

## Supporting Information

### Targeting the Src homology 2 (SH2) domain of signal transducer and activator of transcription 6 (STAT6) with cell-permeable, phosphatase-stable phosphopeptide mimics potently inhibits activation and transcriptional activity.

Pijus K. Mandal,<sup>1</sup> Pietro Morlacchi,<sup>1</sup> J. Morgan Knight,<sup>1</sup> Todd M. Link,<sup>2</sup> Gilbert R. Lee, IV,<sup>2</sup> Roza Nurieva,<sup>3</sup> Divyendu Singh,<sup>3</sup> Ankur Dhanik,<sup>4</sup> Lydia Kavraki<sup>4</sup>, David B. Corry,<sup>5</sup> John E. Ladbury,<sup>2</sup> John S. McMurray<sup>1\*</sup>

<sup>1</sup>Department of Experimental Therapeutics, <sup>2</sup>Department of Biochemistry and Molecular Biology, Center for Biomolecular Structure and Function, <sup>3</sup>Department of Immunology, The University of Texas M. D. Anderson Cancer Center, Houston, Texas, <sup>4</sup>The Department of Computer Science, Rice University, Houston, Texas, <sup>5</sup>Departments of Medicine and Pathology & Immunology, The Baylor College of Medicine, Houston, Texas

Table S1. Characterization of phosphopeptides assayed for binding to STAT6.....	S2
Table S2. Characterization of phosphate analogs of <b>17</b> .....	S3
Table S3. Yields and characterization of prodrug analogs of <b>17</b> .....	S4
Figure S1. Inhibition of pSTAT6 and pAkt in MDA-MB-468 breast cancer cells.....	S5
Figure S2. Inhibition of STAT proteins in CD4 <sup>+</sup> T lymphocytes.....	S6
NMR Characterization of <b>19c</b> .....	S7
NMR Characterization of <b>31</b> .....	S8

**Table S1.** Characterization of phosphopeptides assayed for binding to STAT6.

#	Phosphopeptide	(M+H) Calcd	(M+H) Found	HPLC RT (min) <sup>a</sup>	Yield (mg)
1	Ac-pTyr-Lys-Pro-Phe-Gln-Asp-Leu-Ile-NH <sub>2</sub>	1144.5444	1144.5484	25.4	91
2	Ac-pTyr-Lys-Pro-Phe-Gln-Asp-Leu-NH <sub>2</sub>	1031.4603	1031.4668	21.4	48
3	Ac-pTyr-Lys-Pro-Phe-Gln-Asp-NH <sub>2</sub>	918.3763	918.3787	16.4	75
4	Ac-pTyr-Lys-Pro-Phe-Gln-NH <sub>2</sub>	803.3493	803.3534	16.1	50
5	Ac-pTyr-Lys-Pro-Phe-NH <sub>2</sub>	675.2907	675.2950	17.17	63
6	Ac-pTyr-Lys-Pro-NH <sub>2</sub>	528.2223	528.2234	9.33	28
7	Ac-pTy-Tle-Pro-Phe-NH <sub>2</sub>	660.2798	660.2880	21.73	46
8	Ac-pTy-Nle-Pro-Phe-NH <sub>2</sub>	660.2798	660.2863	24.07	31
9	Ac-pTy-Ala-Pro-Phe-NH <sub>2</sub>	618.2329	618.2415	18.40	39
10	PhCH <sub>2</sub> CH <sub>2</sub> CO-pTyr-Lys-Pro-Phe-NH <sub>2</sub>	765.3377	765.3385	24.16	63
11	PhCH <sub>2</sub> CO-pTyr-Lys-Pro-Phe-NH <sub>2</sub>	751.3220	751.3241	22.52	58
12	PhCO-pTyr-Lys-Pro-Phe-NH <sub>2</sub>	737.3064	737.3081	21.66	71
13	PhCH=CHCO-pTyr-Lys-Pro-Phe-NH <sub>2</sub>	763.3220	763.3239	24.92	42
14	pCinn-Lys-Pro-Phe-NH <sub>2</sub>	616.2536	616.2521	17.9	49
15	p-Indole-Lys-Pro-Phe-NH <sub>2</sub>	629.2489	629.2490	18.17	32
16	p-Benzofuran-Lys-Pro-Phe-NH <sub>2</sub>	630.2329	630.2324	18.04	71

<sup>a</sup> Peptides were tested for purity by reverse phase HPLC on a Waters Alliance HPLC using a Phenomenex Luna C18 (2) 5 $\mu$ M 2.1 x 250 mm column. A gradient of 0-40% MeCN/30 min at a flow rate of 0.4 mL/min was used with both mobile phases containing 0.1% TFA. Elution was monitored at 230 nm, 254 nm, and 275 nm.

**Table S2.** Characterization of phosphate analogs of **17**.

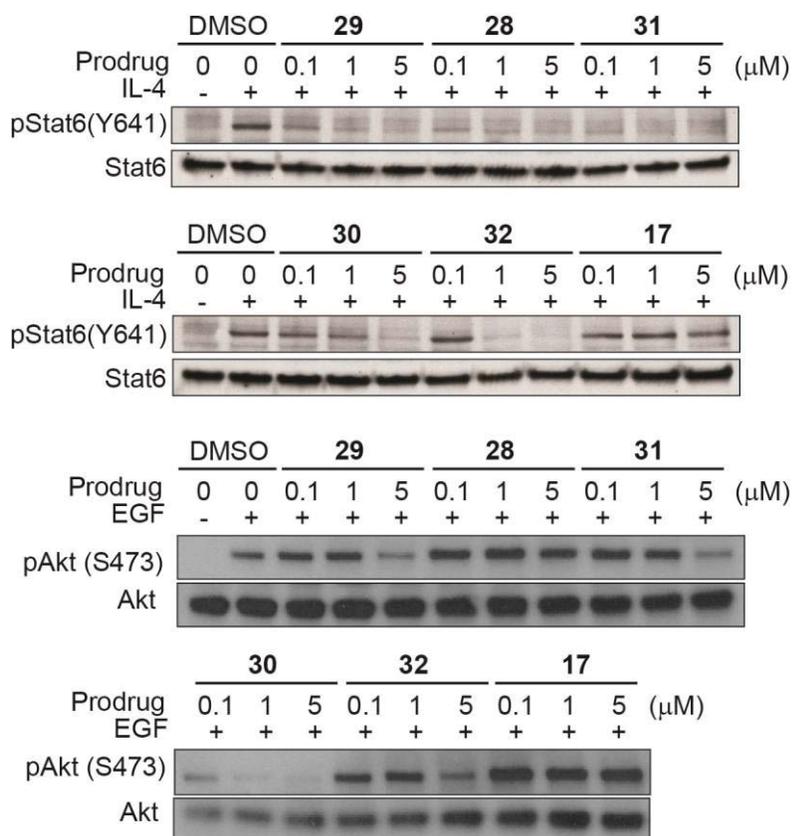
	Sequence	Yield mg	% Yield	HRMS (M+H) Calcd	HRMS (M+H) Found	HPLC RT (min) <sup>a</sup>
<b>18a</b>	pCinn-Tle-Pro-N(4-I-Ph)(Ph)	35	47	732.1336	732.1333	22.5
<b>19a</b>	βMpCinn-Tle-Pro-N(4-I-Ph)(Ph)	39	53	746.1492	746.1458	22.8
<b>20</b>	pInd-Tle-Pro-N(4-I-Ph)(Ph)	61	72	745.1288	745.1273	22.13
<b>18b</b>	pCinn-Tle-Pro-NPh <sub>2</sub>	23	39	606.2369	606.2350	18.4
<b>18c</b>	pCinn-Tle-Pro-N(Me)(Ph)	41	27	544.22	544.20	26.7
<b>18d</b>	pCinn-Tle-Pro-NHPh	34	22	530.21	530.25	30.3
<b>19b</b>	βMpCinn-Tle-Pro-NPh <sub>2</sub>	31	51	620.2526	620.2510	19.46
<b>19c</b>	βMpCinn-Tle-Pro-N(Me)(Ph)	29	56	558.2369	558.2373	15.13
<b>19d</b>	βMpCinn-Tle-Pro-NHPh	25	48	544.2213	544.2181	17.2
<b>19e</b>	βMpCinn-Tle-Pro-NHCH <sub>2</sub> Ph	27	51	558.2369	558.2336	16.01
<b>19f</b>	βMpCinn-Tle-Pro-NHCH <sub>2</sub> CH <sub>2</sub> Ph	31	53	572.2526	572.2526	16.57
<b>19g</b>	βMpCinn-Tle-Pro-N(Me)(C <sub>6</sub> H <sub>11</sub> )	37	67	564.2839	564.2842	17.5
<b>19h</b>	βMpCinn-Tle-Pro-N(CH <sub>3</sub> ) <sub>2</sub>	30	61	496.2213	496.2191	10.7
<b>19i</b>	βMpCinn-Tle-Pro-N(C <sub>2</sub> H <sub>5</sub> ) <sub>2</sub>	27	52	524.2526	524.2502	12.00
<b>21</b>	βMpCinn-Tle-Sar-N(Me)(Ph)	22	42	532.2213	532.2186	15.92
<b>22</b>	βMpCinn-Tle-Ala-N(Me)(Ph)	31	59	532.2213	532.2184	15.15
<b>23</b>	βMpCinn-Tle-N(Me)Ala-N(Me)(Ph)	30	55	546.2369	546.2189	24.4
<b>24</b>	βMpCinn-Tle-pyrrolidine-2-CH=CHPh	22	40	527.2311	527.2312	20.47
<b>25</b>	βMpCinn-Tle-pyrrolidine-2-CH=CHCH <sub>2</sub> CH <sub>2</sub> Ph	19	35	555.2624	555.2867	24.52
<b>26</b>	βMpCinn-Tle-pyrrolidine-2-CH <sub>2</sub> CH <sub>2</sub> CH <sub>2</sub> CH <sub>2</sub> Ph	16	30	557.2780	557.2827	25.52
<b>27</b>	βMpCinn-Tle-pyrrolidine-2-CH <sub>2</sub> N(Me)Ph	44	31	544.25	544.35	28.14

<sup>a</sup> Peptides were tested for purity by reverse phase HPLC on a Waters Alliance HPLC using a Phenomenex Luna C18 (2) 5μM 2.1 x 250 mm column. A gradient of 0-40% MeCN/30 min at a flow rate of 0.4 mL/min was used with both mobile phases containing 0.1% TFA. Elution was monitored at 230 nm, 254 nm, and 275 nm.

**Table S3.** Yields and characterization of prodrug analogs of **1**.

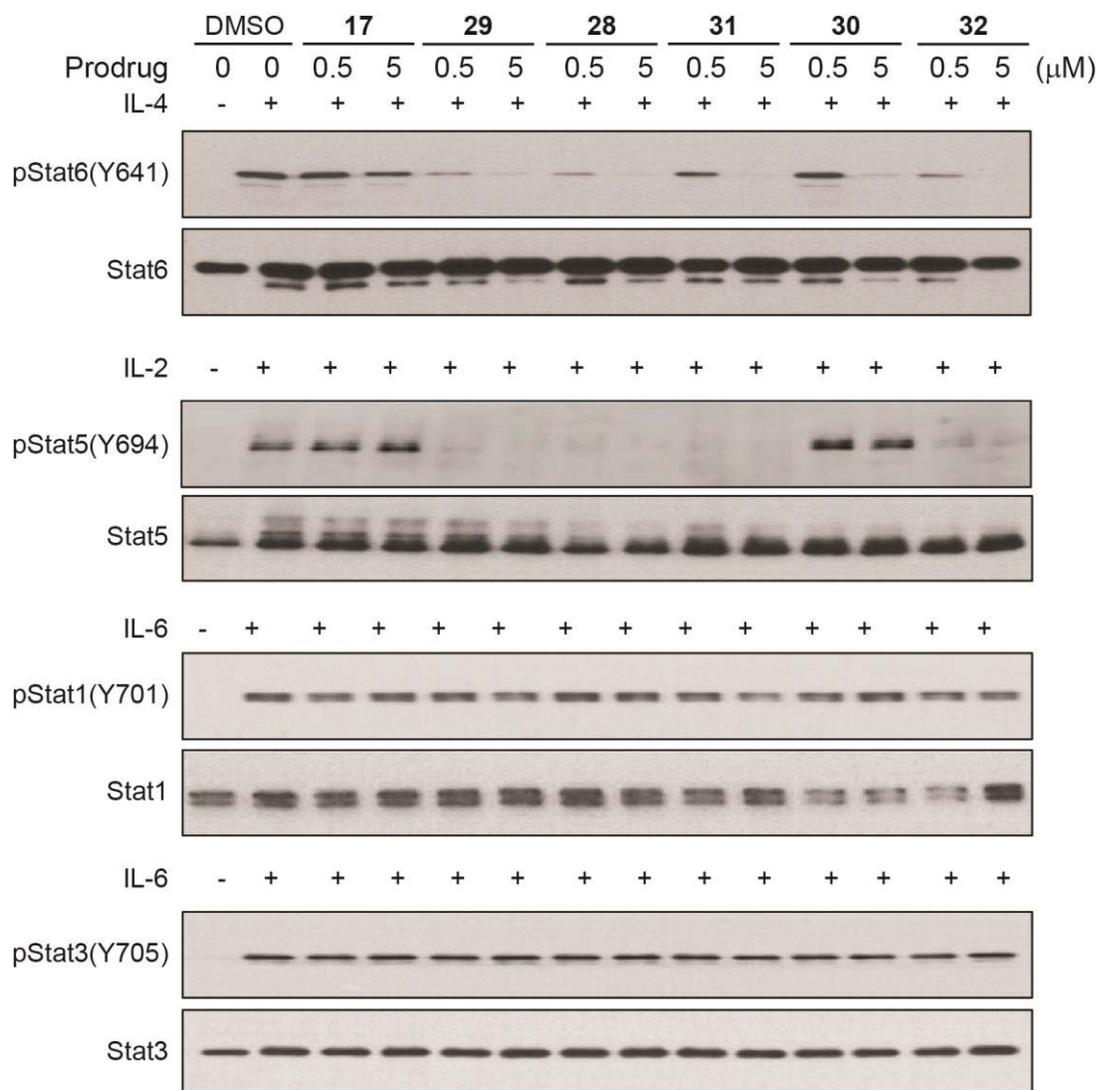
#	Structure	Yield (mg)	% Yield	(M+H) Calcd	(M+H) Found	HPLC RT (min) <sup>a</sup>
<b>17</b>	F2PmCinn(POM <sub>2</sub> )-Tle-Pro-N(4-I-Ph)(Ph)	64	66	994.2716	994.2753	35.03
<b>28</b>	F2PmCinn(POM <sub>2</sub> )-Tle-Pro-NPh <sub>2</sub>	63	58	868.3750	868.3753	35.6
<b>29</b>	F2PmCinn(POM <sub>2</sub> )-Tle-Pro-N(Me)(Ph)	56	53	806.3593	806.3578	31.2
<b>30</b>	F2PmCinn(POM <sub>2</sub> )-Tle-Pro-NHPH	49	38	792.3437	792.3463	33.2
<b>31</b>	βMF2PmCinn(POM <sub>2</sub> )-Tle-Pro-N(Me)Ph	56	53	820.3750	820.3746	34.73
<b>32</b>	βMF2PmCinn(POM <sub>2</sub> )-Tle-Pro-N(Me)(C <sub>6</sub> H <sub>5</sub> )	48	45	826.4219	826.3944	33.77

<sup>a</sup> Prodrugs were tested for purity by reverse phase HPLC on a Waters Alliance HPLC using a Phenomenex Luna C18 (2) 5μM 2.1 x 250 mm column. A gradient of 10-80% MeCN/30 min at a flow rate of 0.4 mL/min was used. Elution was monitored at 230 nm, 254 nm, and 275 nm.



**Figure S1.** Inhibition of STAT6 and Akt phosphorylation in MDA-MB-468 breast cancer cells.

Cells were serum starved overnight and were treated with prodrugs for 2 h. IL-4 or EGF were added and total and phosphoprotein levels were determined by western blot after 1 hr.



**Figure S2.** Inhibition of STAT proteins in CD4<sup>+</sup> T lymphocytes from C57BL/6J mice. CD4<sup>+</sup> T cells isolated from C57BL/6J mice were pre-treated with STAT6 inhibitors for 2 hours and activated with plate-bound anti-CD3/anti-CD28 in the presence or absence of indicated cytokines for 15 minutes. The whole cell lysate was subjected to western blot assay to detect the levels of phosphorylated and total STAT6, STAT5, STAT1 and STAT3.

NMR of Compound **19c**

<sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz)

*Trans* :*cis* based on integration of the Tle CH<sub>α</sub>, Pro CH<sub>α</sub>, and Cinn CH<sub>α</sub> resonances

*Trans* isomer, 83%

	αNH	αCH	βCH <sub>2</sub>	γCH <sub>2</sub>	δCH <sub>2</sub>	Arom	NCH <sub>3</sub>	NPh
Cinn		6.50	2.42 (CH <sub>3</sub> )			7.17, 7.49		
Tle	7.97	4.59		1.01				
Pro		4.19	1.97, 1.76	1.96, 1.73	3.76, 3.63			
NMePh							3.15	7.3-7.5

NOE

*Cis* isomer, 17%

	αNH	αCH	βCH <sub>2</sub>	γCH <sub>2</sub>	δCH <sub>2</sub>	Arom	NCH <sub>3</sub>	NPh
Cinn		6.58	under DMSO			7.19, 7.55		
Tle	7.91	4.38		0.72				
Pro		4.27	2.10, 1.97	1.90, 1.74	3.51, 3.34			
NMePh							3.17	7.3-7.5

NOE TleCH<sub>α</sub>-ProCH<sub>α</sub>, minor isomer, *cis* Tle-Pro

<sup>13</sup>C NMR (DMSO-*d*<sub>6</sub>, 150 MHz):

δ 16.4, 24.7, 26.0, 26.5, 29.0, 34.9, 35.7, 37.1, 47.9, 55.1, 56.8, 119.7, 119.8, 119.9, 127.1, 127.6,

NMR Characterization of **31**

<sup>1</sup>H NMR (CDCl<sub>3</sub>, 600 MHz)

*Trans* :*cis* ratio based on integration of the Tle NH, Tle CH<sub>2</sub>, Cinn CH<sub>α</sub> and NCH<sub>3</sub> resonances

<i>Trans</i> isomer, 90%										
	αNH	αCH	βCH <sub>2</sub>	γCH <sub>2</sub>	δCH <sub>2</sub>	Arom	NCH <sub>3</sub>	NPh	CH <sub>2</sub>	tBu
Cinn		6.02	2.50 (CH <sub>3</sub> )			7.58, 7.47				
Tle	6.39	4.78		1.12						
Pro		4.38	1.85	2.04, 1.75	3.87, 3.73					
NMePh							3.27	7.3-7.6		
POM										1.22
NOE										

*Cis* isomer, 10%

	αNH	αCH	βCH <sub>2</sub>	γCH <sub>2</sub>	δCH <sub>2</sub>	Arom	NCH <sub>3</sub>	NPh		
Cinn		6.11	2.52 (CH <sub>3</sub> )			7.53, 7.58				
Tle	6.30	4.43		1.12						
Pro		4.36	2.06	2.09, 1.79	3.80, 3.50					
NMePh							3.31	7.3-7.6		
POM										1.22

<sup>13</sup>C NMR (CDCl<sub>3</sub>, 150 MHz) δ:

17.6, 25.2, 26.6, 26.8, 29.6, 36.2, 37.7, 38.7, 48.8, 56.7, 57.7, 82.4, 121.2, 126.4, 126.5, 127.7, 127.9, 128.0, 129.8, 143.4, 145.5, 149.9, 165.9, 169.8, 171.7, 176.6

Resonances from the *cis* isomer were not distinguishable from those of the *trans* .