (CYP716A6-Pt), XP\_006338129 (CYP716A13-St), no GB identifier (CYP716A36-Nt), CBN88268.1 (CYP716A12-Mt), BAJ84106 (CYP716A15-Vv), BAJ84106 (CYP716A17-Vv), no GB identifier (CYP716A16-Cp). The text/background colour code refers to the degree of similarity between the different sequences: red/yellow, identical; dark blue/turquoise, conservative; black/green, block of similar; black/white, nonsimilar.



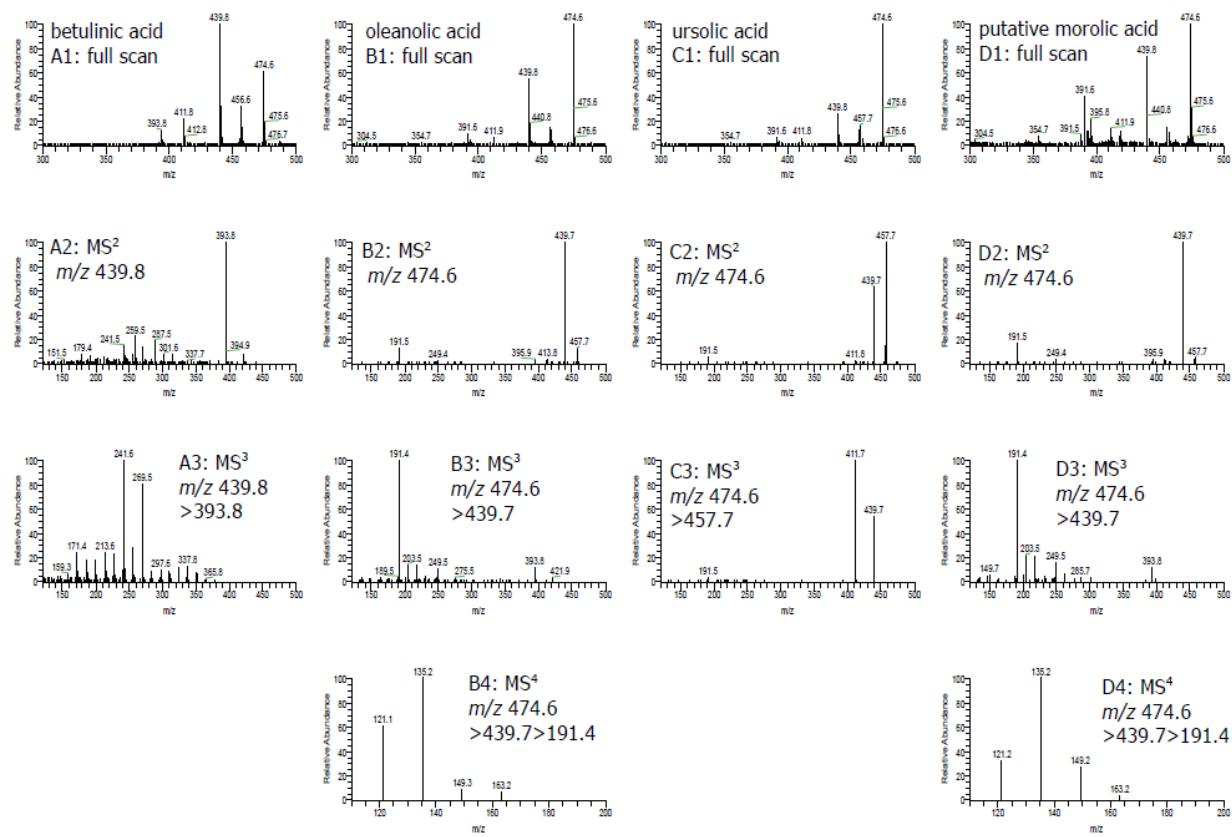
CYP716A2-At	SGNKELEFTNEKLVISWWPDSVNKTIFPSSSTQTSSK-EEAIKTRMILLMPSM
CYP716A1-At	SGNKELEFTNEKLVVSWWPDSVNKTIFPSSMQTSSK-EEARKLRRMILLSQFM
CYP716A20-Vv	AGNKELEFSNENKLVTTSWWPDSVNKTIFPSSLQTSSK-EESMKTFRKILLPAFL
CYP716A19-Vv	AGNKELEFSNENKLVTAWWPSSVNKTIFPSSLQTSSQ-EESKKMRKLLPGFL
CYP716A8-Pt	AGNKELEFSNENKLVTAWWPDSVNKTIFPSSQQTSSQ-EESKKMRKLFPLFF
CYP716A14-Aa	AGNKELEFSNENKLVKAWWPASVEKILPSAKETINQ-----RKMLSRSF
CYP716A3-Pt	AGNKELEFSNENRLVETWWPEFVNKTIFPSSAVEKSPK-EEAKRMRRLFPPFL
CYP716A6-Pt	AGNRFLEFSNKNKLVAWYPDFVCKTIFPSSVORPLI-EQVDRLRRTLLPELL
CYP716A13-St	SGNKELEFSNENKLVQAWWPNSVNKVFEESSTQTSSK-EEAIKMRKMLPNFF
CYP716A36-Nt	TCNKELEFSNENKLVQAWWPDSVNKVFEESSTQTSSK-EEAIKMRKMLPNFF
CYP716A12-Mt	ASNKELEFSNENKLVTAWWPDSVNKTIFPTTSLDNSNLKEESIKMRKLLPQFF
CYP716A15-Vv	AGNKELEFSNENKLVHAWWPSSVDKVFEESSTQTSSK-EEAKKMRKLLPQFF
CYP716A17-Vv	AGNKELEFSNENKLVHAWWPSSVDKVFEESSTQTSSK-EEAKKMRKLLPQFF
CYP716A16-Cp	ACNKELEFSNENKLVTAWWPNSVNKTIFPTSLQTSSI-EESKKMRKLLPQFL
<u>CYP716A175</u>	ACNKELEFSNENKLVTAWWPSSVNKVFEESSTLETSAK-EEAKKMRKMLPNFM

151	200
CYP716A2-At	KPEALRRYVGVMDEIAQKHFEETEWANQDQLIVFPLTKKETFSIACRLFIS
CYP716A1-At	KPEALRRYVGVMDEIAQRHFEETEWANQDQVIVFPLTKKETFSIACRSFIS
CYP716A20-Vv	KPEALDKYVGIMDSIAKWHLDNHWDLNFTVTFPLAKOYTFMVACRLFIS
CYP716A19-Vv	KPEALQRYISIMDVIAQRHFEESWWNNKEEVTVFPLAKMFTFWLACRLFIS
CYP716A8-Pt	RPESLQRYISVMDVIAQRHLASDWEGRQEVSVFPLAKTYTFWLACRLFIS
CYP716A14-Aa	RPESLQRQYVPVMDMAQRHFKTEWLGMDQIVTHEVTQNFTFSLACKIFVS
CYP716A3-Pt	KPEALRRYIGTMDMVTKRHFALEWGNAEVVVFPLAKSYTFELACRLFIS
CYP716A6-Pt	RPDALRKYVGIFDKAAGRHFASEWEWKVVVFPLAKRTFGLACSLFIS
CYP716A13-St	KPEALQRYVGIMDHITQRHFASGWENKEQVVVFPLTKRYTFWLACRLFIS
CYP716A36-Nt	KPEALQRYVGIMDHIAQRHFASSWENKNQEVEFPLAKRYTFWLACRLFVS
CYP716A12-Mt	KPEALQRYVGVMDVIAQRHFTVTHWDNKNEITVYPLAKRYTFLLACRLFMS
CYP716A15-Vv	KPEALQRYIGIMDHIAQRHFADSWDNRDEVVFPLAKRTFWLACRLFMS
CYP716A17-Vv	KPEALQRYIGIMDHIAQRHFADSWDNRDEVVFPLAKRTFWLACRLFMS
CYP716A16-Cp	KPEALQRYIGIMDGIAQRHFESGWENKEEVKVFPLAKSYTFWIACRLFMS
<u>CYP716A175</u>	KPEALQRYIGIMDTVARRHFAEGWENKEEVVFPLAKNYTFWLAARLFVS

201	250
CYP716A2-At	MDDLERVVKLEEPEFTTVMTCVFSIEIDIPGIRFNRAIKASRLLSKEVSTI
CYP716A1-At	MEDPARVRQLEEQFNTVAVGIFSIETIDIPGIRFNRAIKASRLLRKEVSAT
CYP716A20-Vv	IDDPKHIAKFANPFHILAAAGVMSIEINFEGIPFNRAIKAADSVRKELRAI
CYP716A19-Vv	VEDPDHVEKLAEPFNELAAGIATIDIPGICISFNKGIAASNLVRKELHAI
CYP716A8-Pt	MEDEEVQKFAKPFDNLAAAGISIEIDIPWIPFNRGVKASNVVHKELLKI
CYP716A14-Aa	IEDPEEVKHLGSGPFEKFAPGIFSIEIDIPWIPLRAIHAGNFIRKEIIIAI
CYP716A3-Pt	IEDPSHIAFRSHFPFHNIITSGIFTIEIAFECPFPNRAIKATKLIRIELLAI
CYP716A6-Pt	IEDPDHIAKLASPFLNLVVGSIFSIEIDIPGCTPLSRAIKASTIIRTELFAI
CYP716A13-St	VEDPNHVAKFADPFDVLASGLISIEIDIPGCTPFNRAIKASNFINKELVRI
CYP716A36-Nt	VEDPNHVAKFADPFDVLASGLISIEIDIPGCTPFNRAIKASNLRKEILLI
CYP716A12-Mt	VEDENHVAKFSDPFLQIAAGIISIEIDIPGCTPFNRAIKASNFINKELIK
CYP716A15-Vv	IEDPAHVAKEEKPFHVLASGLITVPEIDIPGTPFHRAIKASNFINKELR
CYP716A17-Vv	IEDPNHVERFAEFPFHHLASGVISIEIDIPGCTAFNRGIKAASNFIRKEELLKI
CYP716A16-Cp	IEDSVETAKLGDPFAVLASGISMPLIDFPGTIFYKAIKASNFINKEELTKI
<u>CYP716A175</u>	

251	300
CYP716A2-At	IQRKEEELKAQKVSVE-QDILSHMILMNIGET----KDEDLADKIIALLIG
CYP716A1-At	VRQRKEEELKAQKALEE-HDILSHMILMNIGET----KDEDLADKIIIGLLIG
CYP716A20-Vv	IKQRKIQVLAGKSSSSKHDILSHMILTITDENGQFQNEEMDIADKILGLLIG
CYP716A19-Vv	IKQRKMNADNKASTT-QDILSHMILTCDENGEYMEEDIADKILGLLIG
CYP716A8-Pt	IKQRKIDLAENKASPT-QDILSHMILTTADDNGQCMKKIDIADKILGLLIG
CYP716A14-Aa	IKQRKIDLAEGKASPT-QDILSHMILCDEEISON--IAEADTADVIIIGLLIG
CYP716A3-Pt	IKQRKIDLAEGKASPT-QDILSHMILSNDANGQYMEVEIADKIIALLIG
CYP716A6-Pt	IKQRKIDLAEGKASPK-QDILSHMILAC-DEKGAFMSELDIADTIIALLAS
CYP716A13-St	IKQRKIDLAEGKASPT-QDILSHMILTCDENKGFMGDLADIADKILGLLIG
CYP716A36-Nt	IKQRKVDLAEGKASPT-QDILSHMILTSDENGYMHLDIADKILGLLIG
CYP716A12-Mt	IKQRKIDLAEGTASPT-QDILSHMILTSDENGSKSMNELNADIADKILGLLIG
CYP716A15-Vv	IKQRKIDLAEGKASQN-QDILSHMILATDEDGCHMNEMEIADKILGLLIG
CYP716A17-Vv	IKQRKIDLAEGKASQN-QDILSHMILTSDESGQFMSLDIADKILGLLIG
CYP716A16-Cp	IKQRKIDLAEGKASPT-QDILSHMILLCDEHGSHMKEHDIADKILGLLIG
<u>CYP716A175</u>	

CYP716A2-At		
CYP716A1-At		
CYP716A20-Vv		
CYP716A19-Vv		
CYP716A8-Pt		
CYP716A14-Aa		
CYP716A3-Pt		
CYP716A6-Pt		
CYP716A13-St		
CYP716A36-Nt		
CYP716A12-Mt		
CYP716A15-Vv		
CYP716A17-Vv		
CYP716A16-Cp		
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		351
CYP716A2-At		400
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CYP716A19-Vv		
CYP716A8-Pt		
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CYP716A3-Pt		
CYP716A6-Pt		
CYP716A13-St		
CYP716A36-Nt		
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CYP716A17-Vv		
CYP716A16-Cp		
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CYP716A1-At		
CYP716A20-Vv		
CYP716A19-Vv		
CYP716A8-Pt		
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CYP716A6-Pt		
CYP716A13-St		
CYP716A36-Nt		
CYP716A12-Mt		
CYP716A15-Vv		
CYP716A17-Vv		
CYP716A16-Cp		
<u>CYP716A175</u>		
		451
CYP716A2-At		502
CYP716A1-At		
CYP716A20-Vv		
CYP716A19-Vv		
CYP716A8-Pt		
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CYP716A3-Pt		
CYP716A6-Pt		
CYP716A13-St		
CYP716A36-Nt		
CYP716A12-Mt		
CYP716A15-Vv		
CYP716A17-Vv		
CYP716A16-Cp		
<u>CYP716A175</u>		



**Fig. S6** Mass spectra comparison (MS1, MS2, and MS3) between betulinic acid, ursolic acid, oleanolic acid, and a putative morolic acid.

## References

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