SUPPLEMENTARY INFORMATION

Mitochondrial Ca²⁺ uptake controls cytoskeleton dynamics during cell migration

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Supplementary Figure S1 related to Figure 1. *Mcu*-silencing in Hs578t cells alters mitochondrial Ca²⁺ uptake, but not apoptosis or cell proliferation.

a. Representative traces of the relative changes in the mitochondrial Rhod-2AM fluorescence intensity normalized to the baseline (F/F₀ ratio) reflecting the mitochondrial Ca²⁺ rise following histamine (100 μ M) stimulation in Hs579t cells silenced or not with the 2 *mcu*-siRNA.

b. Histogram depicting the maximum mitochondrial Ca^{2+} increase (F_{max}/F_0 ratio) in Hs578t cells following histamine (100 μ M) treatment relative to control, from Fig S1a. (mean \pm S.E.M; three independent experiments). ***P < 0.001.

c. Representative traces of the relative changes in the Oregon Green-5N fluorescence intensity normalized to the baseline (F/F₀ ratio), reflecting the mitochondrial Ca²⁺ uptake following calcium (20 μ M) stimulation from isolated mitochondria of Hs579t cells silenced or not with the 2 *mcu*-siRNA.

d. Histogram depicting the mitochondrial Ca^{2+} uptake (F_{max}/F_0 ratio) of isolated mitochondria from si-*mcu* treated Hs578t cells following Ca^{2+} (20 μ M) boost relative to control, from Fig S1c. (mean ± S.E.M; three independent experiments). ***P < 0.001.

e. Histogram showing the gap closure of Ru360 (60μ M) + probenecid (2.5 mM) treated Hs578t cells relative to control (probenecid treated only) (mean ± S.E.M; three independent experiments). ***P < 0.001.

f. Histogram showing the Caspase 3/7 signal to confluence ratio for Hs578t cells silenced with the 2 *mcu*-siRNA (mean ± S.E.M; n=3 independent experiments). N.S.= not significant.

g. Histogram showing the proliferation rate relative to control for Hs578t cells silenced with the 2 mcu-siRNA (mean \pm S.E.M; n=3 independent experiments). N.S.= not significant; ***P < 0.001.



Supplementary Figure S2 related to Figure 2. Stable sh*mcu* silencing in Hs578t cells compromises cell migration as well as mitochondrial Ca²⁺ uptake.

a. Representative single cell tracking experiments highlighting the cell paths (blue line) of isolated Hs578t (red dots) stably expressing sh Control, sh1 or sh2 MCU shRNAs; acquisition time 24 hours.

b. Histogram deduced from the results displayed in Fig. S2a showing the distance of cell migration relative to control (mean \pm S.E.M; 30 cells in each experiment; n=3 independent experiments). ***P < 0.001.

c. Representative traces of the relative changes in the fluorescence intensity normalized to the baseline (F/F₀ ratio) reflecting the mitochondrial Ca²⁺ rise following histamine (100 μ M) stimulation. The mitochondrial Ca²⁺ increase was detected using the mitochondrial localized CEPIA2mt Ca²⁺ sensor transiently expressed in Hs578t cells. *Mcu* knockdown using specific

shRNA (sh1 and sh2) significantly decreased the mitochondrial calcium uptake compared to control cells transfected with negative control shRNA (sh Control).

d. Histogram depicting the maximum mitochondrial Ca^{2+} increase (F_{max}/F_0 ration) in Hs578t cells following histamine (100 μ M) treatment, from Fig S2c (mean \pm S.E.M; n=3 independent experiments). ***P < 0.001.



Supplementary Figure S3 related to Figure 3. Mitochondrial Ca²⁺ uptake impairment does not lead to microtubules and intermediate filaments modifications.

Representative confocal images of sh Control and sh2 MCU Hs578t cells stained for intermediate filaments and microtubules using anti-vimentin and anti-tubulin antibodies, respectively. Intermediate filaments and microtubules organization was not affected in the *mcu*-silenced cells compared to controls. Cells were stained additionally with Hoechst 33342 (blue) dye. Scale bar: $20 \mu m$.



Supplementary Figure S4 related to Figure 4. Mitochondrial Ca²⁺ uptake impairment leads to SOCE default but not to ATP production loss.

a. Raw data of Oxygen Consumption Rate (OCR) determined by the SeaHorse XF24 analyser. OCR was measured continuously in Hs578t expressing sh Control, sh1 MCU or sh2 MCU shRNAs throughout the experimental period at baseline followed by the addition of the indicated

drugs. A: Oligomycin (0.5 μ M), B: FCCP (1 μ M), C: Rotenone (0.5 μ M), D: Antimycin A (0.5 μ M). OCR was normalized to protein level. (mean ± S.E.M; n=3 independent experiments).

b. Histogram depicting basal respiration calculated from the first baseline mean (pmol/min/10 μ g protein) in Hs578t cells stably expressing sh Control, sh1 MCU or sh2 MCU shRNAs (mean \pm S.E.M; n=3 independent experiments). N.S.= not significant.

c. Total ATP levels in Hs578t cells stably expressing sh Control, sh1 MCU or sh2 MCU shRNAs. (mean ± S.E.M; n=3 independent experiments). N.S.= not significant.

d. Representative traces of the relative changes of the ER targeted R-CEPIA1er fluorescence intensities normalized to the baseline (F/F₀ ratio). Ca²⁺ ionophore, ionomycin (10 μ M), stimulation in the presence of 2 mM Ca²⁺ induces ER Ca²⁺-depletion followed by rapid ER filling through SOCE. Compared to control cells, sh1 and sh2 MCU expressing cells possess lower ER filling capacities (Fmax-Fmin difference, black arrow).

e. Histogram depicting the ER filling capacities (Fmax-Fmin difference) of Hs578t cells stably expressing sh Control, sh1 MCU or sh2 MCU shRNAs. (mean \pm S.E.M; n=3 independent experiments). *P < 0.05.



Supplementary Figure S5 related to Figure 5. STIM1 silencing leads to cell migration and cell polarization defects.

a. Immunoblot blot showing the effect of three siRNAs (si1, si2 and si3) on STIM1 expression in Hs578t cells. Scrambled siRNA (si Control) was used as negative control. Vinculin was used as loading control.

b. Histogram representing the distance of cell migration of *stim1*-silenced cells relative to control (mean \pm S.E.M; 30 cells in each experiment; n=3 independent experiments). ***P<0.001.

c. Representative single cell tracking experiments highlighting the cell paths (blue lines) of isolated Hs578t cells (red dots), silenced or not for STIM; acquisition time 24 hours.

d. Confocal images of si control and si1, si2 or si3 STIM1 stained for Vinculin and F-actin using anti-Vinculin antibody and phalloidine rhodamine probe, respectively. Images are presented as negative LUT mask. Scale bar: 20µm.