

Supplemental Figure 1

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3610 GATTGATTTGAAATGCATATTTAAATACAAACAGCAGGGTGTGACTATACGTCCAGAAGGATATCAGGAGAAAAAT
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CYBS14 GATTGATTTGAAATGCATATTTAAATACAAACAGCAGGGTGTGACTATACGTCCAGAAGGATATCAGGAGAAAAAT
CYBS26 GATTGATTTGAAATGCATATTTAAATACAAACAGCAGGGTGTGACTATACGTCCAGAAGGATATCAGGAGAAAAAT
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CYBS9 GAAAGCGCATTCTCTTTCGATTTTTTAAAAAAAGTGATGAGGTAATAAGTCCTGCTGTTTTCCAAAAATAGAAAACAG
CYBS14 GAAAGCGCATTCTCTTTCGATTTTTTAAAAAAAGTGATAAAGGTAATAAGTCCTGCTGTTTTCCAAAAATAGAAAACAG
CYBS26 GAAAGCGCATTCTCTTTCGATTTTTTAAAAAAAGTGATAAAGGTAATAAGTCCTGCTGTTTTCCAAAAATAGAAAACAG
CYBS54 GAAAGCGCATTCTCTTTCGATTTTTTAAAAAAAGTGATAAAGGTAATAAGTCCTGCTGTTTTCCAAAAATAGAAAACAG
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CYBS26 TTTGTAGGTATAAAAATCTCTTTCAAAGAGAAGTTTGGCTTAGTCGATTAGGGAAGATTATGTTACATAATGCCG
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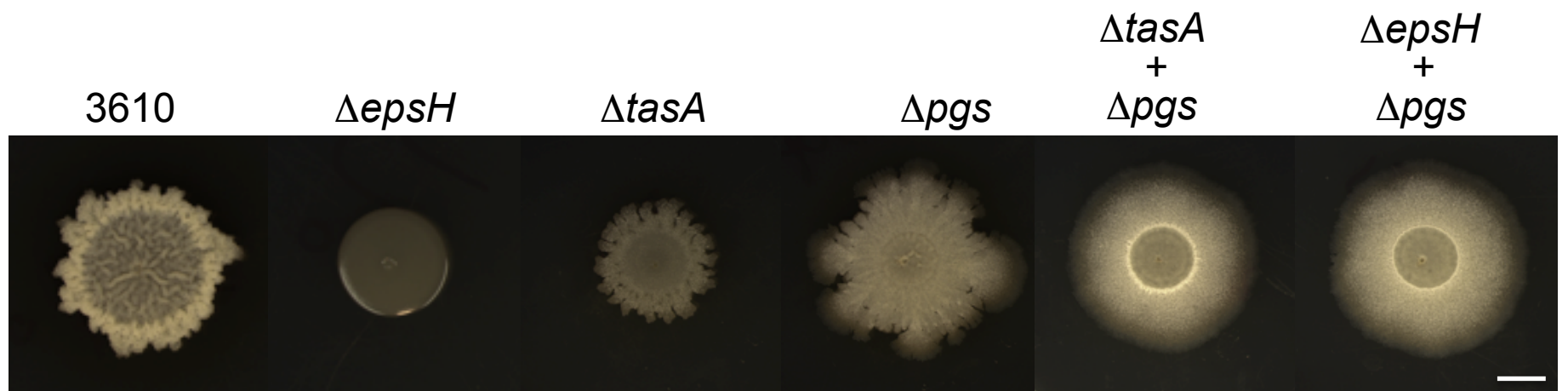
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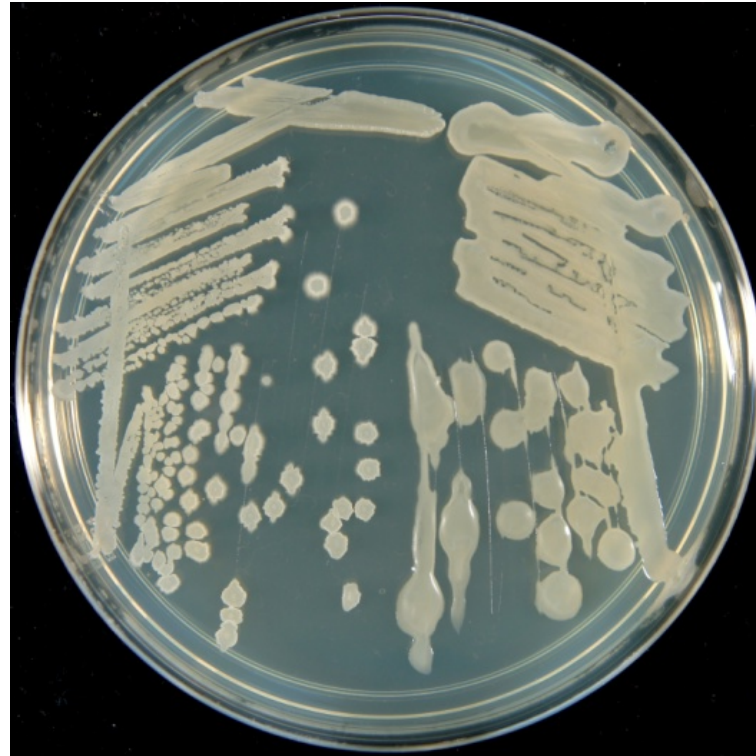
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SM21 ATTGAGAATTCATAGTGATTCTATATACTGATGAATGAATTTACAACAATATAGAAGGAGATGTCGAAAAGCA
CYBS9 ATTGAGAATTCATAGTGATTCTATATACTGATGAATGAATTTACAACAATATAGAAGGAGATGTCGAAAAGCA
CYBS14 ATTGAGAATTCATAGTGATTCTATATACTGATGAATGAATTTACAACAATATAGAAGGAGATGTCGAAAAGCA
CYBS26 ATTGAGAATTCATAGTGATTCTATATACTGATGAATGAATTTACAACAATATAGAAGGAGATGTCGAAAAGCA
CYBS54 ATTGAGAATTCATAGTGATTCTATATACTGATGAATGAATTTACAACAATATAGAAGGAGATGTCGAAAAGCA
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Supplemental Figure 3



3610

$\Delta motA$

Supplemental figure legends

Fig. S1. SDS-PAGE for methylene blue stained samples containing purified γ -PGA from the environmental isolates CYBS54, CYBS26 and their corresponding $\Delta epsH\Delta tasA$ double mutants. Purification of PGA was described in the Methods.

Fig. S2. DNA sequence alignment of the *pgs* promoter regions from various *B. subtilis* environmental strains. The *pgs* promoter sequences were obtained by PCR amplification of the corresponding regions using genomic DNAs from selected *B. subtilis* environmental strains and primers P_{*pgsA*}-F1 and P_{*pgsA*}-R1 (Table S1) and by DNA sequencing. The DNA sequence alignment was performed by using the program ClustalW (<http://www.genome.jp/tools/clustalw/>). The columns with nucleotide polymorphism are highlighted in yellow and green. Yellow represents consensus while green indicates changes. The -10 and -35 motifs of the sigma A-dependent promoter, and the transcription start (+1) were experimentally confirmed and are indicated here (1). A CcpA binding site (*cre* box) was also experimentally characterized in a previous study (2) and indicated here.

Fig. S3. Biofilm complementation by various matrix mutants. Shown here are colony biofilms on LBG agar plates formed by the wild type (3610), the $\Delta epsH$ mutant (RL3852), the $\Delta tasA$ mutant (SB505), the Δpgs mutant (FY6), a 1:1 mixture of the $\Delta tasA$ and Δpgs mutants, and a 1:1 mixture of $\Delta epsH$ and Δpgs mutants. 2- μ l log phase cells were spotted onto the center of the plates and incubated at 37°C for 2 days before images were taken. The scale bar represents 5 mm in length.

Fig. S4. The $\Delta motA$ mutant shows a colony mucoidy phenotype. The wild type strain (3610) and the $\Delta motA$ mutant in 3610 (CY258) were streaked out on the LB agar plate and incubated for 16 hours at 37°C before the picture was taken.

Supplemental references

1. Ohsawa T, Tsukahara K, Ogura M. 2009. *Bacillus subtilis* Response Regulator DegU Is a Direct Activator of *pgsB* Transcription Involved in γ -Poly-glutamic Acid Synthesis. *Bioscience, Biotechnology, and Biochemistry* **73**:2096-2102.
2. Ishii H, Tanaka T, Ogura M. 2013. The *Bacillus subtilis* Response Regulator Gene degU Is Positively Regulated by CcpA and by Catabolite-Repressed Synthesis of ClpC. *Journal of Bacteriology* **195**:193-201.

Table S1. Oligonucleotides used in this study.

Primer name	Primer sequence (5'-3')
pdgS-P1	AAGATACGCTTGGAGAATTTGCGAAGCAAA
pdgS-P2	CAATTCGCCCTATAGTGAGTCGTTTTTATTATCTCCTCCTC
pdgS-P3	CCAGCTTTTGTTCCTTTAGTGAGAAGTATACGGGTGCAATA
pdgS-P4	CGATTTAGGTGTATAATGAACGGGATGGCG
pgs-P1	CGCCGTTAAATCGGTCTTGAGCG
pgs-P2	CAATTCGCCCTATAGTGAGTCGTGTATGACAGCACAGGCTA
pgs-P3	CCAGCTTTTGTTCCTTTAGTGAGAAGTATACGGGTGCAATA
pgs-P4	CGATTTAGGTGTATAATGAACGGGATGGCG
PpgsB-F1	GTACAAGCTTATGACCTTGTCCTTAAGAAACAG
PpgsB-R1	GTACGGATCCCTTCTATATTGTTGTAATTC