Table S1. List of *C. albicans* strains, plasmids and primers used in the study

A. Yeast strains used in this study

Strain	Genotype	Source
SC5314	Wild type diploid	(1)
BWP17	*ura3::imm434/ura3::imm434 his1::hisG/his1::hisG arg4::hisG/arg4::hisG*	(2)
GZY792	*MTLa his4*	(3)
GZY803	*MTLa his4 ura3Δ::HIS4*	(3)
GZY923	*ura3::imm434/ura3::imm434 his1::hisG/his1::hisG arg4::hisG/arg4::hisG ira2Δ::HIS1/ira2Δ::FRT*	(4)
GZY1022	*ura3::imm434/ura3::imm434 his1::hisG/his1::hisG arg4::hisG/arg4::hisG ira2Δ::HIS1/ira2Δ::FRT IRA2-Myc-URA3*	(4)
GZY1162	*ira2Δ/Δ* (GZY923)+TetOff-Myc-AHP1	This study
GZY1163	GZY803+TetOff-Myc-AHP1	This study
GZY1166	BWP17 :: AHP1-Myc	This study
GZY1167	*ira2Δ/Δ* (GZY923) :: AHP1-Myc	This study
GZY1168	GZY803 :: AHP1-Myc	This study
GZY1210	*ura3::imm434/ura3::imm434 his1::hisG/his1::hisG arg4::hisG/arg4::hisG ahp1Δ::HIS1/ahp1Δ::UFP*	This study

B. Plasmids constructed in this study

Construct	Description
CIP10U	*Candida albicans* integration vector with *URA3* as the selection marker; generated by replacing the *RP10* gene in the vector CIp10 (5) with 700 bp *GAL1* untranslated region (UTR) at *Cla*I and *Pst*I sites.
pYGS1273	TetOff-Myc-AHP1-UTR*-TetR/CIP10U; The coding region of *AHP1* gene (1 to 528 bp) was PCR amplified and cloned (between *Cla*I and *Pac*I) in frame after an N-terminal 6x*Myc* epitope which is controlled by a tetracycline repressible promoter (6), and before *UTR* which is followed by TetR, into CIP10U. The plasmid was linearized by *Asc*I (generated by site-directed mutagenesis) within UTR* for integration to generate TetOff-Myc-AHP1.
pYGS1276	AHP1c-Myc-UTR/CIP10U; The partial region of *AHP1* gene (28 to 528 bp) was PCR amplified and cloned (between *Kpn*I and *Xho*I) in frame in front of an N-terminal 6x*Myc* epitope (between *Xho*I and *Cla*I), followed by *UTR* (between *Cla*I and *Pst*I), into CIP10U. The plasmid was linearized by *Hind*III within *AHP1* for integration to generate AHP1-Myc.
pYGS1307	AHP1Δ::UFP/pBKS; *AHP1* promoter region (~500 bp) and terminator region (~450 bp) were amplified by PCR and cloned into the vector pBKS at *Kpn*I-*Xho*I and *Not*I-*Sac*II sites, respectively, to flank the *URA3* flipper (7) (UFP) located between *Xho*I and *Not*I. The knock-out cassette was released by *Kpn*I and *Sac*II for transformation to generate *ahp1Δ::UFP*.
pYGS1308	AHP1Δ::HIS1/pBKS; *HIS1* was amplified by PCR and cloned into pYGS1307 at *Xho*I-*Not*I to replace UFP. The knock-out cassette was released by *Kpn*I and *Sac*II for transformation to generate *ahp1Δ::HIS1*.

C. Primers used in this study

Gene name	Primer sequence	Reference
ACT1	Forward: 5'-GCTTTTGGTGTTTGACGAGTTTCT-3' Reverse: 5'-GTGAGCCGGGAAATCTGTATAGTC-3'	(8)
PMA1	Forward: 5'-TTGCTTATGATAATGCTCCATACGA-3' Reverse: 5'-TACCCCACAATCTTGGCAAGT-3'	(9)
AHP1	Forward: 5'-CTGCTGTGCCTGGTGCTT-3' Reverse: 5'-TTGACGCCCTTGTCTTTGA-3'	(10)
PRX1	Forward: 5'-GTGTGTAGCACCGAGCTTTCTGCGTTC-3' Reverse: 5'-TCCAGTCGGAATTTGCTTCAACAGGGT-3'	(11)
DOT5	Forward: 5'-CCAAAGAACCAGAGGCTGTCACCGAAC-3' Reverse: 5'-AATTTTTTCACCAATCCCCAATCCAGCA-3'	(11)
PCK1	Forward: 5'-AAGGGGTTGAAAAAGGTGATGTC-3' Reverse: 5'-ATGCGGAGAATGTAGCGTGTG-3'	(12)
TSA1	Forward: 5'-TCCAAAGGGTGTCTTGAGACAAATCACCAT-3' Reverse: 5'-TTCCTTGGATGCTTCTGGGCTTGG-3'	(11)
TUP1	Forward: 5'-CAAAATCTTGAGGGGCCACGAAC-3' Reverse: 5'-CTAAGGACCCGGAAGCGATTTGTT-3'	(12)
TRR1	Forward: 5'-TGGACTGAATGGAATGAAGATGCAGAACC-3' Reverse: 5'-GTCACAAACAGCACAGGCAGAAATACCTTG-3'	(11)