Supplementary Data



Supplementary Figure 1 Effect of *Tsc1* loss in cones of wild-type mice. Data shown are from wild-type mice harboring the *Tsc1^{c/c}* allele. (**a**) Scotopic ERG recordings at 12 months of age with flashes of increasing intensities showing no significant difference between *Cre⁻* and *Cre⁺* mice. Data are representative of recordings from at least 3 mice per genotype. (**b**) Quantification of cone number based on cone arrestin staining at the indicated time points in Sector 2 (see Figure **1e** for schematic of the two sectors; *P < 0.05 by Student's *t* test). (**c**) Western blot analysis of indicated proteins in *Cre⁻* and *Cre⁺* mice. (**d**) Immunofluorescence analysis on retinal cryosections from *Tsc1^{c/c}* mice at 2 months of age for indicated proteins (red signal; green: PNA; blue: nuclear DAPI). In each panel, 50% of the PNA and DAPI signal is removed to better appreciate the expression of the labeled protein in the different retinal layers (ONL: outer nuclear layer, INL: inner nuclear layer, GCL: ganglion cell layer; arrows point to expression in INL and GCL; Scale bar: 20µm).



Supplementary Figure 2 Increase in expression of autophagy genes and nuclear FOXO3A is dependent on mTORC1. (a) Immunofluorescence analyses on retinal flat mounts for indicated proteins (red signal) in rd1- $Tsc1^{c/c}Raptor^{c/c}$ mice at 2 months of age showing no difference in expression between Cre^{-} and Cre^{+} mice. (b) Same genotype as in (a) showing the dependence of nuclear FOXO3A expression (red signal) on mTORC1 activity (green is PNA in (a) and SW OPSIN in (b); blue: nuclear DAPI; Scale bar: 20µm).



Supplementary Figure 3 Effect of rapamycin on rd1- $Tsc1^{c/c}$ mice. (a) Immunofluorescence analyses on retinal flat mounts for p62 and UBIQUITIN (red signal) in rd1- $Tsc1^{c/c}Cre^+$ mice at 2 months of age upon vehicle or rapamycin administration (Scale bar: 20µm). (b) Quantification of cone survival in rd1- $Tsc1^{c/c}Cre^+$ mice at 2 months of age when administered with vehicle or rapamycin once every five days from P28 to 2 months. (c, d) Kinetics of phospho-S6 recovery after one injection of rapamycin in rd1- $Tsc1^{c/c}Cre^+$ mice. (c) Recovery seen in cones (red signal) by immunofluorescence on retinal flat mounts (green: PNA; Scale bar: 20µm). (d) Western blot for phospho-S6 with retinal protein extracts of mice harvested 1, 3 or 5 days after rapamycin injection. By western blot pS6 is almost fully recovered by day 5. (e) Quantification of cone survival at 4 months of age in rd1- $Tsc1^{c/c}$ Cre^- mice treated with varying number of rapamycin injections between 1-4 months of age (Cre^- control of Figure **5c-d**).



Supplementary Figure 4 Effect of rapamycin administration in rd1- $Tsc1^{c/c}Cre^+$ mice. (a) Immunofluorescence analyses for p62 (red signal) at 4 months of age in rd1- $Tsc1^{c/c}Cre^+$ mice treated with vehicle, 6 and 2 injections of rapamycin (green: PNA). (b) Dynamic of mTOR (red signal) and LAMP2 (green signal) colocalization post rapamycin injection in rd1- $Tsc1^{c/c}Cre^+$ mice with upper row showing mTOR staining, middle row showing LAMP2 and lower row showing overlay of the two.



Supplementary Figure 5 Loss of *Pten* in cones of wild-type mice does not affect cone function and expression of cone-arrestin. (**a**) Immunofluorescence analyses (red signal) on retinal flat mounts for phosphorylation sites of indicated proteins in rd1-*Pten*^{c/c} mice. SW OPSIN (green signal) was used to label cones (blue: nuclear DAPI; Scale bar: 20µm). (**b-d**) Analyses in *Pten*^{c/c} mice. (**b**) Immuno-fluorescence on retinal flat mounts for p62 (red signal) in cones at 2 months of age (arrow: p62 aggregate in cone segment of Cre^+ mouse; green: PNA; Scale bar: 20µm). Bar graph indicating percentage of cones with p62 aggregates in retinae from *Pten*^{c/c} *Cre*⁺ and *Tsc1*^{c/c}*Cre*⁺ mice at 2 months of age. p62 aggregates were not detected in *Cre*⁻ mice for either genotype and thus not included in the graph. Data are representative of measurements in at least 100 cone segments across 2 animals per genotype (**P < 0.01 by Student's *t* test). (**c**) Evaluation of cone function by photopic ERG recordings from at least 6 mice per group showing b-wave amplitudes over time. No significant difference was detected over time between *Cre*⁻ and *Cre*⁺ littermates. (**d**) Quantification of cone number based on cone arrestin staining in two retinal sectors as described in Figure (**1e**) of at least 2 animals per group. No difference was observed up to 1 year of age between *Cre*⁻ and *Cre*⁺ littermates.