

Establishment of a novel tick-Babesia experimental infection model

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Supplementary Methods

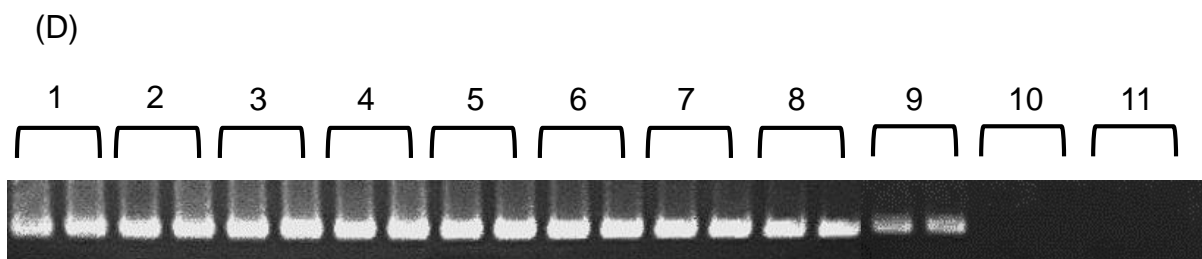
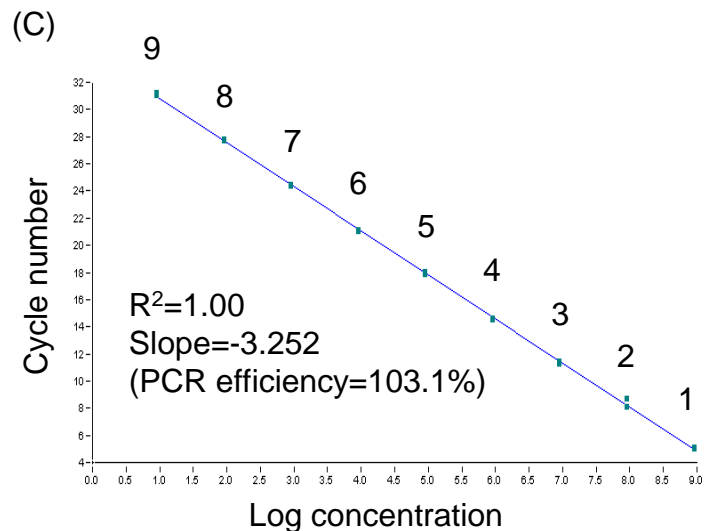
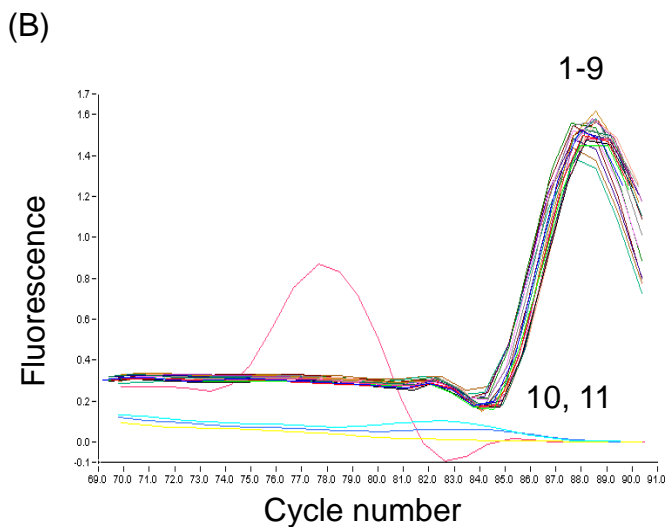
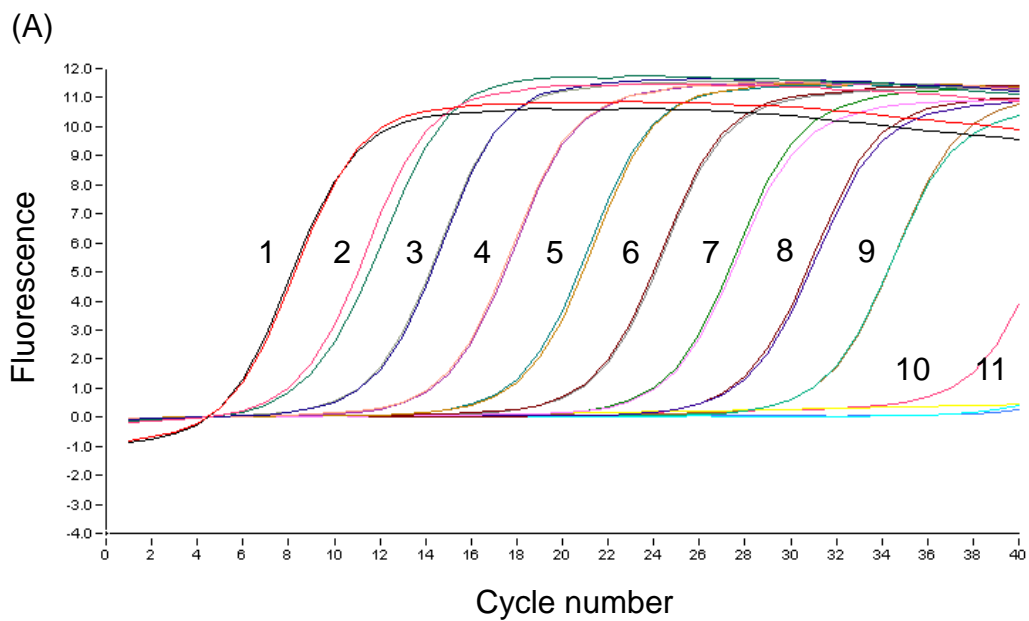
Recombinant protein expression of *B. ovata* P29

The predicted P29 homologous fragments were amplified from *B. ovata* cDNA using the result of RNA sequencing from the midgut of *B. ovata*-infected ticks at 3 DPE (unpublished data). After confirming the sequencing (Supplementary Fig. 2, FASMAC Co., Ltd.), the sequence was cloned into the pRSET B vector (Invitrogen, Carlsbad, CA, USA) using an In-Fusion[®] HD Cloning Kit (TaKaRa) with a specific primer set (Supplementary Table 1). For In-Fusion cloning, the pRSET B vector was digested with *Bam*HI and *Hind*III restriction enzymes (Toyobo) and purified with the MinElute Gel Extraction Kit (Qiagen, Hilden, Germany).

The recombinant plasmid was transformed in Rosetta[™] (DE3) Competent Cells (Novagen, Darmstadt, Germany) and expressed by induction with 1 mM isopropyl- β -D(-)-thiogalactopyranoside (IPTG) at 37°C for 4 h. The expressed recombinant protein was purified using the ProBond Purification System (Invitrogen) from the insoluble fraction under denaturing conditions. The purified recombinant protein was dialyzed against the PBS solution for refolding. The purity of the recombinant proteins was checked by SDS-PAGE. The concentrations of recombinant proteins were determined by a Micro BCA[™] Protein Assay Kit (Thermo Scientific) and stored at -30°C until use.

Production of an anti-serum against *B. ovata* P29

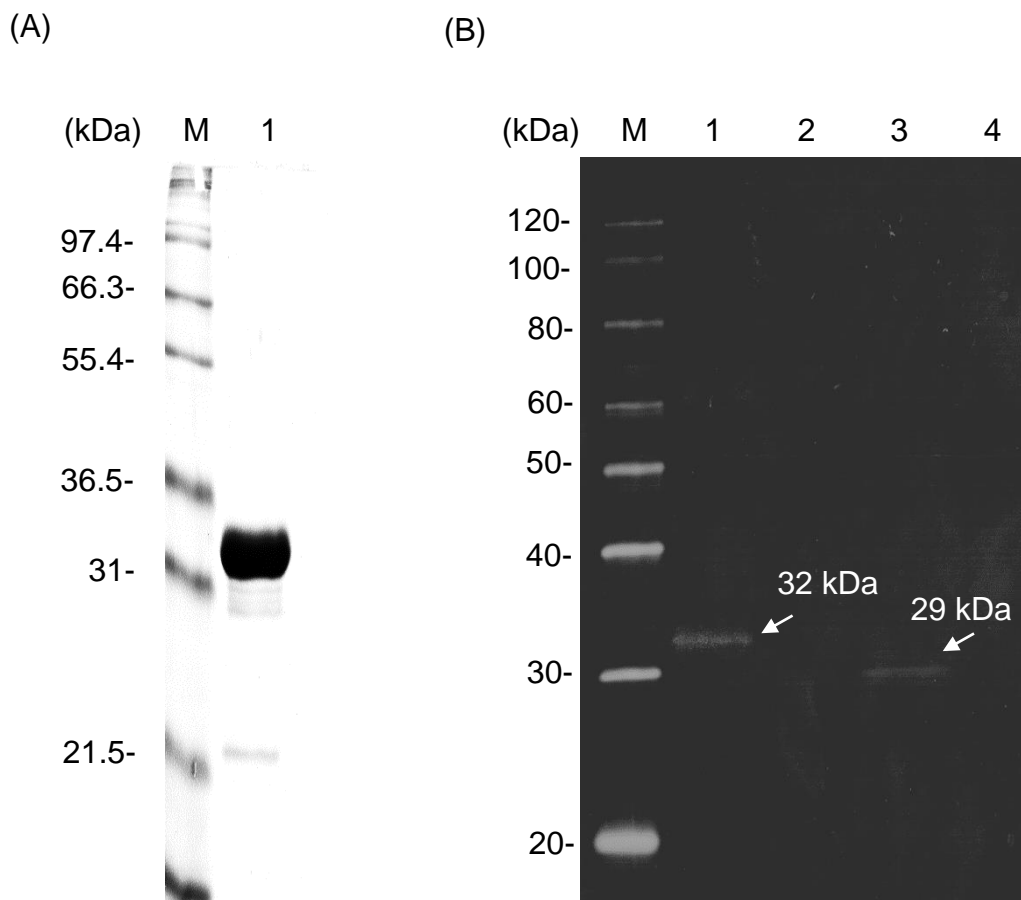
To prepare mouse anti-BoP29 sera, 100 μ g of recombinant BoP29 completely mixed with TiterMax Gold Adjuvant (Sigma-Aldrich) was *subcutaneously* injected into BALB/c mice (Japan SLC). After 2 weeks, these mice were injected again to boost the generation of antibodies. Whole blood was collected from the mice 2 weeks after the second immunization to obtain the specific antisera for BoP29.



Supplementary Fig. 1. Sensitivity of qPCR. To confirm the detection limit of qPCR, a 10-fold serial dilution of Bo β -tub/pTA2 was used for a template. Tick DNA and water were also used for a negative control. (A) Amplification curve (B) Melting curve (C) Standard curve (D) Gel electrophoresis of the PCR products. 1–9, 10-fold serial dilutions of Bo β -tub/pTA2 plasmids mixed with tick genomic DNA; 10, tick DNA; 11, water sample.

1 ATGCAGTGCTGTTCTAGGGACACTCGTCTAATGGACGAGGAGATCGACACCCAAGACGTA
 M Q C C S R D T R L M D E E I D T Q D V 20
 61 CAAGTGAGGACAATCGGCACCGTGGCCGACCGTTCCAGGGCACAAGAAATTGGCCAAAAC
 Q V R T I G T V A D R S R A Q E I G Q N 40
 121 GTCGAGAGGCAATGGGTGCGCCGTTACCACCTACCAGCCCCTTGATACCATTACGAAGACG
 V E R Q W V A V T T Y Q P V D T I T K T 60
 181 GTGGAAATTCCAGTTGTCAAGACCGTTGAACGCGTTGTCCCAAGCCTGTCATCCAGGAA
 V E I P V V K T V E R V V P K P V I Q E 80
 241 CGTGTAATTCAGGTGCCCGCGAAGTTACCCAGGTTGTTGAAAAGGTTGTTGAGATCCCT
 R V I Q V P R E V T Q V V E K V V E I P 100
 301 GATGTAAAGTTCGTCGAGAAGATCATTGAGGTTCCACAGGTCCAATACCGCAACAAACTC
 D V K F V E K I I E V P Q V Q Y R N K L 120
 361 GTACCGAAGGTTGAGGTTGTTGAGAAGATCGTGAAAAGCCGCAAATCATCGAGCAGTGG
 V P K V E V V E K I V E K P Q I I E Q W 140
 421 ACTGAGCGCAAGGTCGAGGTTCCCCAAATTAAGGAGGTTGTGCGCTACAAGGAAATTGAT
 T E R K V E V P Q I K E V V R Y K E I D 160
 481 GAGACAGAGGAGATCATCCGCTACTACCCTAAGGGACATGGCAACATTGACTGGGATAAG
 E T E E I I R Y Y P K G H G N I D W D K 180
 541 GAGTGCGAAAAGGCTCACATTATGATTCCCAGCGAGGTTACGGAATCCAAGGCCGCTAAC
 E C E K A H I M I P S E V T E S K A A N 200
 601 AAGCCCGAGGCAACTGCTTAA
 K P E A T A * 207

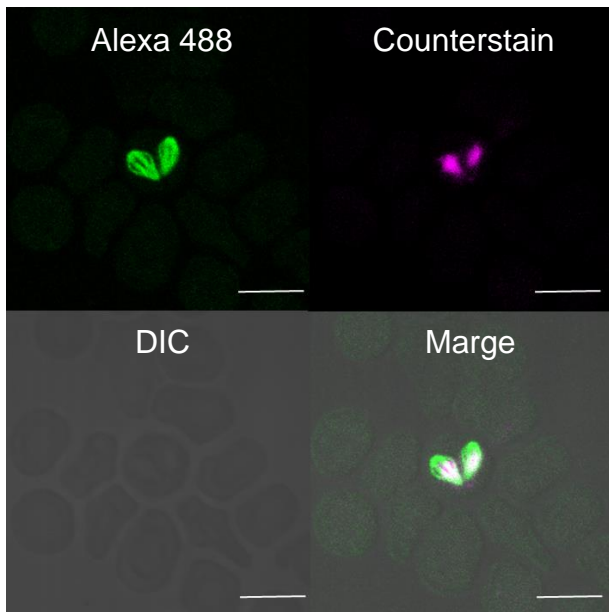
Supplementary Fig. 2. Cloned P29 ORF sequences from *B. ovata*. Cloned BoP29 ORF consists of 621 bp encoding 206 amino acids and shows high homology with *Babesia bigemina* P29 (99%), *Babesia bovis* P29 (93%), and *Babesia gibsoni* P29 (89%).



Supplementary Fig. 3. (A) Purified recombinant BoP29 loaded in 15% SDS-PAGE gel. (B) Sensitivity and specificity of anti-BoP29 mouse serum by western blotting. Lane 1: purified recombinant BoP29; Lane 2: *E. coli* lysate with expression vector; Lane 3: lysate of *B. ovata*-infected RBCs; Lane 4: lysate of normal RBCs.

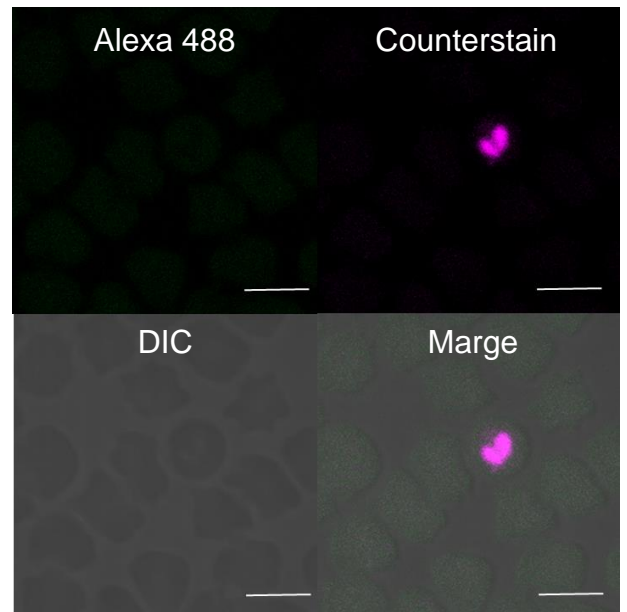
(A)

α -BoP29 mouse serum



(B)

normal mouse serum



Supplementary Fig. 4. IFAT of *B. ovata*-infected RBCs (A) anti-BoP29 mouse serum (B) normal mouse serum. Bar: 5 μ m

Supplementary Table 1 Gene-specific primers used in this paper

Primers	Sequence (5'→3')
Bob-tub F ^{a)}	ACACTGTGCATCCTCACCGTCATAT
Bob-tub R ^{a)}	CTCGCGGATCTTGCTGATCAGCAGA
HIITS2 F ^{b)}	GCGTGTTGGGAAGTCTGAA
HIITS2 R ^{b)}	CGCGGTTTACGAGAGAAAG
BoP29 in-fusion F	ATGACGATAAGGATCCGATGCAGTGCTGTTCTAGGG
BoP29 in-fusion R	GCCAAGCTTCGAATTTAAGCAGTTGCCTCGGG

^{a)}Sivakumar et al, 2014; ^{b)}Hatta et al, 2012

Supplementary Table 2 Parasite burden of *B. ovata* in tick organs

DPE	Experiment 1					Experiment 2				
	Tick ID	BW (mg)	MG	OV	CA	Tick ID	BW (mg)	MG	OV	CA
0	#1	123.3	+	ND	ND	#1	105.1	+	ND	+
	#2	120.9	+	ND	ND	#2	92.8	+	+	ND
	#3	214.7	+	ND	ND	#3	235.9	++	ND	+
						#4	181.4	+	ND	+
1	#4	220.9	ND	ND	++	#5	147.1	ND	ND	+
	#5	241.4	ND	+	+	#6	151.4	ND	++	++
	#6	159.6	ND	++	+	#7	179.9	ND	+++	+
	#7	150.2	++	++	++	#8	202.8	+	++	+
2	#8	188.5	ND	ND	+	#9	154.1	ND	+	+
	#9	277.4	++	ND	+	#10	186.1	ND	ND	+
	#10	145.9	ND	+	+	#11	211.3	ND	++	+
	#11	183.6	ND	++	ND	#12	193.5	+	++	+
3	#12	165.2	ND	+	ND	#13	230.2	ND	ND	ND
	#13	139.4	++	+	++	#14	199.1	ND	++	ND
	#14	173.4	ND	+	+	#15	191.6	ND	++	++
	#15	184.6	ND	++	ND	#16	267.5	+	++	+
4	#16	265.9	ND	++	++	#17	274.3	ND	+	++
	#17	166.6	ND	+	+	#18	229.3	ND	+	++
	#18	160.9	ND	ND	+	#19	220.5	+	+	++
	#19	127.9	++	++	++	#20	253.1	ND	ND	+

The quantified copy number of the *B. ovata* β -*tubulin* gene was normalized by the tick *ITS2* gene. The quantified value was presented as +, 10^{-9} ~ 10^{-6} ; ++, 10^{-6} ~ 10^{-3} ; +++, 10^{-3} ~ 10^0 . ND, not detected