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Supplemental Information

Endoplasmic Reticulum Proteostasis Influences

the Oligomeric State of an Amyloidogenic

Protein Secreted from Mammalian Cells

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SUPPLEMENTAL FIGURES 1



Mass Diff (vs. ^{FT} TTR ^{WT})	Modification	Predicted Site of Modification	Relative Abundance (%)
+199	+SO $_3$ and +Cys	Tyr and Cys10	38 +/- 4
+80	+SO ₃	Tyr	31 +/- 5
+119	Cys	Cys10	12 +/- 2
+160	2 x SO ₃	Tyr	8 +/- 2
Parent			6 +/- 3

Figure S1 (Supplement to Figure 1).

- **A.** Plot showing the compound **1**-TTR conjugate fluorescence of recombinant TTR^{WT} (200 nM) in cell 5 culture media separated by anion exchange chromatography. Compound 1 (10 µM) was added to unconditioned media containing recombinant TTR^{WT} for 16 h prior to analysis by anion exchange 7 8 chromatography.
- **B.** Plot showing the compound **1**-TTR conjugate fluorescence of recombinant ^{FT}TTR^{WT} (200 nM) in cell 9 culture media separated by anion exchange chromatography. Compound 1 (10 µM) was added to 0 unconditioned media containing recombinant ^{FT}TTR^{WT} for 16 h prior to analysis by anion exchange 1 2 chromatography.
- **C.** Plot showing the compound **1**-TTR conjugate fluorescence of ^{FT}TTR^{WT} in conditioned media. 3 Compound **1** at the indicated concentration was added to conditioned media containing ^{FT}TTR^{WT} for 16 4 5 h prior to analysis by anion exchange chromatography.
- **D.** Table showing modifications of ^{FT}TTR^{WT} identified by mass spectrometric analysis of TTR 6 immunopurified from media conditioned for 16 h on HEK293 cells expressing ^{FT}TTR^{WT}. The mass 7 difference from parent, modification, predicted modification site and relative abundance are shown. 8 These modified TTRs integrate into heterotetramers to comprise the 8 distinct peaks identified by 9 compound 1-TTR conjugate fluorescence in UPLC anion exchange chromatograms, as shown in Fig. 0
 - 1D.



3 Figure S2 (Supplement to Figure 2).

- A. Immunoblot of fractions collected from media conditioned on HEK293 cells expressing ^{FT}TTR^{WT} (top) or
 ^{FT}TTR^{A25T} (bottom) separated by gel filtration chromatography. The migration of molecular weight
 standards are shown above. The arrows show ^{FT}TTR. The asterix (*) identifies non-TTR background
 bands that cross-react with this antibody in conditioned media. The migration of ^{FT}TTR^{WT} is identical to
 that observed for recombinant ^{FT}TTR^{WT} (Chen et al., 2014).
- B. Representative SDS-PAGE immunoblot and quantification of total TTR levels in media conditioned for
 16 h on HEK293 cells expressing ^{FT}TTR^{WT} (black), ^{FT}TTR^{V122I} (orange), ^{FT}TTR^{V30M} (green), ^{FT}TTR^{L55P}
 (blue), or ^{FT}TTR^{A25T} (red). The graph shows relative amounts of the different TTR variants normalized to
 ^{FT}TTR^{WT}. Error bars show SEM for n=3.
- C. Immunoblot of ^{FT}TTR^{A25T} incubated in conditioned media at 37 °C for the indicated time. The pH of the
 media was adjusted as indicated to induce protein unfolding. Note that the ^{FT}TTR^{A25T} signal is not lost
 during this incubation at any pH, indicating that no proteolysis is observed.
- 6



Figure S3 (Supplement to Figure 4)



B. Proton NMR for Taf-SO $_3^-$.



Figure S4 (Supplement to Figure 5).

9	Α.	Representative plot showing the compound 1-TTR conjugate fluorescence of media conditioned for 16
0		h on HEK293 cells expressing $^{\text{FT}}\text{TTR}^{\text{L55P}}$ in the presence of vehicle (black), Taf (10 $\mu\text{M};$ red), or Taf-
1		SO ₃ ⁻ (10 μ M; blue). Compound 1 (10 μ M) was added to conditioned media for 16 h prior to analysis by
2		anion exchange chromatography.

B. Representative plot showing the compound 1-TTR conjugate fluorescence of media conditioned for 16
 h on HEK293 cells expressing ^{FT}TTR^{A25T} in the presence of vehicle (black), Taf (10 μM; red), or Taf SO₃⁻ (10 μM; blue). Media was conditioned for 16 h prior to collection. Compound 1 (10 μM) was added
 to conditioned media for 16 h prior to analysis by anion exchange chromatography.

- C. Plot showing relative tetramer levels (measured by compound 1-TTR conjugate fluorescence UPLC assay) of media conditioned on HEK293 cells expressing ^{FT}TTR^{A25T} in the presence of Taf-SO₃⁻ (10
- μ M). The x-axis shows the time media was incubated prior to addition of compound **1** (14.4 μ M).



2 Figure S5 (Supplement to Figure 6)

- A. Immunoblot of lysates prepared from HEK293 cells expressing ^{FT}TTR^{A25T} treated with vehicle or Tg (0.5
 μM). The Tg-dependent increase in BiP is shown in the KDEL immunoblot panel.
- B. Representative plot showing the compound 1-TTR conjugate fluorescence of media conditioned for 16
 h on HEK293 cells expressing ^{FT}TTR^{A25T} conditioned in the presence of vehicle (black) or Tg (0.5 μM;
 red). Media was conditioned the presence of Taf-SO₃⁻ (10 μM). Compound 1 (10 μM) was added to
 conditioned media for 16 h prior to analysis by anion exchange chromatography.
- C. Representative plot showing the compound 1-TTR conjugate fluorescence of media conditioned for 16
 h on HEK293 cells expressing ^{FT}TTR^{A25T} conditioned in the presence of vehicle (black) or increasing Tg
 concentration, as indicated. Media was conditioned the presence of Taf-SO₃⁻ (10 μM). Compound 1 (10
 μM) was added to conditioned media for 16 h prior to analysis by anion exchange chromatography.
- D. Representative CN-PAGE and SDS-PAGE immunoblot of media conditioned for 16 h on HEK293 cells
 expressing ^{FT}TTR^{A25T} conditioned in the presence of vehicle (black) or increasing Tg concentration, as
 indicated. Media was conditioned the presence of Taf-SO₃⁻ (10 μM).
- 6 E. Graph showing normalized TTR tetramers in the conditioned media described in Fig. S5C,D.
 7 Normalized tetramers were calculated by dividing the relative tetramer levels by the total amount of
 8 TTR in each sample measured by SDS-PAGE. This normalization is necessary to account for the
 9 reduced total ^{FT}TTR^{A25T} levels observed in Tg-treated cells. Error bars show SEM for n=3. *p<0.05,
 0 **p<0.01, ***p<0.005.
- F. Graph showing normalized TTR tetramers in the conditioned media prepared on HEK293 cells
 transfected with 5 or 2.5 μg DNA and treated with vehicle (grey) or Tg (0.5 μM; red), as indicated.
 Normalized tetramers were calculated by dividing the relative tetramer levels by the total amount of
 TTR in each sample measured by SDS-PAGE. This normalization is necessary to account for the
 reduced total ^{FT}TTR^{A25T} levels observed in Tg-treated cells. Error bars show SEM for n=2.
 G. Graph showing total (open bars; measured by SDS-PAGE) or tetrameric (closed bars; measured by
- integrating compound 1-TTR conjugate fluorescence from UPLC) TTR in media conditioned for 16 h on
 HEK293 cells expressing ^{FT}TTR^{WT} in the presence of vehicle (grey) or Tg (0.5 µM; red). Media was

9		conditioned in the presence of Taf-SO $_3^-$ (10 μ M). All data are shown relative to vehicle-treated controls.
0		Error bars show SEM for n=6.
1	H.	Graph showing the posttranslational modifications of FTTTR ^{WT} immunopurified from media conditioned
2		on HEK293T cells expressing $^{FT}TTR^{WT}$ and treated in the presence of vehicle (grey) or Tg (0.5 μM ;
3		red). Modifications of ^{FT} TTR ^{WT} were determined using the same LC-MS approach described in Fig. 1E .
4		

5 SUPPLEMENTAL EXPERIMENTAL PROCEDURES

6 Antibodies

7 The antibodies used for immunoblotting were monoclonal mouse M2 anti-FLAG (Sigma), monoclonal mouse
8 anti-tubulin (Sigma) and polyclonal rabbit anti-TTR antibody (Dako).

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0 Cell Culture, Lysates, Plasmids, and Transfections

HEK293 cells used in this study were purchased from the ATCC and cultured in Dulbecco's modified Eagle's 1 2 medium (DMEM) supplemented with glutamine, penicillin/streptomycin, and 10% fetal bovine serum, as previously described (Shoulders et al., 2013). All TTR point mutations were incorporated into the previously 3 described FLAG₂.TTR.pcDNA3.1 vector through site-directed mutagenesis. Transient transfections of ^{FT}TTR 4 5 variants were performed by calcium phosphate transfection using established procedures (Shoulders et al., 2013). Whole cell lysates were prepared in Lysis Buffer (20 mM HEPES [pH 7.4], 100 mM NaCl, 1 mM EDTA, 6 7 1% Triton X-100 supplemented with EDTA-free protease inhibitors [Roche]). Protein concentration in lysates 8 was normalized using the Bio-Rad protein assay.

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0 Characterization of TTR modifications

^{FT}TTR^{WT} was immunopurified from cell-conditioned media by overnight incubation with protein A resin 1 covalently conjugated to anti-TTR antibody at 4 ^oC, as previously described (Sekijima et al., 2003). The beads 2 were washed four times with 10 mM Tris pH 8.0 100 mM NaCl 0.05% saponin and two times with 10 mM Tris 3 pH 8.0 100 mM NaCl. FTTTR^{WT} was then eluted overnight using 100 mM triethylamine pH 11.5 with gentle 4 shaking. For LC-MS analysis, eluates were separated using a 5 µm ID 300 Å pore size C8 RP-HPLC column 5 6 (Agilent) over a 15%/min linear acetonitrile gradient and analyzed on an 1100 MSD SL mass spectrometer 7 (Agilent). Deconvolution parameters were set to 10 kDa \leq MW \leq 20 kDa, q \leq 50, and minimum 6 peaks for identification. 8

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0 Recombinant TTR Aggregation Assay

1 Recombinant TTR^{WT} (7.2 μ M tetramer) in 10 mM Tris pH 7.5 100 mM NaCl 1 mM EDTA was incubated for 1 h 2 with the indicated equivalent of Taf-SO₃⁻ or Taf (1 equivalent = 7.2 μ M molecule). The protein was then diluted

- 3 1:1 into 50 mM Acetate pH 4.4 100 mM NaCl 1 mM EDTA resulting in a final tetramer concentration of 3.6 μM.
- 4 The sample was then incubated for 72 h at room temperature. Aggregation was measured by turbidity at 330
- 5 and 400 nm, as previously described (Razavi et al., 2003; Sekijima et al., 2005).
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